

REVIEW

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Mechanisms and applications of biochar in environmental remediation, energy, and catalysis

Parul Sharma¹ and Ramandeep Kaur^{2*}

*Correspondence:

Ramandeep Kaur
ramansaini@pau.edu

¹Department of Chemistry,
Oklahoma State University,
Stillwater, OK, USA

²Department of Chemistry, Punjab
Agricultural University, Ludhiana,
Punjab, India

Abstract

Biochar is a carbonaceous material produced from the thermochemical treatment of biomass in the absence or with a limited supply of oxygen. Despite the use of carbonized biomass to improve agricultural soils over centuries, the application and interest in biochar have grown dramatically owing to its diverse applications in the environment, energy, industry, etc. This review focuses on key technologies of biochar production, comparing their advantages, disadvantages, and their effect on biochar properties. Emphasis has been placed on the physical-chemical properties of biochar (pore structure, surface chemistry, element composition, specific surface area, etc.) as well as the production parameters that control them. Approaches for modifying and activating biochar to improve performance in targeted applications are also discussed. The review further examines the growing role of biochar in pollutant removal, renewable energy systems, and catalytic transformations. In addition, recent advances in nanobiochar are highlighted, emphasizing how reducing particle dimensions to the nanoscale alters surface reactivity, adsorption behavior, and functional performance. Nanomaterials, generally defined as materials with dimensions below 100 nm, possess exceptionally high surface-to-volume ratios that can result in unique physicochemical properties not observed in their bulk counterparts. Finally, current challenges, emerging opportunities, and future pathways for the development and application of biochar-based materials are critically evaluated.

1 Introduction

Biochar is a carbonaceous substance formed through the pyrolysis of biomass (lack of oxygen) or its anaerobic combustion. Biochar usage in improving soil fertility has a long history, dating back thousands of years, when farmers used char in the soil, and was revived recently because of its potential in solving many problems due to its structure and properties [1]. It gained popularity in recent years in solving many problems because of its unique structure and properties [2]. Different properties of biochar depend on the source of feedstock and processes in its production. Pyrolysis temperature is one of the



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variables affecting the properties of biochar significantly. Several studies have proved that surface area and porosity increase with an increase in pyrolysis temperature [3–5].

Syngas and bio-oil are produced as other products of pyrolysis together with biochar; they have great potential as substitutes for conventional fossil fuels, and the utilization of these by-products could increase the energy neutrality of biochar-producing systems [6–10]. Biochemically, the components of biochar can be divided into two kinds, organic and inorganic. For organic matter, the major elements are C, H, and O, and part of N and S also exist; more than 80–90% of the total amount of biochar composition is carbon, which makes the structure stable and ensures its function in carbon sequestration. H and O elements are primarily in the form of surface functional groups, including hydroxyl (-OH) and carboxyl (-COOH); it is mainly because of these groups that the reactivity of biochar and reactions with contaminants can occur. Inorganic matter is minerals and metals, including Fe, Mn, Zn, Ca, Mg, P, Si, etc.; the species depends on the feedstock nature [11].

Apart from the composition, the aforementioned production parameters affect a wide variety of physical and mechanical properties of biochars. These properties include, among others: porosity, specific surface area, particle size, water holding capacity, mechanical stability and grindability, zeta potential and hardness, strength and Young's modulus of elasticity. The above-mentioned properties are tunable and are the reason for the wide application range of biochars in numerous fields. Biochar has been mostly used as a soil additive [12–16] to increase soil productivity, sequester carbon, and mitigate climate change, but it also serves as a highly efficient adsorbent in air purification, water treatment, and the adsorption of heavy metals and organic contaminants [12, 13]. It has been used as a heterogeneous catalyst in biodiesel synthesis [15, 16], and the use as an additive in feed supplements is also under consideration, where it stimulates growth of desirable bacteria and improves feed assimilation. Recent engineering approaches applied to biochar have enabled production of nanobiochar, a nanosize derivative where surface area, reactivity, and functional complexity have been significantly enhanced. Nanobiochar can be either top-down produced (e.g., by ball milling or ultrasonication) or bottom-up produced (e.g., via thermal chemical exfoliation, microwave pyrolysis, hydrothermal carbonization) or by combined methods. Enrichment with function via physical/chemical activation and doping with heteroatoms could extend the applicability of this material for advanced applications such as photocatalysis, supercapacitors, and energy storage [17, 18]. (Fig. 1)

Biochar and nanobiochar represent promising technologies, but scientific and technical barriers that are essential to overcome prior to safe and broad implementation of biochar and nanobiochar exist. Factors such as impacts on plant, animal, and human health, and potential for mobilizing and transporting contaminants in aquatic ecosystems require greater scientific understanding. Higher yield per production cycle and assessment of whether optimizing production processes or large-scale production of these products could influence their properties and performance need to be addressed. Long-term experimental studies that examine the chemical stability, structural integrity, and fate, as well as overall life cycle analysis of biochar-based products, are critical. Within the scope of this review, there is a current overview of research related to biochar and nanobiochar, on production processes, physicochemical properties and modification approaches, an overview of existing applications, a summary of what currently restricts



Fig. 1 Biochar formation

the usage, and emerging prospects for a better utilization of biochar-based products (Fig. 2).

2 Biochar production

The history of the application and production of biochar is very old. There is archaeological evidence showing that biochar was applied to soils over 2000 years ago [19]. It was produced on purpose by ancient people to improve soil fertility and agricultural productivity.¹⁹ The amendment of soil with biochar helped to increase nutrient availability of soils, improved moisture storage capacity, and enriched the soils' capacity for nutrient holding, which resulted in an improvement of crop yield [20]. Besides its agricultural

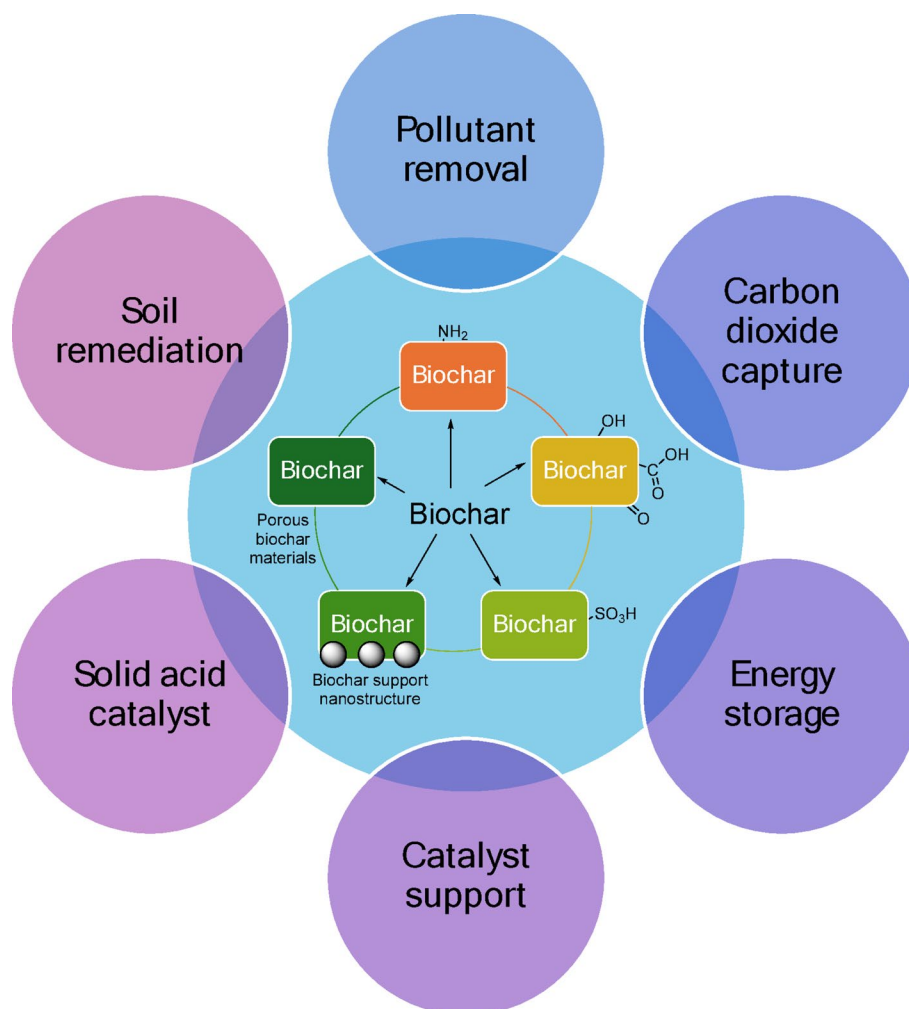
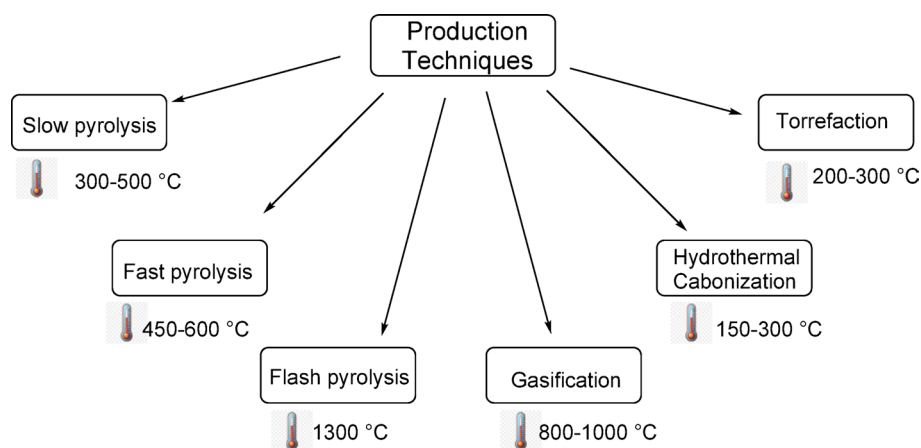


Fig. 2 Biochar for the Synthesis of Various Functional Materials and Their Potential Applications

use, the carbonization process can create side-products that have many uses. It was used for mummification, to make paints and pigments, as binders for building materials, and for arms [19, 21, 22]. In terms of traditional production of biochar, the method was dependent on location, the type of biomass, and the type of kilns. In Spain, underground kilns were used to produce nutrient-rich biochar for agriculture using plant biomass [23]. Indonesia and Sweden used different kinds of kilns, respectively, vertical kilns and a platform-type kiln, adapted to other types of feedstocks and production purposes [24–26]. In common, all types of traditional kilns were built mostly out of clay, and the yield was 20–30% of the initial feedstock [21]. These facts clearly show the awareness about biochar applications and versatility. Scheme 1 summarizes different types of traditional production of biochar.

Due to the growing need to improve the efficiency of biochar production and solve the environmental issues it causes, such as deforestation and air pollution, alternative types of kilns and kiln materials have been designed and fabricated [21, 27]. Metal kilns have been found to be popular for a number of reasons, namely greater durability, efficiency in heat transfer, and greater emission control compared to that achieved in traditional



Scheme 1 Scheme of biochar production methods

kilns made from clay [28]. As a result of their success, metal kilns are still used in many countries for both cooking and small-scale biochar production [19].

Retort kilns were introduced as another technological improvement; these types of kilns recirculate and combust the volatile matter produced during pyrolysis [29]. The heat from the combustion of volatile matter can be recirculated to complete the carbonization of biomass, increasing energy efficiency, reducing emissions, and improving biochar yield [30]. Due to the demand for biochar from various sectors such as agriculture, environment, and industry, a range of new production technologies have been developed. The production technology chosen largely depends on the desired biochar characteristics and the biomass type available; thus, production conditions play a significant role in defining both yield and physicochemical properties of biochar and so their application.

2.1 Pyrolysis

Pyrolysis is one of the oldest and most commonly used methods for biochar production in both traditional and modern contexts. The pyrolysis process relies on the thermal decomposition of organic feedstock in the absence or in a limited presence of oxygen at temperatures typically ranging between 300 and 900 °C [31, 32]. Three distinct products arise from this process, namely: solid biochar, liquid bio-oil, and gaseous syngas. The gases generated consist of carbon monoxide (CO), carbon dioxide (CO₂), hydrogen (H₂), and lighter hydrocarbons. There is a complex interdependence of yields of the three products with regard to operating conditions, though usually it is found that higher temperatures result in greater gas yield, and consequently a lower biochar yield [33–36].

Many operational variables like reaction temperature, heating rate, and residence time play a significant role in the pyrolysis process. By varying operational variables, various types of pyrolysis processes can be operated (Table 1). Catalytic pyrolysis is unlike conventional pyrolysis as the catalysts are used to produce pyrolysis products at a lower temperature, which saves energy consumption [58]. Also, catalysts can produce products with good selectivity and quality, which also depend on various properties such as surface area, pore size, pore volume, and acidity [59]. The thermal pyrolysis process can be a slow or fast type. In slow pyrolysis, a low temperature (150–400 °C) and heating rate are adopted, which allows volatile products to stay inside the reactor for a sufficiently

Table 1 Physical properties of biochar

Property	Influencing Factors	Key Observations	References
Surface Area	Pyrolysis temperature, volatile content, feedstock composition, two-step pyrolysis, particle size (nano vs. macro), ball milling	Increases with temperature due to gas evolution; excessive > 900 °C causes pore collapse. Two-step pyrolysis enhances surface area. Cellulose-rich feedstocks produce a larger surface area. Nanobiochar is often higher than macrobiochar; milling may reduce it.	[37–42]
Water-Holding Capacity	Porosity, pore-size distribution, pyrolysis temperature, residual functional groups	Highly porous biochars retain more water. Low-temp biochars may absorb less due to tar blockage and hydrophobic residues. High-temp biochars have developed pore networks for better water retention.	[43–45]
Density	Pyrolysis temperature, biomass density, particle size/shape, heating rate, atmosphere	Decreases due to mass loss. Linear correlation between biomass and biochar density (coefficient ~0.82 under N ₂ at 15 °C h ⁻¹ up to 900 °C).	[46]
Mechanical Stability	Internal porosity, carbonization rate, and feedstock composition	Slow carbonization reduces cracks, improving strength. Higher porosity decreases stability. Cellulose- and hemicellulose-rich biochars are more fragile, aiding nanobiochar production.	[20, 46, 47]
Zeta Potential	Surface functional groups, particle size, feedstock origin, solution pH	Nanobiochars have more negative potentials than macro-biochars → better dispersion. Woody/agricultural residues are more negative than sludge/manure. pH affects surface charge; significant changes < 4.5 or > 9.5.	[48–50]
Chemical Composition	H/C & O/C ratios, pyrolysis temperature, particle size, feedstock, trace elements	H/C indicates aromaticity; lower H/C → more condensed aromatics. O/C indicates polarity & oxygen functionality; decreases at higher temperatures. Nanobiochar may alter H/C & O/C depending on feedstock. Trace elements: P, Ca, Mg, Na, K, Mn, Fe, Si, Al (Si, K, Ca most abundant).	[39, 49–51]
pH	Pyrolysis temperature, residence time, particle size, feedstock, metal ions	Raw biomass is acidic to mildly basic. Pyrolysis increases alkalinity; the largest increase is in the first 10 min. Nanobiochar may lower pH by 0.6–0.9 units, feedstock-dependent. Metal ions (Mn, Fe) stabilize pH.	[37, 49–57]

long duration and react secondary [60, 61]. The specific surface area (SSA) of biochar produced from slow pyrolysis is usually high, and the SSA is increased as the temperature increases up to about 600 °C [62]. In the case of slow pyrolysis, temperature had a greater effect on biochar properties compared with residence time (Fig. 3).

By contrast, high-temperature fast pyrolysis is normally operated with 450–600 °C at a high heating rate [63]. The rate of breakdown of biomass under these conditions will greatly increase, thus resulting in decreased yield of biochar and increased yield of volatile matter [64]. With prolonged residence time, more volatile matter is released, thus resulting in even lower solid yield [65]. Traditionally, the reactors used for pyrolysis are kilns, retorts, and converters. The kilns are mainly used to carbonize woody biomass into charcoal, and the liquid and gaseous products are recovered in retorts and converters. Slow pyrolysis was commonly carried out in kilns and retorts, while fast pyrolysis must be carried out in a more complex reactor, such as converters [66, 67]. Although

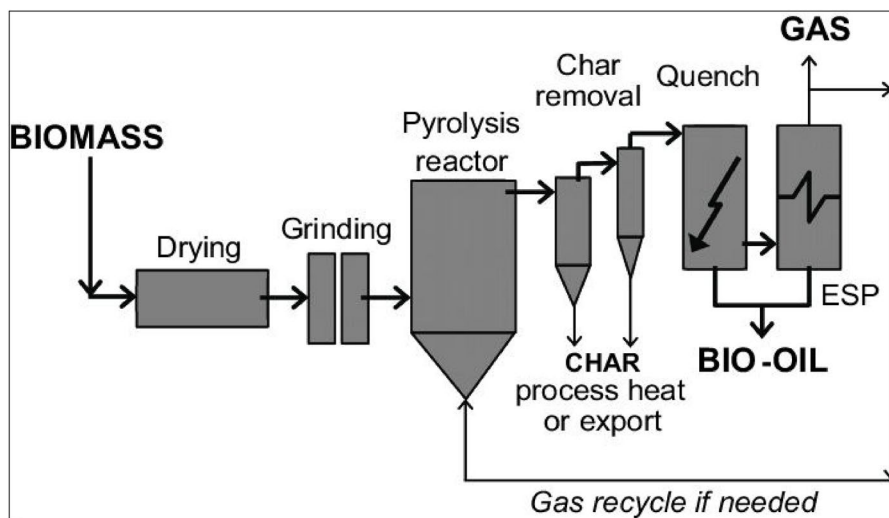


Fig. 3 Biochar production methods by pyrolysis

kilns have low efficiency and cause environmental pollution [68], they have been widely used as economical charcoal producers in developing countries [69, 70]. (Fig. 4)

Retort kilns were originally an enclosed system that enhanced process efficiency by re-burning gaseous and liquid by-products in an efficient way to generate the process heat [71, 72]. However, the performance of the retort kilns demands a constant and rich feedstock material. Converters could process small feedstock particles, enhancing gas permeation efficiency. It allowed continuous fast pyrolysis at higher reaction rates [71]. Current designs of fast pyrolysis reactors could be summarized in rotating cone reactor, ablative reactor, conical spouted bed reactor, bubbling fluidized bed reactor, and circulating fluidized bed reactor [73, 74]. These reactors were designed to produce more bio-oil by providing quick heating rates, a consistent temperature distribution, and a very short vapor residence time [75, 76].

