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Biochar: A Sustainable Solution for Mitigating Greenhouse Gas Emissions and Enhancing Soil Productivity—A Review

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ABSTRACT

Greenhouse gases (GHGs) resulting from human activities significantly impact crop production and agricultural sustainability, necessitating innovative solutions to mitigate their effects. One promising approach is employing biochar for GHG mitigation, providing a potential means to offset emissions and enhance crop productivity sustainably. We conducted a comprehensive review by sourcing reputable academic research from various search engines, focusing on terms such as biochar, methane (CH₄), carbon dioxide (CO₂), nitrous oxide (N₂O), GHG, soil organic carbon, agricultural land and cropland. The whole review was divided into three major portions: GHGs, the effect of GHG emission on crop productivity and biochar as an agent of GHG mitigation and further subtopics were designed under each. The review revealed that GHG emissions, including CO₂, CH₄ and N₂O, detrimentally affect crop productivity, posing a serious threat to global food security. Studies demonstrated that biochar aids in mitigating atmospheric CO₂ by sequestration of C. Studies also demonstrated that biochar can positively influence soil physical properties, such as reducing bulk density and enhancing soil moisture, potentially leading to a decrease in soil N₂O emissions. The decrease in soil N₂O emissions was due to the maintenance of optimal oxygen levels in the soil by biochar. Biochar has been utilized to mitigate methane (CH₄) emissions. The reduction in CH₄ due to biochar can be linked to the inhibitory effect of biochar chemicals on soil methanogens. However, further research and widespread adoption of biochar use are imperative to fully realize its global potential.

1 | Introduction

In recent decades, the issue of climate change has emerged as a pressing concern globally, with greenhouse gas emissions (GHGs) being a significant contributor to this phenomenon [1, 2]. Among the various sectors impacted by these emissions, agriculture stands as a critical player, both as a source and a potential mitigator of GHGs [3, 4]. GHGs, including carbon dioxide, methane and nitrous oxide [5], play a pivotal role in climate change due to their heat-trapping properties [6], directly affecting the Earth's temperature and weather patterns [7]. Although CH₄ and N₂O are emitted in lower quantities than CO₂, their global warming potentials (GWPs) over a 100-year horizon are approximately 28 and 273 times that of CO₂, respectively [8, 9].

Numerous lines of evidence support the assertion that human actions have been the primary driver of global warming since the early 20th century [10]. While natural factors like solar radiation variations, volcanic activity, orbital shifts and the carbon cycle also influence Earth's radiation balance [11], the predominant impact since the late 1700s has been the consistent elevation of GHG concentrations due to human activities [12]. For example, the emissions of GHGs such as CO₂ and N₂O have been reported to occur during dung decomposition [13].

This surge in concentrations of GHG emissions is inducing warming effects and exerting influence over diverse aspects of the climate, including surface and ocean temperatures, precipitation patterns and sea levels [14]. The repercussions of

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climate change extend to human health, agriculture, water resources, forests, wildlife and coastal regions, rendering them all vulnerable [10]. These gases, predominantly released through human activities, profoundly influence crop production and agricultural sustainability.

It is now widely accepted that agriculture is the main source of anthropogenic N_2O [8, 15, 16]. Agriculture contributes to 60% of the global N_2O emissions [17]. Agricultural soils are recognized as the major source of atmospheric N_2O , globally contributing 1.7–4.8 Tg N yr^{-1} [18]. Agriculture accounts for nearly 12% of global anthropogenic GHG emissions [19]. The concentration of carbon dioxide (CO_2) in the atmosphere has continued to rise and is now nearly 100 parts per million higher than it was before the industrial revolution [20].

Based on the aforementioned points, it is imperative to address GHG emissions from agricultural soils, and one proposed method is the use of biochar [21, 22]. Biochar, created through pyrolysis of organic materials at high temperatures in the absence of oxygen, possesses key characteristics such as alkali pH, carbon-rich composition, large surface area and high porosity, making it a suitable soil amendment [23]. Leveraging its physical and chemical properties, biochar application is advocated as a potential approach to enhance soil quality, boost crop yield, mitigate GHG emissions and promote soil carbon sequestration [24].

Although numerous studies have demonstrated the potential of biochar to improve soil quality, enhance microbial activity and reduce GHG emissions, the existing evidence remains fragmented and often inconsistent. For example, Kumar et al. [25] explored biochar's role as a catalyst/support in advanced oxidation processes but did not deeply link that to soil-based GHG mitigation