Flash pyrolysis is an intensified form of fast pyrolysis that operates at heat transfer rates $> 1000\text{ }^{\circ}\text{C s}^{-1}$ and temperature $\sim 1300\text{ }^{\circ}\text{C}$ [77]. At these severe conditions, the amount of biochar formed is reduced to $\sim 17\text{ wt\%}$, but the production of bio-oil can be increased to $65\text{--}70\text{ wt\%}$ [78]. Fine milled biomass and very short contact times, and specially adapted reactors (e.g., fluidized-bed reactors, twin-screw mixers) are needed for flash pyrolysis. The continuous breakdown of the particles and the high particle-to-particle contact achieved in a twin-screw reactor enables effective heat transfer [79]. However, flash pyrolysis is not economically viable for large-scale industry [19].

2.2 Gasification

Gasification is a thermochemical process in which biomass is partially oxidized, under the influence of a gasification agent (air, CO_2 , oxygen, or its mixture), to produce gaseous products [80]. The process conditions, such as temperature and air-to-oxygen ratio, play an important role in changing gas composition as well as characteristics of biochar [81, 82]. Since it focuses on gas products rather than solid products, biochar yield of gasification is typically between 5 and 10%, which is even lower compared to fast and flash pyrolysis [83].

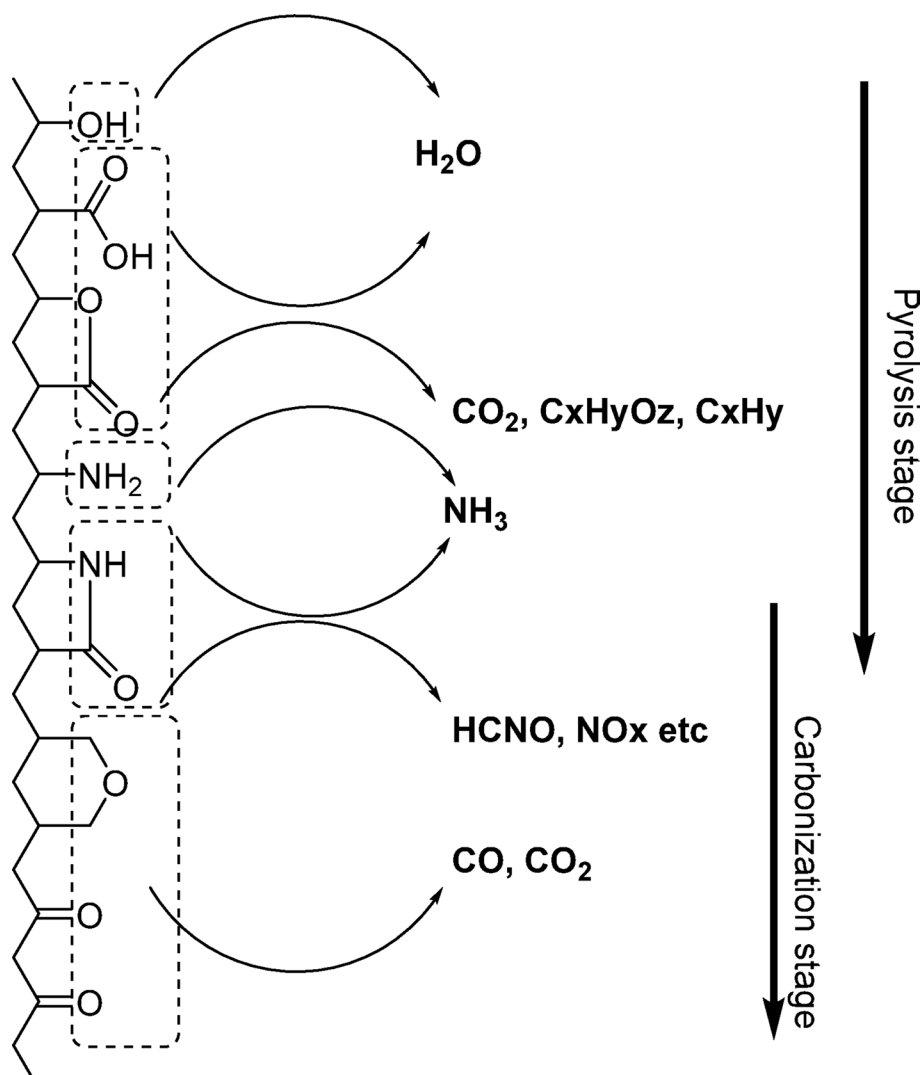
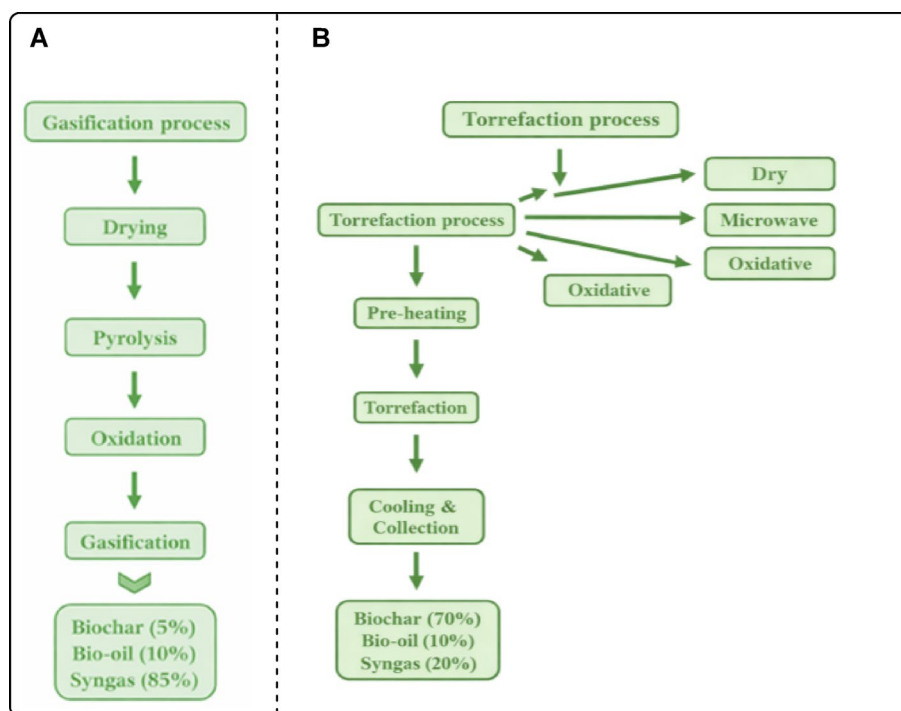


Fig. 4 Decomposition of Biomass with the increase in Pyrolysis Temperature

This process proceeds through four overlapping stages: drying, pyrolysis, oxidation, and gasification. Drying removes moisture from biomass, consuming energy that cannot be recovered. Biomass moisture content below 20% is generally considered suitable for efficient gasification [84]. Pyrolysis usually happens at temperatures around 150–400 °C, followed by oxidation reactions that produce CO_2 [85]. During gasification, char is further converted into gaseous products. (Scheme 2A)

Gasification reactors are classified based on the interaction between biomass and gasifying agents:

1. **Fixed-bed reactors**, commonly used for small-scale power generation due to their high efficiency [86].
2. **Moving-bed reactors**, in which biomass flows continuously through the reactor [87].
3. **Fluidized bed reactors**, in which the biomass particles are entrained in a fluid [88].
4. **Entrained flow reactors**, characterized by very high gas velocities and rapid reaction kinetics [89].



Scheme 2 A) Gasification Steps B) Torrefaction Steps

2.3 Hydrothermal carbonization

Hydrothermal carbonization (HTC) is a thermochemical process to convert biomass into a coal-like carbon-rich solid through sequential dehydration and decarboxylation reactions [90]. HTC possesses many benefits, in particular the possibility of treating wet biomass such as agricultural waste, municipal solid waste (MSW), sewage sludge, etc. Without requiring an energy-consuming drying step before treatment [91, 92]. HTC is performed at relatively moderate temperatures between 150 and 300 °C, which consequently demands less energy compared to conventional thermochemical conversion technologies [93]. Besides producing hydrochar, other liquid and gas fractions can also be recovered and used as additional energy sources, making the whole process more sustainable [93].

HTC systems are generally categorized into batch and continuous reactor configurations, with batch reactors being more widely employed because of their simpler operation and easier pressure management [92]. The most basic HTC reactor consists of a sealed cylindrical metal vessel heated within a furnace, where the biomass is contained in a perforated holder that permits the release and circulation of volatile compounds generated during the reaction [94]. More advanced systems include autoclave reactors, which offer improved control over operating pressure and reaction conditions [95]. Among the available designs, stirred HTC reactors are considered the most sophisticated, as they incorporate magnetic agitation and precise monitoring of temperature and pressure through integrated thermocouples, ensuring enhanced process control and product consistency [96].

2.4 Torrefaction

Torrefaction is a type of moderate thermochemical treatment which has frequently been applied as a biomass pretreatment. Torrefaction takes place under relatively mild temperature and low heating rate in an inert or oxygen-limited atmosphere [97, 98]. Varying the atmosphere, humidity, and method of heating has led to several types of torrefaction, such as dry, wet, oxidative, steam, and microwave torrefaction. Dry torrefaction (DT) is the earliest type of the torrefaction process, which mainly produces solid biochar (70 wt% of original biomass) along with relatively small amounts of bio-oil and non-condensable gases [99]. The properties of DT biochar are good calorific value, hydrophobicity, and grindability [97, 100]. The process usually takes place at temperature ranges of 200–300 °C at atmospheric pressure under an inert atmosphere [101]. In contrast, dry torrefaction requires dry feedstock to be input into the reactor, which will require more energy during pretreatment and make the reactor complex and expensive [102, 103]. (Scheme 2B)

Wet torrefaction (WT), also known as hydrothermal torrefaction, addresses this limitation by processing wet biomass in boiling compressed H₂O at 180–260 °C [104–107]. Hydrochar constitutes the main product (~88 wt%), accompanied by gaseous by-products from decarboxylation reactions [103, 108]. WT reactors include batch systems for laboratory-scale studies [109] and continuous reactors for industrial applications. Oxidative torrefaction introduces controlled amounts of oxygen or partially oxidative gases, combining thermal decomposition with oxidation [110]. Although economically advantageous, this method often reduces solid and energy yields. However, improved performance has been reported for woody biomass, including increased calorific value and solid conversion [111, 112].

Microwave torrefaction (MT) uses electromagnetic radiation to achieve selective, volumetric heating, improving energy efficiency and reducing processing time [113, 114]. Microwave heating activates polar molecules, facilitates removal of bound H₂O, and enhances the lignin and hemicellulose decomposition through localized hot spots [113]. Unlike conventional heating, MT transfers heat from the interior outward [115], resulting in more uniform heating and reduced energy losses [116–118].

2.5 Nanobiochar production

Nanobiochar (NB) is biochar particles with a diameter less than 100 nm [119]. Nanobiochar is usually prepared using either bottom-up or top-down routes. Bottom-up approaches, like chemical precipitation and microwave pyrolysis, prepare nanobiochar through molecular or atomic self-assembly, while top-down routes, which are mostly adopted for preparing nanobiochar, involve physical breakdown of macro-biochar into nanoparticles via arc discharge, laser ablation, and mechanical milling [120]. Of these techniques, the mechanical approaches, especially ball milling, are attractive for their relatively lower energy consumption and ease of operation [121]. In ball milling, biochar is subjected to multiple impacts against metallic balls, which cleaves chemical bonds and enables charge transfer to yield nanoparticles. The size of the nanoparticles could be tuned via modification of milling time, milling speed, and ball-to-particle ratio [122–124].

The properties of nanobiochar are also greatly dependent upon the type of feedstock used, with the majority of applications utilizing agricultural waste, wood, and forest

residue [125]. High content of hemicellulose present in agricultural waste is beneficial for ease of grinding [126, 127]. Properties of nanobiochar can also be tailored by performing certain modification stages before and/or after its production by using acidic solutions [128, 129], iron compounds [130], magnetite [131, 132], hematite [130] or by adding a polymer like styrene-butadiene rubber [133]. Modified characteristics can either be intensified or novel properties are induced by the modification process based on the modification of chemical species and surface groups. Ball milling is currently the most predominantly used method in order to produce modified nanobiochar, and each stage can be performed at different points in the production process [134].

3 Types of biomass and their influence on biochar properties

The nature of the feedstock is crucial in dictating the physicochemical characteristics, in particular the surface area and porosity of biochar. This is due to the differing amounts and types of mineral and organic fractions that different types of biomass comprise. Feedstock types fall into four categories: (1) woody biomass (e.g., tree residues, sawdust), (2) herbaceous/agricultural biomass (e.g., straw, husk, shells, seeds), (3) waste biomass (e.g., Manure, sewage sludge), and (4) algal biomass (e.g., Microalgae and macroalgae). The inherent minerals within the biomass can affect the pores that form during pyrolysis. The presence of some metals may have pore-activating effects [135], but an overwhelming abundance of mineral material may result in a high ash content, which can clog micropores and lower the surface area. Lignocellulosic biochars thus typically have larger surface areas than biochars produced from manure or waste, the former containing larger amounts of minerals and ash-forming materials [135]. Regarding ash content, manure- and waste-derived biomasses generally have the highest content, followed by algal biomass, herbaceous and agricultural residues, and woody biomass last [136, 137].

It has also been shown that certain metallic elements play an activating role. Zhang et al. (2017) [138] indicated that biochar produced from lotus stems and leaves has a high specific surface area owing to high contents of alkali metals, especially potassium(K). Alkali metals and alkaline-earth metals could serve as auto-activating agents during pyrolysis and enhance pore development [139]. In another study, the porosity of biochar prepared from red mud and lignin was increased by increasing the content of iron(Fe) in the feedstock, which implies that a number of metallic species are likely to be active agents responsible for the pore formation.

Besides the chemical structure of biomass material, the original morphology of the biomass is also considered as an influencing factor for pore characteristics [140]. Compared with sewage sludge biochar, lignin-derived biochar (a hollow and tubular structure) will tend to contain more micropores. Further to the above-mentioned morphology features, particle size also affected the characteristics of surface area and pore volumes. Liu et al. (2018) [141] showed that small particle sizes resulted in increased surface area and micropore volume. But Mui et al. (2010) [142] observed that surface area decreased when particle size reduced to a certain point. Small particles allow more homogeneous heating and carbonization and can increase the formation of pores, but it might lead to more homogeneous ash content, which blocks access to pores. So it's difficult to define a universal relation between biomass features and biochar surface properties due to the complicated influence of feedstock characteristics and treatment conditions.

4 Properties of biochar

A variety of properties are found within biochar that are heavily dependent on the feedstock used for its creation and also the conditions that this material was exposed to. The nature of the source material and also the parameters under which it was pyrolyzed result in differing characteristics, including particle size, physical structure, and function. Nano biochar shares much of the functionality with macro biochar but also possesses differing physical and chemical properties, which are outlined below.

4.1 Physical properties

4.1.1 Surface area

Surface area of biochar is important and greatly affected by the evolution of volatile compounds during pyrolysis. A high amount of volatile compounds facilitates the formation of pores and leads to a higher surface area. As a result, at high temperature, the surface area of biochar usually increases with the temperature, as indicated by gas evolution [37]. Whereas the surface area would be reduced at excessively high temperature ($>900\text{ }^{\circ}\text{C}$) due to shrinkage of the matrix and collapse of the pores [38]. At a low pyrolysis temperature ($<500\text{ }^{\circ}\text{C}$), less carbonization resulted in lower surface area. With the increasing temperature up to around $500\text{ }^{\circ}\text{C}$, there was an increasing amount of volatile evolution and a porous carbon matrix formed. The surface area increased with the increasing temperature up to a maximum and then, above $\sim 800\text{ }^{\circ}\text{C}$, a decrease in total pore volume and pore closure caused by pore sintering and graphitization can be detected [39].

It has also been demonstrated that using two-step pyrolysis methods (secondary higher temperature treatment of initial carbonization) yields even greater surface area increases [38]. Feedstock composition is also key; it has been demonstrated that cellulose-rich biomasses produce a higher surface area biochar than hemicellulose or lignin-rich feedstock at equal temperatures, as the smaller thermal stability of cellulose will result in increased volatile release at higher temperature [41]. Nanobiochar does show increased surface area compared to macrobiochar, but not in all cases [39]; it can even result in a decrease due to pore closure and collapse upon milling [40]. For example, rice straw nanobiochar, synthesized at $400\text{ }^{\circ}\text{C}$, showed reduced surface area compared to its macrobiochar counterpart, while at $700\text{ }^{\circ}\text{C}$ it showed a substantial increase in surface area [41]. Ball milling primarily affects the external surface rather than the internal pore structure. Lyu et al. (2019) [42] reported only marginal increases in micropore and nanopore surface areas following milling, suggesting that size reduction mainly promotes particle fragmentation and dispersion rather than altering pre-existing internal porosity.

4.1.2 Water retention characteristics

Water retention is dependent on intra-pore structure, i.e., Pore volume and pore size distribution (rather than just particle size) of the biochar, not simply particle size [43]. Large volumes of water can be contained in highly porous networks while simultaneously increasing aeration of the soil. Pyrolysis temperature had the greatest effect on water-retention compared with other production parameters [44]. Low-temperature pyrolyzed biochars have high proportions of oxygenated functional groups, allowing them to interact with water via hydrogen bonding. However, these materials frequently exhibit lower water-storage capacity because residual tars may obstruct pore channels

and limit accessible surface area [45]. Furthermore, the presence of unconverted aliphatic compounds can impart hydrophobic characteristics. In contrast, high-temperature pyrolysis promotes the formation of extensive pore networks and larger surface areas. Although the concentration of polar functional groups decreases, the enhanced pore structure improves the physical retention of water, often resulting in superior moisture-holding performance [44].

4.1.3 Density

During the thermal conversion of biomass, some volatile elements are released, and the overall mass decreases, which in turn causes the density of the material to decrease. The strong relationship between feedstock and biochar densities was previously confirmed by researchers [46]. The final density is subject to parameters such as feedstock shape, particle size, heating rate, reaction temperature and pressure, as well as the atmospheric environment. In particular, under an atmosphere of nitrogen and a slow heating rate up to 900 C, the density coefficient is about 0.82 [46].

4.1.4 Mechanical properties

The structural durability of biochar is controlled by both production conditions and the degree of pore development within the carbon matrix. Slower carbonization processes generally reduce crack formation and produce materials with greater mechanical integrity [46, 47]. However, extensive pore formation often compromises structural strength because highly porous materials are more susceptible to fragmentation [47]. Feedstocks rich in cellulose and hemicellulose tend to generate biochars with lower mechanical resistance. While this may limit some engineering applications, it is advantageous for nanobiochar production because weaker structures can be milled more efficiently into nanoscale particles [20].

4.1.5 Surface charge and zeta potential

Zeta potential is often applied as an indicator of surface charge and suspension stability for biochar particles. Nearly all biochar nanoparticles exhibit negative surface charges due to the dissociation of hydroxyl-containing functional groups. Absolute zeta potential of nano-biochar tends to be higher in magnitude relative to conventional biochars, thus resulting in enhanced colloidal stability and reduced aggregation [48]. Enhanced stability is especially significant when considering particle motion and pollutant transport in the aqueous phase [49]. The magnitude of the zeta potential depends on the nature of the feedstock. Biochar made from woody biomass and agricultural residues usually exhibits greater negative surface charge than that made from sewage sludge and manure. This can mainly be explained by the fact that woody materials and agricultural residues contain more lignin-derived phenolics and carboxylic groups than sewage sludge and manure [49, 50]. The solution pH has a significant impact on the charge of the surface as well. In acidic conditions, the protonation of functional groups can reduce the magnitude of negative charge, lowering the colloidal stability. A dramatic alteration is typically seen below 4.5 and above 9.5, where there is acid/base behavior of carboxyl and phenolic groups on the biochar surface [49].

4.2 Chemical characteristics

4.2.1 Elemental composition

The elemental analysis results for biochar are often discussed using H/C and O/C ratios. These are indicative of structure and can provide information about specific properties of biochar. The H/C ratio is generally an indicator of aromaticity. Lower H/C values typically indicate more aromatic condensation and carbonization. The O/C ratio represents the surface polarity and is related to the number of oxygenated groups present on the biochar surface, which affects polar contaminant interactions. Materials with H/C ratios > 1 still possess a high amount of aliphatic nature from original cellulosic, hemicellulosic, and lignin structures. These elemental ratios are tunable by altering conditions, and so an application may benefit from a biochar that has been designed to have a particular structure. The O/C ratio generally decreases with pyrolysis temperature, as oxygenated groups are continually removed [39]. However, particle size can also play a role in the elemental analysis results. Several studies have demonstrated that the elemental analysis of biochar conversion to nano-biochar may increase or decrease both the H/C and O/C ratio depending on the feedstock and milling process applied [49]. Beyond the major constituents carbon, hydrogen, and oxygen, biochars contain a variety of inorganic elements, typically in trace concentrations. Commonly detected minerals include aluminum, silicon, iron, manganese, potassium, sodium, magnesium, calcium, and phosphorus [51]. Silicon, potassium, and calcium are often the dominant species, although their abundance is highly dependent on the original biomass source. Current investigations comparing macro- and nanoscale biochars have not established a consistent trend in mineral composition following particle-size reduction [51].

4.3 pH characteristics

The biochar pH has the property of influencing its suitability for environmental and agricultural applications. Excessively acidic or alkaline materials may alter soil chemistry and affect nutrient availability. Most untreated biomass feedstocks exhibit neutral to mildly acidic characteristics [52]. During pyrolysis, acidic organic groups are progressively removed, causing the resulting biochar to become increasingly alkaline as processing temperature rises [37]. Residence time also contributes to pH development, with the largest changes typically occurring during the initial stages of thermal treatment [53]. In general, biochars produced through conventional pyrolysis possess higher pH values than hydrochars obtained from hydrothermal carbonization, where organic acid formation is favored [37]. Mechanical downsizing to produce nanobiochar frequently results in a slight decrease in pH. This effect is commonly led to the generation of additional O₂-containing acidic functional groups during milling [50, 54]. Reported decreases generally range from approximately 0.6 to 0.9 pH units [55]. Nevertheless, the response is not universal. Depending on feedstock composition and processing conditions, some studies have observed little change or even an increase in pH following particle-size reduction [49, 51, 57]. Mineral materials present (e.g., Iron and manganese) might even have additional effects on the pH behavior, as they provide positively charged surface sites which buffer pH variations [56]. Based on current evidence, it seems clear that the effect of nanosizing on biochar pH is very system-dependent.

4.4 Influence of biochar properties on environmental behavior

Reducing biochar to the nanoscale introduces unique physicochemical characteristics that can substantially alter its environmental performance. One of the most important consequences of nanosizing is the dramatic increase in surface area relative to particle volume. This enlarged surface provides greater reactive sites, encouraging increased interaction with contaminants, nutrients, and biological systems. As a result, nanobiochar often demonstrates enhanced adsorption, faster reaction rates, and improved catalytic activity compared with conventional biochar materials. Changes in particle size can also modify the electronic and surface properties of biochar. At the nanoscale, variations in surface energy, redox behavior, and electronic structure may influence chemical reactivity and interactions with surrounding environments. These changes can enhance the ability of nanobiochar to participate in oxidation–reduction processes, pollutant transformation, and contaminant immobilization.

The fate of nanobiochar in the environment depends on factors such as small size, surface charge, and surface functional groups. Nanoparticles can move more readily through soil pores and aquatic environments than larger particles, increasing their mobility and potential bioavailability. Their transport behavior is strongly affected by aggregation tendencies, colloidal stability, and interactions with natural organic matter. Surface characteristics also influence transformation processes such as oxidation, dissolution, and photochemical reactions that occur after environmental release [49, 51, 57]. The high reactivity of nanobiochar can provide significant benefits for environmental remediation by improving contaminant removal and accelerating degradation processes. This same property can also result in environmental problems. Enhanced surface activity can promote the formation of ROS species, which can result in microorganisms, plants, and aquatic life experiencing oxidative stress [49, 51, 57]. Consequently, evaluating the environmental implications of nanobiochar requires consideration of its entire life cycle, including production, application, transformation, and eventual disposal. Optimization of particle size, morphology, and surface functionality is essential to maximize environmental benefits while minimizing potential ecological and health risks.

5 Activation methods

As synthesized biochars obtained directly through thermochemical treatment usually has low adsorption and catalytic capacity due to partially blocking of pore networks by remaining tars, ash and other carbons, various activation methods (such as steam activation, CO₂ activation, chemical activation and so on) are developed to alter the pore structure, increase specific surface area and add active surface groups, which facilitates the accessibility of the active sites and the interaction between biochar and targets. Activation strategies generally aim to increase chemisorption capacity by creating chemically active surface functionalities capable of forming stronger interactions with pollutants and other adsorbates. Depending on the desired application, activation can be accomplished through thermal treatment, chemical modification, biological approaches, or surface functionalization methods (Table 2) [163–165].

5.1 Physical activation

Physical activation or thermal activation is the process where biochar is treated with oxidizing gases such as steam, CO₂, and air at high temperatures. Physical activation helps

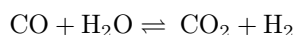
Table 2 Summary of biochar activation methods and their effects on surface area and pore characteristics

Activation Strategy	Processing Conditions	BET Surface Area (m ² g ⁻¹)	Pore Volume (mL g ⁻¹)	Main Outcome	Ref.
Aluminum chloride-assisted activation	AlCl ₃ impregnation for 6 h, dried at 80 °C for 48 h	212.58–418.14	0.056–0.077	Promoted pore development and improved incorporation of metal species	[143]
Phosphoric acid treatment	5% H ₃ PO ₄ , 70–80 °C, 2 h	199–557	0.026–0.22	Increased porosity and expanded surface area	[144]
Alkali sulfide or potassium hydroxide activation	Mixing with Na ₂ S or KOH solution	32.85–59.23	–	Generated additional surface sites and moderate porosity enhancement	[145]
Steam-assisted physical activation	Steam treatment at 900 °C for 1 h	702–2079	0.53–1.212	Produced extensive micro- and mesoporous networks	[146]
Zinc chloride activation	ZnCl ₂ impregnation followed by carbonization at 800 °C	583–1067	0.001–0.511	Favored formation of a predominantly microporous structure	[147]
High-temperature pyrolysis under inert atmosphere	900 °C under N ₂ for 4 h	204–1080	0.118–0.95	Improved pore characteristics and adsorption capability	[148]
Sodium hydroxide activation	NaOH treatment 2 h at 800 °C	583–1470	0.01–0.705	Enhanced porosity and contaminant uptake performance	[149]
Carbon dioxide activation	CO ₂ exposure 11 h at 750 °C	429–621	–	Developed a more homogeneous pore structure	[150]
Microwave-assisted activation	Heating at 900 °C for 75 min	702–2079	–	Increased micropore content and catalytic potential	[151]
Sulfuric acid modification	H ₂ SO ₄ treatment at 180–380 °C	5.3–29.6	0.18–0.30	Introduced acidic surface groups and improved pollutant removal	[152]
Sodium sulfide-assisted activation	Na ₂ S treatment with drying and mixing at room temperature	32.85–59.23	–	Increased availability of adsorption-active sites	[153]
Steam activation of lignocellulosic residues	Lignin and pruning waste treated at 850 °C	204–1080	–	Enhanced catalytic behavior through increased porosity	[154]
Mixed-gas activation	CO ₂ /N ₂ treatment at 750–920 °C for 0.5 h	435–687	0.18–0.30	Improved pore distribution and structural uniformity	[155]
Moderate-temperature thermal activation	Heating at 500 °C under N ₂ for 4 h	13–16	–	Improved surface characteristics for adsorption applications	[65]
Dilute phosphoric acid activation	Low-concentration H ₃ PO ₄ treatment at 70–80 °C	199–557	0.026–0.22	Increased acidic functionalities beneficial for pollutant capture	[156]
Tungsten precursor impregnation	Ammonium tungstate treatment followed by 1000 °C calcination	> 1500	–	Generated thermally stable tungsten carbide phases	[157]
Iron nitrate modification	Fe(NO ₃) ₃ impregnation and thermal treatment at 800 °C	496–716	0.08–0.27	Improved catalytic activity through well-dispersed Fe species	[158]
Oxidative thermal activation	Air treatment	450–800	0.2–0.5	Enhanced pore accessibility for gas adsorption	[159]
Plasma-assisted surface activation	Plasma exposure for 20 min	600–1000	–	Introduced reactive functional groups onto the carbon surface	[160]
Potassium hydroxide activation	KOH treatment at 700 °C	1200–1800	–	Produced highly porous carbons suitable for energy-storage devices	[161]
Combined chemical-physical activation	Sequential H ₃ PO ₄ and CO ₂ treatment	800–1400	0.3–0.6	Increased active-site density and overall porosity	[162]

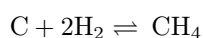
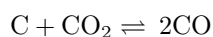
in removing carbon deposits and opening the pore channels. It leads to an increase in pore availability and surface area [166]. Although the process is traditionally described as “physical” activation, the underlying mechanism involves chemical reactions between carbon and activating gases. For example, steam becomes highly reactive at temperatures above approximately 750 °C and participates directly in carbon gasification reactions rather than merely removing volatile residues [167].

Physical activation success is commonly quantified by burn-off percentage (weight lost during treatment). Moderate burn-off percentage levels favor pore development and enhance adsorption efficiency; too much carbon loss leads to the structural breakdown of the pore framework and causes structural collapse. Therefore, activation parameters need to be properly controlled in the case of both developing porosity and maintaining the structure. Steam activation is a broadly adopted physical activation method and could be conducted directly after pyrolysis. Under temperatures typically ranging from 200 to 800 °C with a heating rate of 5–10 °C min and a treatment time of 30–60 min, a highly porous structure containing micropores and mesopores may be generated, leading to good adsorption and catalytic performance [16, 112, 168]. The activation process generally involves three major steps [80]:

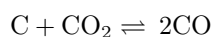
- a) Water molecules interact with the carbon surface, forming oxygen-containing surface complexes and carbon monoxide.
- b) These intermediate surface species subsequently participate in gasification reactions, generating carbon dioxide and creating additional pore space.
- c) In parallel with this process, the water-gas shift reactions are also taking place, leading to a more advanced structural modification and development of porosity of the carbon matrix.



The generation of CO₂ and H₂ is critical for surface activation, as these species further react with the carbon matrix through:



The CO activation also follows the same principles and operates within the 200–900 °C range. Both steam and CO are helpful in the formation and growth of pores, and both of them resulted in little or no differences in adsorption capacity and micropore volume of the activated samples [23, 86]. At the same time, the steam-activated biochar is reported to be more reactive with a greater number of oxygen-containing functional groups [21]. However, the reaction kinetics of the CO activation are found to be slow, which allows better control of oxidation, and a more homogeneous pore structure with higher surface area is attained [27, 139]. At temperatures higher than about 700 °C, CO is very reactive through the reverse Boudouard reaction:



While this reaction is essential for pore development, it also reduces overall biochar yield by converting solid carbon into gaseous products. Consequently, optimizing activation conditions requires balancing surface development against excessive carbon loss.

5.2 Chemical activation

Chemical activation is an extensively employed method to enhance the physicochemical properties of biochar. In this technique, the biochar is treated with chemicals that are active and can induce changes in the surface chemistry and pore structures of biochar, through methods such as oxidation, sulfonation, amination, and metal loading. Various activating agents can be applied, including acids, alkalis, and oxidizing agents, depending on the target applications. The selection of the activating agents is especially critical in the adsorption process, where the target functional groups are necessary to achieve desired interactions with the specific contaminant species. High concentrations of the reagents or harsh temperatures may damage the carbon framework structure and affect its performance.

5.2.1 Alkali activation

Among chemical activation methods, alkali treatment using potassium carbonate (K_2CO_3) and potassium hydroxide (KOH) has received considerable attention due to its effectiveness in generating highly porous carbon materials [169]. The activation temperature plays a critical role in determining the resulting pore structure and surface characteristics.

Studies have shown that both K_2CO_3 and KOH increase the surface area of biochar as activation temperature rises; however, their effects on pore development differ significantly. Activation with K_2CO_3 generally produces biochars with greater surface areas and more extensive pore networks, particularly at elevated temperatures. This treatment promotes the formation of both micropores and mesopores, resulting in a well-developed hierarchical pore structure.

In contrast, KOH activation primarily increases the overall surface area but may alter the distribution of pore sizes. Although pore formation is enhanced, a reduction in microporous volume has been reported in some cases. Consequently, the selection between K_2CO_3 and KOH should be guided by the desired pore architecture and the specific application of the activated biochar.

5.2.2 Metal-assisted activation

Incorporation of metal species is an effective approach for increasing the catalytic and adsorption properties of biochar. Metal-functionalized biochars can be prepared either by introducing metal precursors before carbonization or by modifying the biochar after its formation. During one-step synthesis, metal salts are combined directly with the biomass feedstock prior to thermal treatment. Under these conditions, volatile compounds and carbonaceous intermediates generated during pyrolysis can act as reducing agents, while the metal species facilitate carbon transformation and pore development [170, 171]. Alternatively, a two-step procedure involves the impregnation of preformed biochar with metal-containing solutions. This method enables metal ions or nanoparticles to be anchored onto the carbon surface, providing additional active sites and improving material functionality. An instance of this preparation is the synthesis of iron oxide-modified nanobiochar from rice husks [172]. In this process, ball-milled biomass was combined with iron sulfate and urea before pyrolysis under reducing conditions at 600 °C. The resulting Fe_3O_4 -containing nanobiochar exhibited superior adsorption performance

for arsenic removal compared with the unmodified material, demonstrating the benefits of metal incorporation for environmental remediation applications (Fig. 5).

5.2.3 Oxidative activation

The oxidative activation method is widely used to install oxygen-containing functional groups on the biochar surface in order to enhance adsorption performance. Among the oxidizing agents used, hydrogen peroxide (HO) is preferred for its low cost, high availability, and eco-friendly character. Indeed, in case of decomposition, HO leads to only water and oxygen as byproducts and avoids any secondary pollution [173]. The treatment with hydrogen peroxide leads to a functionalization of the biochar surface with oxygen-containing groups (hydroxyl, carbonyl, and carboxyl groups), increasing polarity and thus leading to better adsorption of diverse contaminants. This oxidation agent does not introduce other heteroatoms than the initial carbon backbone, contrary to other oxidation processes [173].

5.2.4 Activation using alkali hydroxides (KOH and NaOH)

Hydroxides of both potassium and sodium are some of the most commonly utilized chemical activating agents for generating ultra-porous biochar [80, 174]. In the thermal activation process, the alkali hydroxides react with the carbon matrix via several redox and gasification reactions, creating new pore structure and significantly enhancing the surface area. The mechanism of activation includes reactions between hydroxides and carbon producing metallic species, gaseous products, and intermediaries that etch the carbon structure. Such interactions are responsible for forming additional micro- and mesopores, thus enhancing access to the intra-surface active sites. Therefore, both KOH- and NaOH-activated biochar are used in large volumes for processes where large surface area materials are required, such as adsorption of pollutants, catalysis, energy storage, etc. [80, 174].

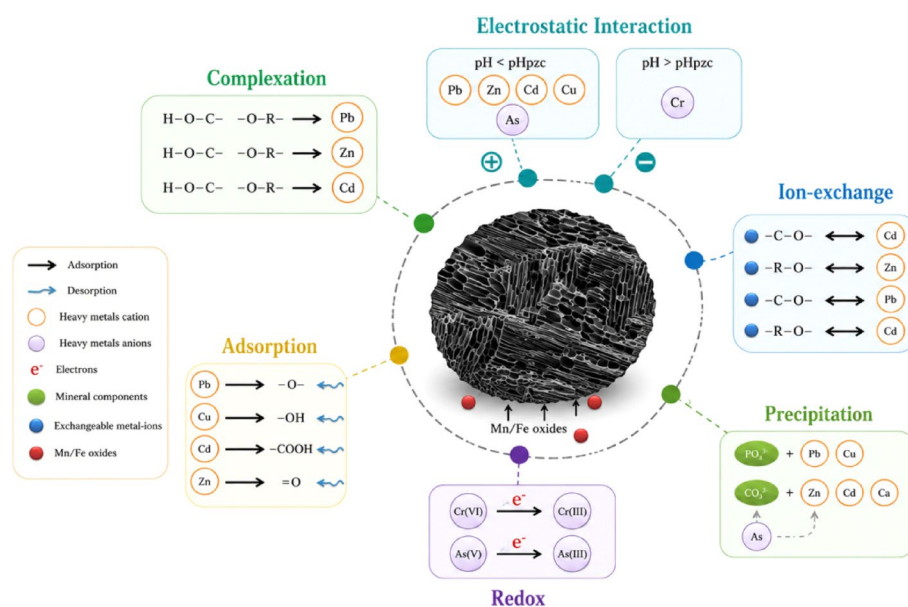


Fig. 5 Biochar interaction mechanisms proposed for organic contaminant removal

Table 3 Biomass-derived biochars: production conditions, properties, and adsorption performance

Feedstock Source	Preparation Method	Pollutant Removed	Key Characteristics and Removal Efficiency
<i>Spirulina</i> microalgae	Pyrolyzed in an inert atmosphere at 750 °C for 2 h	Tetracycline	Produced a carbon material with low oxygen functionality and limited surface area (2.63 m ² g ⁻¹). Despite this, adsorption capacity reached 147.9 mg g ⁻¹ , with approximately 61% desorption efficiency.
Wheat straw	Pyrolysis at 500 °C followed by KMnO ₄ /KOH activation	Tetracycline	Activation produced a very porous material that had a surface area greater than 1500 m ² /g and a pore volume of 0.85 cm ³ /g. This material showed good tetracycline uptake (584.19 mg/g) and good resilience against interfering dissolved ions.
Sunflower seed husk	Carbonization at 700 °C combined with KMnO ₄ , KOH, and ZnCl ₂ activation	Tetracycline	Displayed a well-developed pore structure (1578.3 m ² g ⁻¹) and large pore volume (1.138 cm ³ g ⁻¹). The maximum adsorption capacity reached 673 mg g ⁻¹ while maintaining high performance in the presence of common cations.
<i>Chlorella</i> microalgae	Wet and micro-wave-assisted torrefaction at 160–170 °C	Methylene blue	Generated biochar with modest porosity and a low specific surface area. Nevertheless, dye removal efficiency exceeded 85%, with adsorption capacities up to 113 mg g ⁻¹ across a broad pH range.
Penicillin fermentation residues	Nitrogen-assisted torrefaction followed by KOH impregnation	Methylene blue	Produced a highly porous carbon material with a surface area approaching 1810 m ² g ⁻¹ and a pore volume of 1.02 cm ³ g ⁻¹ . Maximum methylene blue uptake was reported as 620 mg g ⁻¹ .
Bamboo biomass	Pyrolysis at 700 °C with KHCO ₃ and urea treatment	Methylene blue	Resulted in a nitrogen-modified porous carbon possessing a surface area of 1693 m ² g ⁻¹ and substantial pore volume. Adsorption capacity toward methylene blue reached 499 mg g ⁻¹ .
Orange peel	Slow pyrolysis at 400 °C followed by KOH activation	Chromium species [Cr(III)/Cr(VI)]	The activated biochar exhibited a high surface area (998 m ² g ⁻¹) and significant pore development. Cr(VI) adsorption capacity was reported at 285.5 mg g ⁻¹ under acidic conditions.
Rice husk/polyethylene waste blend	Co-pyrolysis at 390 °C under nitrogen	Cr(III)	Produced a material with very limited surface area (< 5 m ² g ⁻¹). Consequently, chromium adsorption capacity was comparatively low at 9.23 mg g ⁻¹ .
<i>Potamogeton crispus</i> algae	Pyrolysis at 300 °C for 2 h	Cr(VI)	Generated biochar with minimal porosity and low surface area. The material relied primarily on surface functional groups rather than pore structure for chromium removal.



The aforementioned reactions create a large increase in the surface area and pore volume. Biochar made from brewer's draff activated by 2 M KOH resulted in an increase in total pore volume from 0.01 mL/g to 8.74 mL/g and an increase in surface area from 9.8 m²/g to 11.6 m²/g [175]. NaOH has consistently proved less effective than KOH for creating surface area and porosity [176].

5.2.5 ZnCl₂ activation

The reason for the high activity of Zinc chloride is due to its dehydrating properties and inhibition effect for tar formation, high diffusion rate into biomass even at low temperature, because ZnCl has a wide range of temperatures in the liquid state. This helps for good impregnation [177, 178]. The biochar derived from rice husk, when activated by ZnCl in a ratio of 3:1 to obtain a specific surface area, was found to be 645 m²/g, while only 28 m²/g of non-activated biochar possessed. Similarly, for safflower seed press cake

and corn stalk biochar, an increased activation temperature and impregnation ratio increased the surface area, pore size, and adsorption capacity [179, 180].

5.3 Thiol functionalization

A common method for introducing thiol functional groups is 3-mercaptopropyltrimethoxysilane (3-MPTS), which reacts with the hydroxyl groups of the surface to obtain stable thiol-modified surfaces while methanol is generated. The common method for the preparation is impregnation and ball milling. The adsorption capacities of thiol-functionalized biochar prepared by ball milling for Hg (320.1 mg/g) and CHHg (104.9 mg/g) were significantly higher than the modified biochar before ball milling [181].

5.4 Nitrogen doping

One of the most common strategies applied to modify surface chemistry and improve catalytic properties of biochar is nitrogen doping. This can be done either by employing nitrogen-containing precursors such as ammonium hydroxide, ammonium nitrate, urea, and ammonia, or by employing a synthesis in a nitrogen-containing atmosphere. The introduction of nitrogen atoms to the carbon framework leads to the creation of more active sites and affects the electronic properties and adsorbents/reactants adsorption. The effectiveness of nitrogen doping in increasing catalytic activity has been proven by a number of research works. For example, Xing et al. [182] have synthesized N-enriched carbon nanoparticles from graphite powder and a nitrogen-containing atmosphere that possess excellent activity for the oxygen reduction reaction. Pyrolysis temperature plays a significant role in the doping efficiency, as overly high temperatures remove oxygenated surface groups, which are preferential adsorption sites for nitrogen-doped precursors, and lead to lower doping efficiency [183].

Compared with higher-temperature-produced biochars, biochars produced at moderate temperatures tend to show better nitrogen incorporation efficiency. For instance, biochars produced at 450 °C after nitrogen modification were found to have a larger surface area than that initially produced at 600 °C [183]. Oxygen-containing functional groups can react readily with ammonia and similar nitrogen sources to form more stable nitrogen functional groups within the carbon network. However, it should be noted that the resulting nitrogen functionalities, including pyridinic, pyrrolic, or graphitic nitrogen, are highly dependent on the synthesis parameters and treatment conditions [182–184]. Recently, research also pointed out that modifying mechanical processing parameters like ball milling can help to optimize the dispersion and preferential formation of nitrogen-containing groups [183, 184].

6 Biochar applications

The high surface area, modifiable pore network, and multitude of surface chemistries the biochar possesses contribute to the diversity in potential applications that exist. The ability to tailor the biochar for specific interactions with pollutants, nutrients, gases, and biological systems has given the material broad appeal for applications as varied as adsorption, catalysis, energy, and environmental management. As production techniques improve and biochar modifications continue to emerge, the material's potential use continues to grow to now include applications for agriculture, wastewater treatment, biomedical, animal production, and advanced materials. Biochar has many potential

applications in various fields. In agriculture, biochar is used as a soil conditioner to enhance carbon retention of nutrients. In biomedicine, targeted biochars are studied as drug carriers and adsorbent platforms. In the animal industry, biochar application as a feed additive to diminish methane emissions is being researched. In the construction sector, biochar addition has been made to improve the mechanical performance, insulate, and extend durability. It is projected that biochar will be further used in various fields, contributing to more sustainable technologies and increased circularity of resources.

6.1 Environmental remediation

The uses of biochar are manifold; however, the area in which it has been one of the most studied is that of remediation. The large specific surface area, porous structure, and a plethora of reactive surface groups present in biochar allow it to adsorb or fix many contaminants from water, soil, and air. In aquatic systems, the effectiveness of biochar in the remediation of organic pollutants and toxic metals has been extensively studied. Agricultural and forestry-derived biochars are highly effective in the removal of drugs and other organic contaminants (like dyes) and show removal efficiencies of above 90% under optimal conditions [185–187]. Adsorption of species such as tetracycline and methylene blue occurs via a mechanism involving pore filling, electrostatic interaction, hydrogen bonding, and pi-pi interaction. Removal of heavy metals from contaminated environments represents another area that is highly researched. Lead, cadmium, copper, and zinc, among others, can be removed or immobilized using biochar. Their removal depends largely on the feedstock composition, the amount of minerals contained in the feedstock, and the surface chemistry of the char. The performance in the remediation of heavy metals can be significantly improved in biochars produced from biosolids or other mineral-rich feedstocks since these chars contain an increased amount of inorganic materials, responsible for mechanisms of ion exchange and precipitation [188].

There are several mechanisms of contaminant removal, including electrostatic attraction, forming complexes with surface oxygen-containing functional groups, ion-exchange reactions, precipitation with carbonates and phosphates, and so forth [189, 190]. These adsorption mechanisms are highly influenced by the preparation process conditions of biochar. Generally, an increase in pyrolysis temperature causes the formation of a porous structure and increases surface area, leading to the improvement of adsorption efficiency toward organic and inorganic pollutants [68, 191–193]. Solution factors such as pH have great effects on contaminant speciation and surface functional group ionization status; thus, divalent metal capture capacity by biochar is improved with increasing pH due to enhancement of electrostatic interaction and surface complexation [188].

Biochar has attracted more interest as a material for air pollution control. Various biochars were employed to capture volatile organic compounds (VOCs), S-containing gases, and NO₂ from the gas phase [194]. Activation treatment can significantly improve the gas adsorption efficiency by improving porous accessibility and surface reactivity, which includes steam activation or alkali activation. In addition, doping metal oxides such as iron- and aluminum-based materials can increase the capture efficiency of CO₂ by increasing active adsorption sites [194]. Recent developments in advanced production methods, including microwave-assisted pyrolysis and engineered surface

modification, have further improved the gas adsorption properties of biochar [195–198]. These approaches can increase both physical and chemical adsorption processes, resulting in significantly enhanced pollutant removal efficiency.

In soil remediation, biochar contributes not only to contaminant immobilization but also to improvements in soil quality. Iron-modified and magnetic biochars have demonstrated excellent performance in the removal of toxic metals while offering the added advantage of magnetic recovery and reuse [199, 200]. In addition, biochar amendments can increase cation-exchange capacity, improve nutrient retention, and reduce bioavailability and mobility of heavy metals, thereby lowering environmental risks and improving agricultural productivity [196]. Overall, the versatility of biochar in soil restoration, air treatment, and water purification highlights its value as a sustainable remediation material [201–205]. High removal efficiencies, broad contaminant applicability, and adaptability through surface modification continue to drive interest in biochar-based technologies for environmental protection and resource recovery [11, 194, 197] (Table 3).

7 Environmental implications of biochar production and use

Biochar has considerable potential in sustainable technologies because of its usefulness in remediation, energy storage, and catalysis. However, its overall environmental value depends not only on its final application but also on how it is produced, modified, transported, and disposed of. Therefore, the benefits of biochar must be evaluated across its full life cycle. Biochar can effectively remove heavy metals and organic contaminants and minimize their discharge to aquatic systems during wastewater treatment [43, 174]. Such removal of pollutants is strongly related to its porous structure, large surface area, and surface functional groups [175]. However, volatile organic matter, particulates, etc. Can be released during the process of biochar preparation if the pyrolysis conditions are not optimal [176, 177]. Therefore, the type of feedstock and the temperature of pyrolysis determine whether the biochar has a net positive environmental effect [176]. Biochar is also being explored as a renewable material for batteries and supercapacitors. Its use can reduce dependence on conventional activated carbons and support circular use of biomass resources [178, 179]. However, activation or chemical modification steps used to improve electrochemical performance may require additional energy and chemicals. As a result, optimization of activation methods is necessary to improve performance without increasing environmental burdens [175, 176].

In catalysis, biochar can function either as a catalyst or as a support material for active catalytic species. Its use may reduce reliance on conventional metal-based catalysts, which can be expensive, toxic, or difficult to dispose of [180, 181]. Biochar surface chemistry can also be adjusted to promote catalytic activity in reactions such as biodiesel production and pollutant degradation [43, 180]. However, emissions such as PAHs and VOCs during production must be carefully monitored to ensure that catalytic applications remain environmentally sustainable [176, 177]. Overall, biochar can contribute to greener remediation, energy, and catalytic technologies. However, its sustainability depends on responsible production, controlled emissions, appropriate feedstock selection, and detailed environmental assessment. Future studies should place greater emphasis on life-cycle analysis, emission monitoring, and long-term environmental behavior to confirm the true sustainability of biochar-based systems.

8 Scalability and practical implementation of biochar technologies

Although biochar has shown strong promise at the laboratory scale, large-scale production remains challenging. Industrial implementation requires consistent product quality, reliable reactor operation, sufficient biomass supply, and economic feasibility. All these are critical issues for such applications as soil amendment, carbon sequestration, water treatment, or energy storage [182]. One major problem is ensuring the homogeneity of biochar properties when scaling up biochar production: properties such as surface area, adsorption capacity, porosity, pH, or chemical stability depend on several parameters related to the pyrolysis, such as the temperature and residence time during the heating phase or the heating rate and the feedstock properties [183, 206–208]. Laboratory systems allow close control of these variables, but industrial reactors often experience temperature gradients and operational fluctuations. These variations can lead to inconsistent biochar quality, reducing performance reliability and market acceptance.

8.1 Pyrolysis control, energy use, and co-product recovery

Slow pyrolysis is commonly used for large-scale biochar production because it favors high fixed-carbon retention. However, controlling temperature and residence time becomes more difficult as reactor size increases. Industrial systems must therefore be designed to process large biomass volumes while maintaining stable thermal conditions [182, 209–212]. Another essential aspect in a commercial-scale operation is energy efficiency. The pyrolysis process gives rise to several co-products, including bio-oil and syngas, which can be utilized in the biochar plant as a source of energy, heat, or electricity. By means of their recovery, the dependency on external energy sources can be reduced and thus operating costs decreased, and a higher overall sustainability of the process increased [182]. A highly integrated system may have biochar plants that move towards partial or total energy self-sufficiency due to the exploitation of co-products.

8.2 Feedstock availability and optimization

The choice of biomass feedstock is central to successful scale-up. While many materials can be tested in laboratory studies, industrial production depends on feedstocks that are affordable, locally available, and obtainable in large quantities. Agricultural residues, forestry wastes, and other lignocellulosic materials are especially attractive because they are abundant, carbon-rich, and generally low in toxicity [183]. However, biomass variability remains a major limitation. Differences in moisture content, ash content, mineral composition, and organic structure can affect pyrolysis efficiency and final biochar properties [182, 183, 213–218]. Pretreatment steps such as drying, grinding, or sorting may improve consistency but can also increase processing costs. Therefore, feedstock optimization must balance product quality with economic practicality.

8.3 Economic feasibility and policy support

The commercial success of biochar depends on both production cost and market demand. Agricultural use, water treatment, carbon sequestration, and replacement of activated carbon are among the most promising markets. However, high capital costs, operational expenses, and competition with established materials can limit adoption. Policy support can play an important role in improving economic feasibility. Carbon credits and/or climate incentives, as well as subsidies for carbon removal technologies,

may contribute to offset production costs and motivate investment in biochar production plants [182]. In addition, integrating biochar production into biorefinery systems can improve profitability by generating multiple products, including biochar, syngas, bio-oil, and heat [184].

8.4 Commercial biochar producers and certification standards

Biochar production has moved beyond academic research, with several companies now operating at commercial scale. These companies often use certification systems to demonstrate product quality, environmental safety, and carbon-removal credibility. Certification is particularly important for participation in carbon markets and for building confidence among customers, regulators, and investors.

8.4.1 NetZero

NetZero operates in France and Brazil and produces biochar mainly from agricultural residues. The company follows the Puro Standard developed by Puro Earth, which focuses on durable carbon removal. This certification entails independent verification, life cycle assessment, and full carbon storage and emission calculation concerning the use and production of biochar [185, 186]. The use of a certification will increase transparency and the credibility of biochar as a carbon credit.

8.4.2 Biochar works

Biochar Works, based in the United States, uses certification approaches such as the VM0044 methodology for biochar-related greenhouse gas mitigation projects. This framework evaluates long-term carbon storage, environmental safeguards, and climate benefits, particularly for soil applications [187]. These requirements help ensure that certified biochar projects contribute meaningfully to emissions reduction.

8.4.3 Carbon gold

Carbon Gold, based in the UK, adheres to schemes such as the International Biochar Initiative standard and European Biochar Certificate. The schemes stipulate quality and safety requirements for the production and application of biochar, particularly in agricultural applications. The IBI requirements outline composition, stability, and acceptable contaminant levels of biochar, while the EBC addresses emissions tracking, product quality, and European regulations [187, 188] (Table 4).

9 Environment and toxicity

The inherent toxicity and environmental risk of biochar, particularly nanobiochar, have grown to be significant research topics given its emerging application into various environmental and agricultural systems [219]. Though generally regarded as environmentally sound, biochar in nano size is associated with a higher surface reactivity and mobility than biochar, which may contribute to increased interactions with living organisms and ecosystems. Research suggests that nanobiochar may lead to a decrease in soil microbial communities, influence plant growth patterns, and may also pose a risk to aquatic organisms due to increased dispersion and bioavailability [220–224]. Furthermore, materials like biochar may absorb co-contaminants such as heavy metals and organic pollutants and distribute these pollutants within soil and water systems instead of immobilizing

Table 4 Summary of biochar studies indicating lignocellulosic feedstocks, key biochar properties evaluated for different applications, analytical techniques employed, and multi-criteria decision-making (MCDM) tools applied

End-Use Category	Biomass Sources Investigated	Main Evaluation Parameters	Characterization Methods Employed	Decision-Making Approach
Energy and Fuel Production	Grape pomace	Biochar recovery, carbon enrichment, calorific value, and combustion behavior	Proximate and elemental analyses, calorimetry, TG/DTG measurements	Pareto-based compromise optimization
Energy and Fuel Production	Grape pomace, cherry and peach pits, rapeseed, sunflower husks, softwood residues	Moisture content, ash fraction, heating value, and thermal degradation characteristics	Proximate analysis, calorific testing, thermogravimetric analysis	Pareto dominance and compromise programming
Energy and Fuel Production	Fruit-derived seed wastes	Carbon enrichment, energy enhancement, and energy–mass performance indices	Elemental composition and calorific analyses	Multi-criteria optimization using Pareto methods
Energy and Fuel Production	Spent coffee grounds and brewery by-products	Atomic H/C and O/C ratios, energy yield, and upgrading fuel efficiency	Elemental and calorimetric analyses	Compromise programming framework
Energy and Fuel Production	Tree-pruning biomass	Fixed carbon content, ash concentration, elemental ratios, and heating value	Proximate, ultimate, and calorific analyses	Pareto-based ranking strategy
Catalytic Material Development	Grape pomace, fruit stones, oilseed residues, and softwood	Mineral composition (K, Ca, P), carbon concentration, and surface area	EDS, BET surface analysis, and elemental characterization	Compromise programming with Pareto evaluation
Agricultural Soil Improvement	Fruit seed biomass	Nutrient content, elemental ratios, and carbon enhancement metrics	EDS and ultimate analysis	Pareto dominance assessment
Agricultural Soil Improvement	Grape pomace	Carbon content, nutrient availability (N, P, Mg, K), and elemental ratios	Elemental and EDS analyses	Multi-objective optimization approach
Carbon Sequestration, Soil Conditioning, and Energy Storage	Grape pomace, cherry stones, peach stones, rapeseed, sunflower husks, and softwood	Mineral content, carbon concentration, porosity, bulk density, electrical conductivity, and pH	BET analysis, EDS, elemental analysis, conductivity, and pH measurements	Pareto compromise decision model
Water Purification and Pollutant Removal	Algal biomass, wheat straw, sunflower husks, penicillin residues, bamboo, orange peel, rice husk, and paper sludge	Surface chemistry, O/C ratio, pore characteristics, graphitic order (ID/IG), and adsorption/desorption behavior	Physicochemical characterization, Raman spectroscopy, elemental analysis, and surface analytical techniques	Not reported

the contaminants. Long-term impacts concern the persistence and accumulation of these particles within the environment, and also the generation of reactive oxygen species from the biochar materials [225–227]. Although at present data are contradictory because of differences in feedstock, methods of preparation, and experimental conditions, standardized toxicity tests, environmental monitoring, and lifecycle assessments are of necessity for responsible application of biochar and nanobiochar on a large scale [228–232].

10 Conclusion

Although several review articles have discussed specific aspects of biochar applications, most have focused on individual sectors such as environmental remediation, agriculture, energy storage, or catalysis. The novelty of the present review lies in its comprehensive and integrated evaluation of biochar across multiple application domains, including environmental remediation, energy generation and storage, catalysis, agriculture, biomedicine, animal nutrition, and advanced materials. In addition to summarizing recent developments, this review emphasizes the relationship between biochar's physicochemical properties such as surface area, porosity, surface functionality, and mineral composition and its performance in diverse applications. Furthermore, it critically examines how feedstock selection, pyrolysis conditions, and post-synthesis modification strategies influence these properties and ultimately determine the efficiency of application. The review also highlights current challenges, including variability in biochar characteristics, long-term stability, catalyst deactivation, economic feasibility, and scalability concerns, while identifying future research directions focused on process standardization, advanced modification technologies, and life-cycle sustainability assessment. By bringing together these diverse perspectives within a single framework, this review provides a broader understanding of biochar's multifunctional role in supporting circular economy principles and the transition toward sustainable, low-carbon technologies.

Abbreviations

HTC	Hydrothermal Carbonization
DT	Dry Torrefaction
WT	Wet Torrefaction
MT	Microwave Torrefaction
NB	Nanobiochar
SSA	Specific Surface Area
O/C	Oxygen-to-Carbon ratio
H/C	Hydrogen-to-Carbon ratio
BET	Brunauer–Emmett–Teller analysis
EDS	Energy Dispersive Spectroscopy
ID/IG	Intensity ratio of D-band to G-band (Raman spectroscopy)
HHV	Higher Heating Value
EF	Enhancement Factor
EY	Energy Yield
EMCI	Energy–Mass Coefficient Index
CEI	Carbon Enhancement Index
FC	Fixed Carbon
ROS	Reactive Oxygen Species
MB	Methylene Blue
IBI	International Biochar Initiative
EBC	European Biochar Certificate

Author contributions

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Ramandeep Kaur: Drafting and revising the manuscript, giving final approval of the version to be published. All authors have read and approved the final version of the manuscript submitted for publication and are responsible for the final content.

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References

1. Lehmann J. Terra Preta Nova - where to from here? Amazonian Dark Earths: Wim Sombroek's Vis. 2009;473–86. https://doi.org/10.1007/978-1-4020-9031-8_28.
2. Liu WJ, Jiang H, Yu HQ. Science. Emerging applications of biochar-based materials for energy storage and conversion. *Energy Environ Sci*. 2019;12:1751–79. <https://doi.org/10.1039/C9EE00206E>.
3. Zhou Z, Shi D, Qiu Y. Sorptive domains of pine chars as probed by benzene and nitrobenzene. *Environ Pollut*. 2010;158:201–6.
4. Ding W, Dong X, Ime IM, Gao B, Ma LQ. Pyrolytic temperatures impact lead sorption mechanisms by bagasse biochars. *Chemosphere*. 2014;105:68–74. <https://doi.org/10.1016/j.chemosphere.2013.12.042>.
5. Ahmad M, Lee SS, Dou X, Mohan D, Sung JK, Yang JE, Ok YS. Effects of pyrolysis temperature on soybean stover- and peanut shell-derived biochar properties and TCE adsorption in water. *Bioresour Technol*. 2012;118:536–44. <https://doi.org/10.1016/j.biortech.2012.05.042>.
6. Ko CH, Park SH, Jeon JK, Suh DJ, Jeong KE, Park YK. Upgrading of biofuel by the catalytic deoxygenation of Biomass. *Korean J Chem Eng*. 2012;29:1657–65. <https://doi.org/10.1007/s11814-012-0199-5>.
7. No SY. Application of bio-oils from Lignocellulosic biomass to transportation, heat and power generation—a review, *Renew. Sustain —Energy Rev*. 2014;40:1108–25. <https://doi.org/10.1016/j.rser.2014.07.127>.
8. Zhang L, Liu R, Yin R, Mei Y. Upgrading of bio-oil from biomass fast pyrolysis in China: a review, *Renew. Sustain Energy Rev*. 2013;24:66–72. <https://doi.org/10.1016/j.rser.2013.03.027>.
9. Richardson Y, Blin J, Julbe A. A short overview on purification and conditioning of syngas produced by biomass gasification: catalytic strategies, process intensification and new concepts. *Prog Energy Combust Sci*. 2012;38:765–81. <https://doi.org/10.1016/j.peecs.2011.12.001>.
10. Abdoulmoumine N, Adhikari S, Kulkarni A, Chattanathan S. A review on biomass gasification Syngas cleanup. *Appl Energy*. 2015;155:294–307. <https://doi.org/10.1016/j.apenergy.2015.05.095>.
11. Liu N, Charrua AB, Weng CH, Yuan X, Ding F. Characterization of biochars derived from agriculture wastes and their adsorptive removal of atrazine from aqueous solution: a comparative study. *Bioresour Technol*. 2015;198:55–62. <https://doi.org/10.1016/j.biortech.2015.08.129>.
12. Ahmad M, Rajapaksha AU, Lim JE, Zhang M, Bolan N, Mohan D, Vithanage M, Lee SS, Ok YS. Biochar as a sorbent for contaminant management in soil and water: a review. *Chemosphere*. 2014;99:19–33. <https://doi.org/10.1016/j.chemosphere.2013.10.071>.
13. Mohan D, Sarswat A, Ok YS, Pittman CU Jr. Organic and inorganic contaminants removal from water with biochar, a renewable, low-cost and sustainable adsorbent—a critical review. *Bioresour Technol*. 2014;160:191–202. <https://doi.org/10.1016/j.biortech.2014.01.120>.
14. Lone AH, Najar GR, Ganie MA, Sofi JA, Ali T. Biochar for sustainable soil health: a review of prospects and concerns. *Elsevier*. 2015;25:639–53. [https://doi.org/10.1016/S1002-0160\(15\)30045-X](https://doi.org/10.1016/S1002-0160(15)30045-X).
15. Konwar LJ, Boro J, Deka D. Review on latest developments in biodiesel production using carbon-based catalysts, *Renew. Sustain. Energy Rev*. 29 (2014) (2014) 546–564. <https://doi.org/10.1016/j.rser.2013.09.003>.
16. Shen Y. Chars as carbonaceous adsorbents/catalysts for tar elimination during biomass pyrolysis or gasification. *Renew Sustain Energy Rev*. 2015;43:281–95. <https://doi.org/10.1016/j.rser.2014.11.061>.
17. Jiang J, Zhang L, Wang X, Holm N, Rajagopalan K, Chen F, Ma S. Highly ordered macroporous woody biochar with ultra-high carbon content as supercapacitor electrodes. *Electrochim Acta*. 2013;113:481–9. <https://doi.org/10.1016/j.electacta.2013.09.121>.
18. Gupta RK, Dubey M, Kharel P, Gu Z, Fan QH. Biochar activated by Oxygen Plasma for supercapacitors. *J Power Sources*. 2015;274:300–1305. <https://doi.org/10.1016/j.jpowsour.2014.10.169>.
19. Gabhane JW, Bhange VP, Patil PD, et al. Recent trends in biochar production methods and its application as a soil health conditioner: a review. *SN Appl Sci*. 2020;2:1307. <https://doi.org/10.1007/s42452-020-3121-5J.W>.
20. Ramanayaka S, Vithanage M, Alessi DS, Liu WJ, Jayasundera ACA, Ok YS. Nanobiochar: production, properties, and multi-functional applications. *Environ Sci Nano*. 2020;7:3279–302.
21. Koves M, Madar V, Ringer M, Kocsis T. Overview of traditional and contemporary industrial production technologies for Biochar along with quality standardization methods. *Land (Basel)*. 2024;13:1388. <https://doi.org/10.3390/land13091388>.
22. The Technology of Wood.
23. Handbook of Charcoal Making: The Traditional and Industrial Methods - Walter Emrich - Google Libri. 2024. Available online.
24. Olarieta JR, Padro R, Masip G, Rodríguez-Ochoa R, Tello E. Formiguers', a historical system of soil fertilization (and Biochar Production?). *Agric Ecosyst Environ*. 2011;140:27–33. <https://doi.org/10.1016/J.AGEE.2010.11.008>.
25. Sulistyo J, Marsoem N, Kholik A, Yatagai M, Nagatsuka Y. *Charcoal quality improvement using double layer walls in a movable kiln*.
26. Carrari E, Ampoorter E, Verheyen K, Coppi A, Selvi F. Former charcoal kiln platforms as microhabitats affecting understorey vegetation in Mediterranean forests. *Appl Veg Sci*. 2016;19:486–97. <https://doi.org/10.1111/avsc.12238>.
27. El-Gamal EH, Saleh M, Elsokkary I, Rashad M, Abd El-Latif MM. *Comparison between properties of biochar produced by traditional and controlled pyrolysis*; Alexandria Sci. Exch J. 2017;38:412–25.
28. Pandit NR, Mulder J, Hale SE, Schmidt HP, Cornelissen G. Biochar from Kon Tiki flame curtain and other kilns: Effects of nutrient enrichment and kiln type on crop yield and soil chemistry. *PLoS ONE*. 2017;12:e0176378. <https://doi.org/10.1371/journal.pone.0176378>.
29. Bailis R. Modeling climate change mitigation from alternative methods of charcoal production in Kenya. *BioMass BioEnergy*. 2009;33:1491–502. <https://doi.org/10.1016/J.BIOMBIOE.2009.07.001>.
30. Sparrevik M, Cornelissen G, Sparrevik M, Adam C, Martinsen V, Cornelissen G, Cornelissen G. Emissions of gases and particles from charcoal/ biochar production in rural areas using medium-sized traditional and improved retort kilns. *BioMass BioEnergy*. 2015;72:65–73. <https://doi.org/10.1016/j.biombioe.2014.11.016>.

31. Lee EH, Su Park R, Kim H, Park SH, Jung SC, Jeon JK, Kim SC, Park YK. Hydrodeoxygenation of guaiacol over Pt-loaded zeolitic materials. *J Indust Eng Chem*. 2016;37:18–21. <https://doi.org/10.1016/J.JIEC.2016.03.019>.
32. Heidari A, Stahl R, Younesi H, Rashidi A, Troeger N, Ghoreysi AA. Effect of process conditions on product yield and composition of fast pyrolysis of eucalyptus Grandis in fluidized bed reactor. *J Indust Eng Chem*. 2014;20:2594–602. <https://doi.org/10.1016/J.JIEC.2013.10.046>.
33. Demirbas A. Effects of temperature and particle size on bio-char yield from pyrolysis of agricultural residues. *J Anal Appl Pyrol*. 2004;72:243–8. <https://doi.org/10.1016/J.JAAP.2004.07.003>.
34. Gristina L, Scalenghe R. Biochar boom-and-bust cycle. *Nat Sustain*. 2026;9:338–9. <https://doi.org/10.1038/s41893-025-01748-w>.
35. Rasool A, Brožová K, Chromíková J, et al. Conventional and microwave-assisted pyrolysis biochars: Comparative mechanistic insights, structural evolution, and environmental remediation applications. *Biochar 8 Article*. 2026;98. <https://doi.org/10.1007/s42773-026-00601-3>.
36. Xu T, Xu Q, Lei Y, et al. Warming increases CO₂ emissions in biochar-amended cropland soil. *Biochar*. 2026;8. <https://doi.org/10.1007/s42773-026-00628-6>. Article 106.
37. Weber K, Quicker P. Properties of Biochar. *Fuel*. 2018;217:240–61. <https://doi.org/10.1016/J.FUEL.2017.12.054>.
38. Pulido-Novicio L, Hata T, Kurimoto Y, Doi S, Ishihara S, Imamura Y. Adsorption capacities and related characteristics of wood charcoals carbonized using a one-step or two-step process. *J Wood Sci*. 2001;47:48–57. <https://doi.org/10.1007/BF00776645>.
39. Freire MCLC, Alexandrino F, Marcelino HR, Picciani PHdeS, Silva KGdeHe, Genre J, de Oliveira AG. E.S.T. do Egito, Understanding drug release data through thermodynamic analysis. *Mater (Basel)*. 2017;10:651. <https://doi.org/10.3390/MA10060651>.
40. Gholizadeh M, Meca S, Zhang S, Clarens F, Hu X. Understanding the dependence of biochar properties on different types of biomass. *Waste Manag*. 2024;182:142–63. <https://doi.org/10.1016/j.wasman.2024.04.011>.
41. Zhang W, Yan L, Wang Q, Li X, Guo Y, Song W, Li Y. Ball milling boosted the activation of peroxymonosulfate by Biochar for tetracycline removal. *J Environ Chem Eng*. 2021;9:106870. <https://doi.org/10.1016/j.jece.2021.106870>.
42. Raczkiwicz M, Ostolska I, Mašek O, Oleszczuk P. Effect of the pyrolysis conditions and type of feedstock on nanobiochars obtained as a result of ball milling. *J Clean Prod*. 2024;458:142456. <https://doi.org/10.1016/j.jclepro.2024.142456>.
43. Dhir B. Biochar amendment improves crop production in problematic soils, Handb. Assist. Amendment-Enhanced Sustain. Remediat Technol. 2021;189–204. <https://doi.org/10.1002/9781119670391.CH10>.
44. Das O, Sarmah AK. The love–Hate relationship of pyrolysis biochar and water: a perspective, *Sci. Total Environ*. 2015;512–3. <https://doi.org/10.1016/j.scitotenv.2015.01.061>.
45. Gray M, Johnson MG, Dragila MI, Kleber M. Water uptake in biochars: the roles of porosity and hydrophobicity. *Biomass Bioenergy*. 2014;61:196–205. <https://doi.org/10.1016/j.biombioe.2013.12.010>.
46. Byrne CE, Nagle DC. Carbonization of wood for advanced materials applications. *Carbon*. 1997;35(96):259–66. <https://doi.org/10.1016/S0008-6223>.
47. Kumar M, Verma BB, Gupta RC. Mechanical properties of Acacia and eucalyptus wood chars. *Energy Sources*. 1999;21:675–85. <https://doi.org/10.1080/00908319950014425>.
48. Chaubey AK, Pratap T, Preetiva B, Patel M, Singsit JS, Pittman CU, Mohan D. Definitive review of Nanobiochar, *ACS. Omega* (2023).
49. Song B, Chen M, Zhao L, Qiu H, Environment XC-S. of T.T., Physicochemical Property and Colloidal Stability of Micron-and Nano-particle Biochar Derived from a Variety. of Feedstock Sources, Elsevier; 2019.
50. Chaubey AK, Pratap T, Preetiva B, Patel M, Singsit JS, Pittman CU, Mohan D. Definitive review of Nanobiochar, *ACS. Omega* (2023).
51. Oleszczuk P.W. Cwiłka-Bundyra, A.B.-J. of A., Characterization of Nanoparticles of Biochars from Different Biomass, Elsevier; 2016.
52. Vassilev SV, Baxter D, Andersen LK, Vassileva CG. An overview of the chemical composition of Biomass. *Fuel*. 2010;89:913–33. <https://doi.org/10.1016/j.fuel.2009.10.022>.
53. Wang Y, Hu Y, Zhao X, Wang S, Xing G. Comparisons of biochar properties from wood material and crop residues at different temperatures and residence times. *Energy Fuels*. 2013;27:5890–9. <https://doi.org/10.1021/EF400972Z>.
54. Xiang W, Zhang X, Chen K, Fang J, He F, Hu X, Tsang DCW, Ok YS, Gao B. Enhanced adsorption performance and governing mechanisms of ball-milled biochar for the removal of volatile organic compounds (VOCs). *Chem Eng J*. 2020;385:123842. <https://doi.org/10.1016/j.cej.2019.123842>.
55. Lyu H, Gao B, He F, Zimmerman A. C.D.-C.E., Experimental and Modeling Investigations of Ball-Milled Biochar for the Removal of Aqueous Methylene Blue. Elsevier; 2018.
56. Shen T, Tang Y, Lu X, Production ZM. -J. of C., Mechanisms of Copper Stabilization by Mineral Constituents in Sewage Sludge Biochar. Elsevier; 2018.
57. Wang D, Zhang W, Hao X, Zhou D. Transport of biochar particles in saturated granular Media: effects of pyrolysis temperature and particle size. *Environ Sci Technol*. 2013;47:821–8. <https://doi.org/10.1021/ES303794D>.
58. Sekar M, Ponnusamy VK, Pugazhendhi A, Nizetic S, Praveenkumar TR. Production and utilization of pyrolysis oil from solid plastic wastes: a review on pyrolysis process and influence of reactor design. *J Environ Manage*. 2022;302:114046. <https://doi.org/10.1016/J.JENVMAN.2021.114046>.
59. Pinto F, Costa P, Gulyurtlu I, Cabrita I. Pyrolysis of plastic wastes: 2. Effect of catalyst on product yield. *J Anal Appl Pyrol*. 1999;51:57–71. [https://doi.org/10.1016/S0165-2370\(99\)00008-X](https://doi.org/10.1016/S0165-2370(99)00008-X).
60. Chen G, Andries J, Luo Z, Spliethoff H. Biomass pyrolysis/gasification for product gas production: the overall investigation of parametric effects, *Energy Convers. Manage*. 2003;44(02):1875–84. <https://doi.org/10.1016/S0196-8904>.
61. Ayllon M, Aznar M, Sanchez JL, Gea G, Arauzo J. Influence of temperature and heating rate on the fixed bed pyrolysis of meat and bone meal. *Chem Eng J*. 2006;121:85–96. <https://doi.org/10.1016/J.CEJ.2006.04.013>.
62. Wang K, Lin M, Ciucu F, Wierman A, Lin C. Characterizing the impact of the workload on the value of dynamic resizing in data centers. *Perform Eval*. 2015;85–6. <https://doi.org/10.1016/J.PEVA.2014.12.001>.

63. Ge L, Li X, Feng H, Xu C, Lu Y, Chen B, Li D, Xu C. Analysis of the pyrolysis process, kinetics and products of the base components of waste wind turbine blades (Epoxy Resin and Carbon Fiber). *J Anal Appl Pyrol.* 2023;170:105919. <https://doi.org/10.1016/J.JAAP.2023.105919>.
64. Choi YJ, Lee Y, Kim K-M, Park S, Chung Y-S. Higher free thyroxine levels are associated with sarcopenia in elderly Koreans. *Osteoporos Sarcopenia.* 2015;1:127–33. <https://doi.org/10.1016/J.AFOS.2015.11.001>.
65. Xie Y, Wang L, Li H, Westholm LJ, Carvalho L, Thorin E, Yu Z, Yu X, Skreiberg Ø. A critical review on production, modification and utilization of biochar. *J Anal Appl Pyrol.* 2022;161:105405. <https://doi.org/10.1016/j.jaap.2021.105405>.
66. Emrich W. *Handbook of Charcoal Making: The Traditional and Industrial Methods*, Springer Dordrecht, 2013. <https://doi.org/10.1007/978-94-017-0450-2>.
67. Bridgwater AV. Review of fast pyrolysis of biomass and product upgrading. *Biomass Bioenergy.* 2012;38:68–94. <https://doi.org/10.1016/j.biombioe.2011.01.048>.
68. O’Laughlin J, McElligott K, Biochar for environmental management: science and technology, Johannes Lehmann, Stephen M. Joseph, Earthscan, London UK. (2009), 448. *Forest Policy Econ.* 11 (2009), 535–536. <https://doi.org/10.1016/j.forpol.2009.07.001>.
69. Kammen D, energy, D.L.-R. and, appropriate. Review of technologies for the production and use of charcoal, Renew. Approp. Energy Laborat. Report 2005 (2005).
70. Elsevier Y Schenkel, PBertaux S, Elsevier Y, Schenkel PB. *Bioenergy 1998 (1998). Biomass Bioenergy 1998 (1998). Elsevier.*
71. Southern YS-B-CO. in E., Charcoal production: opportunities and barriers for improving efficiency and sustainability, Environment portal. Bio-Carbon Opportun. Eastern South. Afr. 2009 (2009).
72. Klavina K, Klavins J, Veidenbergs I, Procedia DB-E. Charcoal production in a continuous operation retort. *Experimental data processing, Energy Procedia* 2016 (2016). *Elsevier.*
73. Emrich W. *Handbook of Charcoal Making: The Traditional and Industrial Methods*, 2013.
74. Garcia-Nunez JA, Pelaez-Samaniego MR, Garcia-Perez ME, Fonts I, Abrego J, Westerhof RJM, Garcia-Perez M. Historical developments of pyrolysis reactors: a review. *Energy Fuels.* 2017;31:5751–75. <https://doi.org/10.1021/ACS>.
75. bioenergy AB-B. Review of fast pyrolysis of biomass and product upgrading, Elsevier AV, Bridgwater Biomass Bioenergy 2012 (2011). <https://doi.org/10.1016/j.biombioe.2011.01.048>. *Elsevier.*
76. Meier D. technology, O.F.-B., Elsevier D Meier, O Faix, State of the art of applied fast pyrolysis of lignocellulosic materials—A review, *Bioresour. Technol.* 1999 (1999). *Elsevier.*
77. Demirbas, A, Arin G. An overview of biomass pyrolysis. *Energy Sources.* 2002;24:471–82. <https://doi.org/10.1080/00908310252889979>.
78. Ighalo JO, Iwuchukwu FU, Eyankware OE, Iwuozor KO, Olotu K, Bright OC, Igwegbe CA. Flash pyrolysis of Biomass: a review of recent advances. *Clean Technol Environ Policy.* 2022;24:2349–63.
79. Ayyadurai S, Arunachalam KD. Experimental investigations on sugarcane bagasse pyrolytic oil production from flash pyrolysis using a rotary screw reactor. *Biofuels Bioprod Biorefin.* 2022;16:576–86. <https://doi.org/10.1002/bbb.2330>.
80. Cha JS, Park SH, Jung SC, Ryu C, Jeon JK, Shin MC, Park YK. Production and utilization of Biochar: a review. *J Indust Eng Chem.* 2016;40:1–15.
81. Arora S, Jung J, Liu M, Li X, Goel A, Chen J, Song S, Anderson C, Chen D, Leong K, et al. Gasification biochar from horticultural waste: an exemplar of the circular economy in Singapore, *Sci. Total Environ.* 2021;781:146573. <https://doi.org/10.1016/J.SCI.TOTENV.2021.146573>.
82. Shen Y. Chars as carbonaceous adsorbents/catalysts for tar elimination during biomass pyrolysis or gasification. *Renew Sustain Energy Rev.* 2015;43:281–95. <https://doi.org/10.1016/J.RSER.2014.11.061>.
83. Gomez-Barea A, Ollero P, Leckner B. Optimization of char and tar conversion in fluidized bed biomass gasifiers. *Fuel.* 2013;103:42–52. <https://doi.org/10.1016/J.FUEL.2011.04.042>.
84. Demol R, Dufour A, Rogaume Y, Mauviel G. Production of purified H₂, heat, and biochar from wood: comparison between gasification and autothermal pyrolysis based on advanced process modeling. *Energy Fuels.* 2022;36:488–501. <https://doi.org/10.1021/acs.energyfuels.1c03528>.
85. Cha JS, Park SH, Jung SC, Ryu C, Jeon JK, Shin MC, Park YK. Production and utilization of Biochar: a review. *J Indust Eng Chem.* 2016;40:1–15.
86. Brynda J, Skoblia S, Pohorely M, Beno Z, Soukup K, Jeremiš M, Mosko J, Zach B, Trakal L, Syc M, et al. Wood chips gasification in a fixed-bed multi-stage gasifier for decentralized high-efficiency CHP and biochar production: long-term commercial operation. *Fuel.* 2020;281:118637. <https://doi.org/10.1016/J.FUEL.2020.118637>.
87. Ochnio M, Kluska J, Kardaś D. Effects of biochar and ash outflow during updraft partial gasification on process parameters in a moving bed reactor. *Chem Pap.* 2020;74:4047–55. <https://doi.org/10.1007/s11696-020-01214-1>.
88. Puig-Gamero M, Pio DT, Tarelho LAC, Sanchez P, Sanchez-Silva L. Simulation of biomass gasification in a bubbling fluidized bed reactor using Aspen Plus®. *Energy Convers Manage.* 2021;235:113981. <https://doi.org/10.1016/J.ENCONMAN.2021.113981>.
89. Ferreira AI, Ferreira AF, Fernandes EC, Coelho P. Influence of process parameters on biomass gasification: a review of experimental studies in entrained flow reactors and drop-tube furnaces. *BioMass BioEnergy.* 2024;185:107217. <https://doi.org/10.1016/J.BIOMBIOE.2024.107217>.
90. Gonzalez-Arias J, ME, Sanchez J, Cara-Ji, Menez. Hydrothermal carbonization of biomass and waste: a review. *Environ Chem Lett.* 2022;20:211–21.
91. Reza MT, Andert J, Wirth B, Busch D, Pielert J, Lynam JG, Mumme J. Hydrothermal carbonization of biomass for energy and crop production. *Appl Bioenergy.* 2014;1. <https://doi.org/10.2478/apbi-2014-0001>.
92. Khan N, Mohan S, Dinesha P. Regimes of hydrochar yield from hydrothermal degradation of various lignocellulosic biomass: a review. *J Clean Prod* 288 (2021).
93. Karthik V, Kumar PS, Vo DVN, Sindhu J, Sneha D, Subhashini B, Saravanan K, Jeyanthi J. Hydrothermal production of algal biochar for environmental and fertilizer applications: a review. *Environ Chem Lett.* 2021;19:1025–42.
94. Sadaka S, Sharara MA, Ashworth A, Keyser P, Allen F, Wright A. Characterization of biochar from switchgrass carbonization. *Energies (Basel).* 2014;7:548–67. <https://doi.org/10.3390/en7020548>.

95. Sevilla M, Macia-Agullo JA, Fuertes AB. Hydrothermal carbonization of biomass as a route for the sequestration of CO₂: chemical and structural properties of the carbonized products. *BioMass BioEnergy*. 2011;35:3152–9. <https://doi.org/10.1016/j.biombioe.2011.04.032>.
96. Nizamuddin S, Mubarak NM, Tiripathi M, Jayakumar NS, Sahu JN, Ganesan P. Chemical, dielectric and structural characterization of optimized hydrochar produced from hydrothermal carbonization of palm shell. *Fuel*. 2016;163:88–97. <https://doi.org/10.1016/j.fuel.2015.08.057>.
97. Jiang H, Ye Y, Lu P, Zhao M, Xu G, Chen D, Song T. Effects of torrefaction conditions on the hygroscopicity of biochars. *J Energy Inst*. 2021;96:260–8. <https://doi.org/10.1016/j.joei.2021.03.018>.
98. Chen Z, Wang M, Ren Y, Jiang E, Jiang Y, Li W. Biomass torrefaction: a promising pretreatment technology for Biomass utilization, in: *Proceedings of the IOP Conference Series: Earth and Environmental Science*, Institute of Physics Publishing, 2018, p. 113. February 22, 2018.
99. Kiel JHA. *Torrefaction for Biomass Co-firing in Existing Coal-fired Power Stations*, 2005.
100. Mamvura TA, Danha G. Biomass torrefaction as an emerging technology to aid in energy production. *Heliyon* 6 (2020).
101. Chew JJ, Doshi V. Recent advances in biomass pretreatment – Torrefaction fundamentals and technology. *Renew Sustain Energy Rev*. 2011;15:4212–22. <https://doi.org/10.1016/j.rser.2011.09.017>.
102. Fagernas L, Brammer J, Wilen C, Lauer M, Verhoeff F. Drying of biomass for second generation synfuel production. *BioMass BioEnergy*. 2010;34:1267–77. <https://doi.org/10.1016/j.biombioe.2010.04.005>.
103. Bach QV, Skreiberg O. Upgrading biomass fuels via wet torrefaction: a review and comparison with dry torrefaction, *Renew. Sustain Energy Rev*. 2016;54:665–77. <https://doi.org/10.1016/j.rser.2015.10.014>.
104. Yan W, Acharjee TC, Coronella CJ, Vasquez VR. Thermal pretreatment of lignocellulosic Biomass. *Environ Prog Sustain Energy*. 2009;28:435–40. <https://doi.org/10.1002/ep.10385>.
105. Bach QV, Tran KQ, Khalil RA, Skreiberg Ø, Seisenbaeva G. Comparative assessment of wet torrefaction. *Energy Fuels*. 2013;27:6743–53. <https://doi.org/10.1021/ef401295w>.
106. Kostyniuk A, Likozar B. Wet torrefaction of biomass waste into high quality hydrochar and value-added liquid products using different zeolite catalysts. *Renew Energy*. 2024;227:120509. <https://doi.org/10.1016/j.renene.2024.120509>.
107. Kostyniuk A, Likozar B. Wet torrefaction of biomass waste into value-added liquid product (5-HMF) and high quality solid fuel (Hydrochar) in a nitrogen atmosphere. *Renew Energy*. 2024;226:120450. <https://doi.org/10.1016/j.renene.2024.120450>.
108. Hoekman SK, Broch A, Robbins C. Hydrothermal carbonization (HTC) of lignocellulosic biomass. *Energy Fuels*. 2011;25:1802–10. <https://doi.org/10.1021/ef101745n>.
109. Yan W, Acharjee TC, Coronella CJ, Vasquez VR. Thermal pretreatment of lignocellulosic Biomass. *Environ Prog Sustain Energy*. 2009;28:435–40. <https://doi.org/10.1002/ep.10385>.
110. Devaraja UMA, Dissanayake CLW, Gunarathne DS, Chen WH. Oxidative torrefaction and torrefaction-based biorefining of Biomass: a critical review. *Biofuel Res J*. 2022;9:1672–96. <https://doi.org/10.18331/BRJ2022.9.3.4>.
111. Uemura Y, Saadon S, Osman N, Mansor N, Tanoue KI. Torrefaction of oil palm kernel shell in the presence of oxygen and carbon dioxide. *Fuel*. 2015;144:171–9. <https://doi.org/10.1016/j.fuel.2014.12.050>.
112. Esetline D, Thanapal SS, Annamalai K, Ranjan D. Torrefaction of Woody biomass (Juniper and Mesquite) using inert and non-inert gases. *Fuel*. 2013;113:379–88. <https://doi.org/10.1016/j.fuel.2013.04.085>.
113. Yan B, Jiao L, Li J, Zhu X, Ahmed S, Chen G. Investigation on microwave torrefaction: parametric influence, TG-MS-FTIR analysis, and gasification performance. *Energy*. 2021;220:119794. <https://doi.org/10.1016/j.energy.2021.119794>.
114. Gronnow MJ, Budarin VL, Ma'sek O, Crombie KN, Brownsort PA, Shuttleworth PS, Hurst PR, Clark JH. Torrefaction/biochar production by microwave and conventional slow pyrolysis - comparison of energy properties. *GCB Bioenergy*. 2013;5:144–52. <https://doi.org/10.1111/gcbb.12021>.
115. Neves D, Thunman H, Matos A, Tarelho L, Gomez-Barea A. Characterization and prediction of biomass pyrolysis products. *Prog Energy Combust Sci*. 2011;37:611–30. <https://doi.org/10.1016/j.peecs.2011.01.001>.
116. Mohd Fuad MAH, Hasan MF, Ani FN. Microwave torrefaction for viable fuel production: a review on theory, affecting factors, potential and challenges. *Fuel*. 2019;253:512–26. <https://doi.org/10.1016/j.fuel.2019.04.151>.
117. Huang YF, Kuan WH, Chang CC, Tzou YM. Catalytic and atmospheric effects on microwave pyrolysis of corn stover. *Biore-sour Technol*. 2013;131:274–80. <https://doi.org/10.1016/j.biortech.2012.12.177>.
118. Sarker TR, Azargohar R, Dalai AK, Meda V. Enhancement of fuel and physicochemical properties of Canola residues via microwave torrefaction. *Energy Rep*. 2021;7:6338–53. <https://doi.org/10.1016/j.egyr.2021.09.068>.
119. Naghdi M, Taheran M, Brar SK, Rouissi T, Verma M, Surampalli RY, Valero JR. A green method for production of nanobiochar by ball milling- optimization and characterization. *J Clean Prod*. 2017;164:1394–405. <https://doi.org/10.1016/j.jclepro.2017.07.084>.
120. Wallace C, Afzal M, Bioprocessing GS-B. effect of feedstock and microwave pyrolysis temperature on physio-chemical and nano-scale mechanical properties of biochar, *Bioresour. Bioprocess*. 6 (2019). <https://doi.org/10.1186/s40643-019-0268-2>. Springer.
121. Xiao D, Yuan D, He H. Luminescence, M.G.-J., Microwave assisted one-step green synthesis of fluorescent carbon nanoparticles from ionic liquids and their application as novel fluorescence probe for quercetin, *J. Lumin*. 2013 (2013). <https://doi.org/10.1016/j.jlumin.2013.02.032>. Elsevier.
122. Production PK-J. of C., A comprehensive evaluation of inherent properties and applications of nano-biochar prepared from different methods and feedstocks, *J. Clean. Prod*. 2021 (2021). Elsevier.
123. Kumar M, Xiong X, Wan Z, Sun Y, D T-B. Ball milling as a mechanochemical technology for fabrication of novel biochar nanomaterials, *Bioresour. Technol*. 2020 (2020). Elsevier.
124. Karinkanta P. A. Am̄ mala, M. Illikainen, bioenergy, J.N.-B., Fine grinding of wood—Overview from wood breakage to applications, *Biomass Bioenergy* 2018 (2018). Elsevier.
125. Ramezanzadeh H, Reyhanitabar A, Oustan S, Mohammadi MH, E.A.T.M. S, van der Zee. Enhanced sorption of cadmium by using biochar nanoparticles from ball milling in a sandy soil. *Euras Soil Sci*. 2021;54:201–11. <https://doi.org/10.1134/S1064229321020125>.
126. Li R, Deng H, Zhang X, Wang J. A.-B. M, High-Efficiency Removal of Pb (II) and Humate by a CeO₂–MoS₂. Hybrid Magnetic Biochar, Elsevier; 2019.

127. Mahmoud E, El Baroudy A, Ali N, Sleem M. Spectroscopic studies on the phosphorus adsorption in salt-affected soils with or without nano-biochar additions. *Environ Res.* 2020;184:109277. <https://doi.org/10.1016/J.ENVRRES.2020.109277>.
128. Naghdi M, Taheran M, Pulicharla R, Rouissi T, Brar SK, Verma M, Surampalli R. Pine-wood derived nanobiochar for removal of Carbamazepine from aqueous Media: adsorption behavior and influential parameters. *J Chem.* 2019;12:5292–301. <https://doi.org/10.1016/j.arabjc.2016.12.025>.
129. Naghdi M, Taheran M, Brar SK, Kermanshahi-Pour A, Verma M, Surampalli R. Immobilized laccase on oxygen functionalized nanobiochars through mineral acids treatment for removal of carbamazepine. *Sci Total Environ.* 2017;584–5. <https://doi.org/10.1016/j.scitotenv.2017.01.021>.
130. Shan D, Deng S, Zhao T, Wang B, Wang Y, Huang J, Yu G, Winglee J, Wiesner MR. Preparation of ultrafine magnetic biochar and activated carbon for pharmaceutical adsorption and subsequent degradation by ball milling. *J Hazard Mater.* 2016;305:156–63. <https://doi.org/10.1016/j.jhazmat.2015.11.047>.
131. Li R, Zhang Y, Deng H, Zhang Z, Wang JJ, Shaheen SM, Xiao R, Rinklebe J, Xi B, He X, Du J. Removing tetracycline and Hg(II) with ball-milled magnetic nanobiochar and its potential on polluted irrigation water reclamation. *J Hazard Mater.* 2020;384:121095. <https://doi.org/10.1016/j.jhazmat.2019.121095>.
132. Li Y, Zimmerman AR, He F, Chen J, Han L, Chen H, Hu X, Gao B. Solvent-free synthesis of magnetic biochar and activated carbon through ball-mill extrusion with Fe₃O₄ nanoparticles for enhancing adsorption of methylene blue. *Sci Total Environ.* 2020;722:137972. <https://doi.org/10.1016/j.scitotenv.2020.137972>.
133. Jiang C, Bo J, Xiao X, Zhang S, Wang Z, Yan G, Wu Y, Wong C, He H. Converting waste lignin into nano-biochar as a renewable substitute of carbon black for reinforcing styrene-butadiene rubber. *Waste Manag.* 2020;102:732–42. <https://doi.org/10.1016/j.wasman.2019.11.019>.
134. Kumar M, Xiong X, Wan Z, Sun Y, Tsang DCW, Gupta J, Gao B, Cao X, Tang J, Ok YS. Ball milling as a mechanochemical technology for fabrication of novel biochar nanomaterials. *Bioresour Technol.* 2020;312:123613. <https://doi.org/10.1016/j.biortech.2020.123613>.
135. Leng L, Xiong Q, Yang L, Li H, Zhou Y, Zhang W, Jiang S, Li H, Huang H. An overview on engineering the surface area and porosity of biochar. *Sci Total Environ.* 2021;763:144204. <https://doi.org/10.1016/j.scitotenv.2020.144204>.
136. Aller MF. Biochar properties: transport, fate, and impact. *Crit Rev Environ Sci Technol.* 2016;46:1183–296. <https://doi.org/10.1080/10643389.2016.1212368>.
137. Zhao L, Cao X, Ma'sek O, Zimmerman A. Heterogeneity of biochar properties as a function of feedstock sources and production temperatures. *J Hazard Mater.* 2013;256–7. <https://doi.org/10.1016/J.JHAZMAT.2013.04.015>.
138. Zhang Y, Liu S, Zheng X, Wang X, Xu Y, Tang H, Kang F, Yang QH, Luo J. Biomass organs control the porosity of their pyrolyzed carbon. *Adv Funct Mater.* 2017;27. <https://doi.org/10.1002/ADFM.201604687>.
139. Raymundo-Pinero E, Cadek M, Beguin F. Tuning carbon materials for supercapacitors by direct pyrolysis of seaweeds. *Adv Funct Mater.* 2009;19:1032–9. <https://doi.org/10.1002/ADFM.200801057>.
140. Cho DW, Yoon K, Ahn Y, Sun Y, Tsang DCW, Hou D, Ok YS, Song H. Fabrication and environmental applications of multifunctional mixed metal-biochar composites (MMBC) from red mud and lignin wastes. *J Hazard Mater.* 2019;374:412–9. <https://doi.org/10.1016/J.JHAZMAT.2019.04.071>.
141. Liu R, Liu G, Yousaf B, Production QA-J. of C., Operating Conditions-Induced Changes in Product Yield and Characteristics during Thermal-Conversion of Peanut Shell to Biochar. Relation to Economic Analysis. Elsevier; 2018.
142. Mui ELK, Cheung WH, McKay G. Tyre char preparation from waste Tyre rubber for dye removal from effluents. *J Hazard Mater.* 2010;175:151–8. <https://doi.org/10.1016/J.JHAZMAT.2009.09.142>.
143. Yin Q, Ren H, Wang R, Zhao Z. Evaluation of nitrate and phosphate adsorption on Al-modified biochar: influence of Al content. *Sci. Total Environ.* 2018;631–2. <https://doi.org/10.1016/J.SCITOTENV.2018.03.091>.
144. Nieva Lobos ML, Sieben JM, Comignani V, Duarte M, Volpe MA, Moyano EL. Biochar from pyrolysis of cellulose: an alternative catalyst support for the electro-oxidation of methanol. *Int J Hydrogen Energy.* 2016;41:10695–706. <https://doi.org/10.1016/J.IJHYDENE.2016.04.041>.
145. Chen T, Luo L, Deng S, Shi G, Zhang S, Zhang Y, Deng O, Wang L, Zhang J, Wei L. Sorption of tetracycline on H₃PO₄ modified biochar derived from rice straw and swine manure. *Bioresour Technol.* 2018;267:431–7. <https://doi.org/10.1016/J.BIORTECH.2018.07.074>.
146. Yang XB, Ying GG, Peng PA, Wang L, Zhao JL, Zhang LJ, Yuan P, He HP. Influence of biochars on plant uptake and dissipation of two pesticides in an agricultural soil. *J Agric Food Chem.* 2010;58:7915–21. <https://doi.org/10.1021/JF1011352>.
147. Yazdani MR, Duimovich N, Tiraferri A, Laurell P, Borghei M, Zimmerman JB, Vahala R. Tailored mesoporous biochar sorbents from Pinecone biomass for the adsorption of natural organic matter from lake water. *J Mol Liq.* 2019;291:111248. <https://doi.org/10.1016/J.MOLLIQ.2019.111248>.
148. Tsai WT, Liu SC, Hsieh CH. Preparation and fuel properties of biochars from the pyrolysis of exhausted coffee residue. *J Anal Appl Pyrol.* 2012;93:63–7. <https://doi.org/10.1016/J.JAAP.2011.09.010>.
149. Yazdani MR, Duimovich N, Tiraferri A, Laurell P, Borghei M, Zimmerman JB, Vahala R. Tailored mesoporous biochar sorbents from Pinecone biomass for the adsorption of natural organic matter from lake water. *J Mol Liq.* 2019;291:111248. <https://doi.org/10.1016/J.MOLLIQ.2019.111248>.
150. Garcia-Perez M, Wang XS, Shen J, Rhodes MJ, Tian F, Lee WJ, Wu H, Li CZ. Fast pyrolysis of oil mallee woody biomass: effect of temperature on the yield and quality of pyrolysis products. *Ind Eng Chem Res.* 2008;47:1846–54. <https://doi.org/10.1021/IE071497P>.
151. Yang XB, Ying GG, Peng PA, Wang L, Zhao JL, Zhang LJ, Yuan P, He HP. Influence of biochars on plant uptake and dissipation of two pesticides in an agricultural soil. *J Agric Food Chem.* 2010;58:7915–21. <https://doi.org/10.1021/JF1011352>.
152. Anto S, Sudhakar MP, Shan Ahamed T, Samuel MS, Mathimani T, Brindhadevi K, Pugazhendhi A. Activation strategies for Biochar to use as an efficient catalyst in various applications. *Fuel.* 2021;285:119205. <https://doi.org/10.1016/J.FUEL.2020.119205>.
153. Tan Xfei, Liu Yguo, Gu Yling, Xu Y, Zeng Gming, Hu Xjiang, Liu Sbo, Wang X, Liu Smian, Li J. Biochar-based nano-composites for the decontamination of wastewater: a review. *Bioresour Technol.* 2016;212:318–33. <https://doi.org/10.1016/J.BIORTECH.2016.04.093>.
154. Tsai WT, Liu SC, Hsieh CH. Preparation and fuel properties of biochars from the pyrolysis of exhausted coffee residue. *J Anal Appl Pyrol.* 2012;93:63–7. <https://doi.org/10.1016/J.JAAP.2011.09.010>.

155. Garcia-Perez M, Wang XS, Shen J, Rhodes MJ, Tian F, Lee WJ, Wu H, Li CZ. Fast pyrolysis of oil mallee woody biomass: effect of temperature on the yield and quality of pyrolysis products. *Ind Eng Chem Res*. 2008;47:1846–54. <https://doi.org/10.1021/IE071497P>.
156. Nieva Lobos ML, Sieben JM, Comignani V, Duarte M, Volpe MA, Moyano EL. Biochar from pyrolysis of cellulose: an alternative catalyst support for the electro-oxidation of methanol. *Int J Hydrogen Energy*. 2016;41:10695–706. <https://doi.org/10.1016/J.IJHYDENE.2016.04.041>.
157. Yin Q, Ren H, Wang R, Zhao Z. Evaluation of nitrate and phosphate adsorption on Al-modified biochar: influence of Al content. *Sci. Total Environ*. 2018;631–2. <https://doi.org/10.1016/J.SCITOTENV.2018.03.091>.
158. Tafjord J, Regli SK, Dugulan AI, Rønning M, Rytter E, Holmen A, Myrstad R, Yang J. Influence of temperature during pyrolysis of Fe-alginate: unraveling the pathway towards highly active Fe/C catalysts. *Appl Catal Gen*. 2022;644:118834. <https://doi.org/10.1016/J.APCATA.2022.118834>.
159. Amalina F, Krishnan S, Zularisam AW, Nasrullah M. An extensive analysis and environmental sustainability applications of multifunctional biochar developments: current trends and technological advances. *Green Technol Sustain*. 2025;3:100174. <https://doi.org/10.1016/J.GRETS.2025.100174>.
160. Liu X, Zhou J, Liu D. Plasma regulates active sites on biochar to boost peroxomonosulfate activation for phenol degradation. *J Environ Chem Eng*. 2022;10:107833. <https://doi.org/10.1016/J.JECE.2022.107833>.
161. Tong XJ, Li JY, Yuan JH, Xu RK. Adsorption of Cu(II) by biochars generated from three crop straws. *Chem Eng J*. 2011;172:828–34. <https://doi.org/10.1016/J.CEJ.2011.06.069>.
162. Wang L, Xie L, Wu J, Li X, Ma H, Zhou J. Sequential H₃PO₄–CO₂ assisted synthesis of lignin-derived porous carbon: CO₂ activation kinetics investigation and textural properties regulation. *Renew Energy*. 2022;191:639–48. <https://doi.org/10.1016/J.RENENE.2022.04.036>.
163. Naseer U, Du Z, Ahmad A, Farooq S, Yousaf M, Yue T. Advanced electrode materials in electrokinetic technology for remediation of heavy metal-contaminated soil: Recent progress and challenges. *Adv Sustainable Syst*. 2025;9(8):2500197. <https://doi.org/10.1002/adsu.202500197>.
164. Naseer U, Ali M, Younis MA, Du Z, Mushtaq A, Yousaf M, Qiu C, Yue T. Sustainable permeable reactive barrier materials for electrokinetic remediation of heavy metals-contaminated soil: Correction(s) for this article. *Adv Sustainable Syst*. 2024;9(3):2400722. <https://doi.org/10.1002/adsu.202400722>.
165. Naseer U, Ahmad A, Adnan M, Yousaf M, Du Z, Qiu C, Yue T. Spatial distribution and health risk assessment of potentially toxic elements along GT road from Sialkot to Rawalpindi. *Environ Adv*. 2025;20:100632. <https://doi.org/10.1016/j.envadv.2025.100632>.
166. Anto S, Sudhakar MP, Ahamed TS, Samuel MS, Mathimani T, Brindhadevi K, Pugazhendhi A. Activation strategies for Biochar to use as an efficient catalyst in various applications. *Fuel*. 2021;285:119205. <https://doi.org/10.1016/j.fuel.2020.119205>.
167. Hagemann N, Spokas K, Schmidt HP, Kagi R, Bohler MA, Bucheli TD. Activated carbon, biochar and charcoal: linkages and synergies across Pyrogenic Carbon's ABCs. *Water (Switzerland)*. 2018;10. <https://doi.org/10.3390/W10020182>.
168. Igalavithana AD, Choi SW, Shang J, Hanif A, Dissanayake PD, Tsang DCW, Kwon JH, Lee KB, Ok YS. Carbon dioxide capture in biochar produced from pine sawdust and paper mill sludge: effect of porous structure and surface chemistry. *Sci Total Environ*. 2020;739:139845. <https://doi.org/10.1016/J.SCITOTENV.2020.139845>.
169. Medynska-Juraszek A, Alvarez ML, Białowiec A, Jerzykiewicz M. Characterization and sodium cations sorption capacity of chemically modified biochars produced from agricultural and forestry wastes. *Mater (Basel)*. 2021;14:4714. <https://doi.org/10.3390/MA14164714>.
170. Liu WJ, Tian K, Jiang H, Zhang XS, Ding HS, Yu HQ. Selectively improving the bio-oil quality by catalytic fast pyrolysis of heavy-metal-polluted biomass: take copper (Cu) as an example. *Environ Sci Technol*. 2012;46:7849–56. https://doi.org/10.1021/ES204681Y.SUPPL_FILE/ES204681Y_SI_001.PDF.
171. Shen Y, Zhao P, Shao Q, Ma D, Takahashi F, Yoshikawa K. In-situ catalytic conversion of tar using Rice Husk char-supported nickel-iron catalysts for biomass pyrolysis/gasification. *Appl Catal B*. 2014;152:3. <https://doi.org/10.1016/J.APCATB.2014.01.032>.
172. Nath BK, Chaliha C, Kalita E. Iron oxide permeated mesoporous rice-husk nanobiochar (IPMN) mediated removal of dissolved arsenic (As): chemometric modeling and adsorption dynamics. *J Environ Manage*. 2019;246:397–409. <https://doi.org/10.1016/J.JENVMAN.2019.06.008>.
173. Gamiz B, Hall K, Spokas KA, Cox L. Understanding activation effects on low-temperature biochar for optimization of herbicide sorption. *Agronomy*. 2019;9:588. <https://doi.org/10.3390/AGRONOMY9100588>.
174. Tay T, Ucar S, Karagoz S. Preparation and characterization of activated carbon from waste biomass. *J Hazard Mater*. 2009;165:481–5. <https://doi.org/10.1016/J.JHAZMAT.2008.10.011>.
175. Trakal L, Sigut R, Silleroval H, Faturikov a D. Kom arek, Copper removal from aqueous solution using biochar: effect of chemical activation. *Arab J Chem*. 2014;7:43–52. <https://doi.org/10.1016/J.ARABJC.2013.08.001>.
176. Ros A, Lillo-Rodenas MA, Fuente E, Montes-Morán MA, Martín MJ, Linares-Solano A. High surface area materials prepared from sewage sludge-based precursors. *Chemosphere*. 2006;65:132–40. <https://doi.org/10.1016/J.CHEMOSPHERE.2006.02.017>.
177. Ahiduzzaman M, Sadrul AKM, Islam. Preparation of porous bio-char and activated carbon from rice husk by leaching ash and chemical activation. *Springerplus*. 2016;5:1–14. <https://doi.org/10.1186/S40064-016-2932-8/FIGURES/13>.
178. Cheng F, Li X. Preparation and application of biochar-based catalysts for biofuel production. *Catalysts*. 2018;8:346. <https://doi.org/10.3390/CATAL8090346>.
179. Angin D, Altintig E, Kose TE. Influence of process parameters on the surface and chemical properties of activated carbon obtained from biochar by chemical activation. *Bioresour Technol*. 2013;148:542–9. <https://doi.org/10.1016/J.BIORTECH.2013.08.164>.
180. Zhu L, Zhao N, Tong L, Lv Y. Structural and adsorption characteristics of potassium carbonate activated biochar, RSC. *Adv*. 2018;8:21012–9. <https://doi.org/10.1039/C8RA03335H>.
181. Boden AC, Bhawe M, Cipolla L, Kingshott P. Solution deposition conditions influence the surface properties of 3-mercaptopropyl (Trimethoxysilane) (MPTS) films. *Appl Surf Sci*. 2022;602:154282. <https://doi.org/10.1016/J.APSUSC.2022.154282>.

182. Xing T, Sunarso J, Yang W, Yin Y, Glushenkov AM, Li LH, Howlett PC, Chen Y. Ball milling: a green mechanochemical approach for synthesis of nitrogen-doped carbon nanoparticles. *Nanoscale*. 2013;5:7970–6. <https://doi.org/10.1039/C3NR02328A>.
183. Xu X, Zheng Y, Gao B, Cao X. N-Doped biochar synthesized by a facile ball-milling method for enhanced sorption of CO₂ and reactive red. *Chem Eng J*. 2019;368:564–72. <https://doi.org/10.1016/J.CEJ.2019.02.165>.
184. Kundu S, Xia W, Busser W, Becker M, Schmidt DA, Havenith M, Muhler M. The formation of nitrogen-containing functional groups on carbon nanotube surfaces: a quantitative XPS and TPD study. *Phys Chem Chem Phys*. 2010;12:4351–9. <https://doi.org/10.1039/B923651A>.
185. Mu Y. Design HM-CER. NaOH-Modified Mesoporous Biochar Derived from Tea Residue for Methylene Blue and Orange II Removal. Elsevier; 2021.
186. Deng R, Huang D, Zeng G, Wan J, Xue W, Wen X, Liu X, Chen S, Li J, Liu C, et al. Decontamination of lead and tetracycline from aqueous solution by a promising carbonaceous nanocomposite: interaction and mechanisms insight. *Bioresour Technol*. 2019;283:277–85. <https://doi.org/10.1016/j.biortech.2019.03.086>. Elsevier.
187. Sivaranjane R, Senthil Kumar P, Chitra B, Rangasamy G. A critical review on biochar for the removal of toxic pollutants from water environment. *Chemosphere*. 2024;360:142382. <https://doi.org/10.1016/J.CHEMOSPHERE.2024.142382>.
188. Lu J, Wu M, Luo L, Lu R, Zhu J, Li Y, Cai Y, Xiang H, Song C, Yu B. Incorporating iron oxide nanoparticles in polyvinyl alcohol/starch hydrogel membrane with biochar for enhanced slow-release properties of compound fertilizers. *Carbohydr Polym*. 2025;348. <https://doi.org/10.1016/j.carbpol.2024.122834>.
189. Tan X, Liu Y, Zeng G, Wang X, Hu X, Gu Y, Yang Z. Application of biochar for the removal of pollutants from aqueous solutions. *Chemosphere*. 2015;125:70–85. <https://doi.org/10.1016/J.CHEMOSPHERE.2014.12.058>.
190. Amdeha E. Biochar-based nanocomposites for industrial wastewater treatment via adsorption and photocatalytic degradation and the parameters affecting these processes. *BioMass Convers Biorefin*. (2023).
191. Ko JH, Kwak YH, Yoo KS, Jeon JK, Park SH, Park YK. Selective catalytic reduction of NO_x using RDF char and municipal solid waste char based catalyst. *J Mater Cycles Waste Manage*. 2011;13:173–9. <https://doi.org/10.1007/S10163-011-0015-Z>.
192. Li P, Oyang X, Zhao Y, Tu T, Tian X, Li L, Zhao Y, Li J, Xiao Z. Occurrence of perfluorinated compounds in agricultural environment, vegetables, and fruits in regions influenced by a fluorine-chemical industrial park in China. *Chemosphere*. 2019;225:659–67. <https://doi.org/10.1016/J.CHEMOSPHERE.2019.03.045>.
193. Mirbagheri SA, Safieddin Ardebili SM, Kiani Deh M, Kiani. Modeling of the engine performance and exhaust emissions characteristics of a single-cylinder diesel using nano-biochar added into ethanol-biodiesel-diesel blends. *Fuel*. 2020;278:118238. <https://doi.org/10.1016/J.FUEL.2020.118238>.
194. Gwenzi W, Chaukura N, Wenga T, Mtisi M. Biochars as Media for Air Pollution Control Systems: contaminant removal, applications and future research directions. *Sci Total Environ*. 2021;753. <https://doi.org/10.1016/J.SCITOTENV.2020.142249>.
195. J. Li, L. Lin, T. Ju, F. Meng, S. Han, K.C.-... and S.E., microwave-assisted pyrolysis of solid waste for production of high-value liquid oil, syngas, and carbon solids: a review, *Renew. Sustain. Energy Rev*. 2024 (2024). Elsevier.
196. Xiang W, Zhang X, Chen J, Zou W, He F, Hu X, Tsang DCW, Ok YS, Gao B. Biochar technology in wastewater treatment: a critical review. *Chemosphere*. 2020;252:126539. <https://doi.org/10.1016/J.CHEMOSPHERE.2020.126539>.
197. Xiao B, Jia J, Wang W, Zhang B, Ming H, Ma S, Kang Y, Zhao M. A review on magnetic biochar for the removal of heavy metals from contaminated soils: preparation, application, and microbial response. *J Hazard Mater Adv*. 2023;10:100254. <https://doi.org/10.1016/J.HAZADV.2023.100254>.
198. Tran TK, Huynh L, Nguyen HL, Nguyen MK, Lin C, Hoang TD, Hung NTQ, Nguyen XH, Chang SW, Nguyen DD. Applications of engineered biochar in remediation of heavy metal(Loid)s pollution from wastewater: current perspectives toward sustainable Development Goals. *Sci. Total Environ*. 2024;926:171859. <https://doi.org/10.1016/J.SCITOTENV.2024.171859>.
199. Xiao B, Jia J, Wang W, Zhang B, Ming H, Ma S, Kang Y, Zhao M. A review on magnetic biochar for the removal of heavy metals from contaminated soils: preparation, application, and microbial response. *J Hazard Mater Adv*. 2023;10:100254. <https://doi.org/10.1016/J.HAZADV.2023.100254>.
200. Ahn Y, Cho D, Ahmad W, Jo J. J.J.-J., Efficient Removal of Formaldehyde Using Metal-Biochar Derived from Acid. Mine Drainage Sludge and Spent Coffee Waste, Elsevier; 2021.
201. Kouzu M, Kasuno T, Tajika M, Yamanaka S, Hidaka J. Active phase of calcium oxide used as solid base catalyst for transesterification of soybean oil with refluxing methanol. *Appl Catal A: Gen*. 2008;87:2798–806. <https://doi.org/10.1016/j.fuel.2007.10.019>.
202. Kastner JR, Mani S. Catalytic decomposition of tar using iron supported biochar. *Fuel Process Technol*. 2015;130:31–7. <https://doi.org/10.1016/j.fuproc.2014.09.038>.
203. Yan Q, Lu Y, To F, Li Y, Technology FY-CS. synthesis of tungsten carbide nanoparticles in biochar matrix as a catalyst for dry reforming of methane to Syngas. *Catal. Sci. Technol*. 2015 (2015). pubs.rsc.org
204. Safieddin Ardebili SM, Taghipoor A, Solmaz H, Mostafaei M. The effect of nano-biochar on the performance and emissions of a diesel engine fueled with fusel oil-diesel fuel. *Fuel*. 2020;268. <https://doi.org/10.1016/J.FUEL.2020.117356>.
205. Ramachandran S, Thangavelu M, Kamaraj L, Sorakka Ponnappan V, Arumugam R. Ignition analysis of diesel engine propelled with neat biodiesel containing nanoparticles. *Energy Sourc Part A: Recov Util Environ Eff*. 2021. <https://doi.org/10.1080/15567036.2021.1917731>.
206. Tomishige K, Miyazawa T, Asadullah M, Ito SI, Kunimori K. Catalyst performance in reforming of tar derived from Biomass over noble metal catalysts. *Green Chem*. 2003;5:399–403. <https://doi.org/10.1039/B303371F>.
207. Shen Y. Chars as carbonaceous adsorbents/catalysts for tar elimination during biomass pyrolysis or gasification. *Renew Sustain Energy Rev*. 2015;43:281–95. <https://doi.org/10.1016/J.RSER.2014.11.061>.
208. Kastner JR, Mani S, Juneja A. Catalytic decomposition of tar using iron supported biochar. *Fuel Process Technol*. 2015;130:31–7. <https://doi.org/10.1016/J.FUPROC.2014.09.038>.
209. Wang D, Yuan W, Ji W. Char and Char-supported nickel catalysts for secondary Syngas cleanup and conditioning. *Appl Energy*. 2011;88:1656–63. <https://doi.org/10.1016/J.APENERGY.2010.11.041>.
210. Shen Y, Zhao P, Shao Q, Ma D, Takahashi F, Yoshikawa K. In-situ catalytic conversion of tar using Rice Husk char-supported nickel-iron catalysts for biomass pyrolysis/gasification. *Appl Catal B*. 2014;152:3. <https://doi.org/10.1016/J.APCATB.2014.01.032>.

211. Mani S, Kastner JR, Juneja A. Catalytic decomposition of toluene using a biomass derived catalyst, *Fuel Process. Technol.* 114 (2013) 118–125. <https://doi.org/10.1016/J.FUPROC.2013.03.015>
212. Kang M, Park JH, Choi JS, Park ED, Yie JE. Low-temperature catalytic reduction of nitrogen oxides with ammonia over supported manganese oxide catalysts. *Korean J Chem Eng.* 2007;24:191–5. <https://doi.org/10.1007/S11814-007-5031-2>.
213. Singh S, Nahil MA, Sun X, Wu C, Chen J, Shen B, Williams PT. Novel application of cotton stalk as a waste-derived catalyst in the low-temperature SCR-DeNOx process. *Fuel.* 2013;105:585–94. <https://doi.org/10.1016/J.FUEL.2012.09.010>
214. Nguyen HKD, Pham VV, Do HT. Preparation of Ni/biochar catalyst for hydrotreating of bio-oil from microalgae biomass. *Catal Lett.* 2016;146:2381–91. <https://doi.org/10.1007/s10562-016-1873-8>.
215. Cancelliere R, Di Tinno A, Di Lellis AM, Contini G, Micheli L, Signori E. Cost-effective and disposable label-free voltammetric immunosensor for sensitive detection of interleukin-6. *Biosens Bioelectron.* 2022;213:114467. <https://doi.org/10.1016/J.BIOS.2022.114467>.
216. Xiao X, Cao J, Meng X, Le D, Li L, Ogawa Y. K.S. Fuel synthesis gas production from catalytic gasification of waste biomass using nickel-loaded brown coal char, *Fuel* (2013). *Elsevier*.
217. Ren S, Lei H, Wang L, Bu Q, Chen S. J.W.-J. of A., *Biofuel Production and Kinetics Analysis for Microwave Pyrolysis of Douglas Fir Sawdust Pellet.* Elsevier; 2012.
218. Alkharabsheh HM, Seleiman MF, Battaglia ML, Shami A, Jalal RS, Alhammad B, Almutairi KF, Al-Saif AM, Sa AMA. Biochar production and characteristics, its impacts on soil health, crop production, and yield enhancement: a review. *MDPI Com.* 2021. <https://doi.org/10.3390/agronomy11050993>.
219. Technology B. L.-E.S., energy diversity brings stability, pure.psu.edu (2006).
220. Kant Bhatia S, Palai AK, Kumar A, Kant Bhatia R, Kumar Patel A, Kumar Thakur V, Yang YH. Trends in renewable energy production employing biomass-based biochar. *Bioresour Technol.* 2021;340:125644. <https://doi.org/10.1016/J.BIORTECH.2021.125644>.
221. Ahuja V, Palai A, Kumar A. A.P.-J. of A, Biochar: empowering the future of Energy production and storage, *J. Anal. Appl. Pyrolysis.* 2024 (2024). *Elsevier*.
222. Ahuja V, Palai AK, Kumar A, Patel AK, Farooque AA, Yang YH, Bhatia SK. Biochar: empowering the future of energy production and storage. *J Anal Appl Pyrol.* 2024;177:106370. <https://doi.org/10.1016/J.JAAP.2024.106370>.
223. Senthil C, Lee CW. Biomass-derived biochar materials as sustainable energy sources for electrochemical energy storage devices. *Renew Sustain Energy Rev.* 2021;137:110464. <https://doi.org/10.1016/J.RSER.2020.110464>.
224. Zheng D, Wang K, Bai B. A critical review of sodium alginate-based composites in water treatment. *Carbohydr Polym* 331 (2024).
225. Ma ZW, Liu HQ, Lü QF. Porous biochar derived from tea saponin for supercapacitor electrode: effect of preparation technique. *J Energy Storage.* 2021;40:102773. <https://doi.org/10.1016/j.est.2021.102773>.
226. Lee J, Kim KH, Kwon EE. Biochar as a catalyst. *Renew Sustain Energy Rev.* 2017;77:70–9. <https://doi.org/10.1016/J.RSER.2017.04.002>.
227. Melero J, Iglesias J, Chemistry GM-G. heterogeneous acid catalysts for biodiesel production: current status and future challenges. *Green Chem.* 2009. <https://doi.org/10.1039/b902086a.pubs.rsc.org>.
228. Atadashi I, Aroua M. A.A.-J. of Industrial, The effects of catalysts in biodiesel production: a review, *J. Indust. Eng. Chem.* (2013). *Elsevier*.
229. Li M, Zheng Y, Chen Y, Zhu X. Biodiesel production from waste cooking oil using a heterogeneous catalyst from pyrolyzed rice husk. *Bioresour Technol.* 2014;154:345–8. <https://doi.org/10.1016/j.biortech.2013.12.070>.
230. Kostić MD, Bazargan A, Stamenković OS, Veljković VB, McKay G. Optimization and kinetics of sunflower oil methanolysis catalyzed by calcium oxide-based catalyst derived from Palm kernel Shell Biochar. *Fuel.* 2016;163:304–13. <https://doi.org/10.1016/J.FUEL.2015.09.042>.
231. Bazargan A, Kostić MD, Stamenković OS, Veljković VB, McKay G. A calcium oxide-based catalyst derived from palm kernel shell gasification residues for biodiesel production. *Fuel.* 2015;150:519–25. <https://doi.org/10.1016/j.fuel.2015.02.046>
232. Liu X, Piao X, Wang Y, Zhu S. Calcium ethoxide as a solid base catalyst for the transesterification of soybean oil to biodiesel. *Energy Fuels.* 2008;22:1313–7. <https://doi.org/10.1021/EF700518H>.
233. Yung MM, Jablonski WS, Magrini-Bair KA. Review of catalytic conditioning of biomass-derived syngas. *Energy Fuels.* 2009;23:1874–87. <https://doi.org/10.1021/EF800830N>.

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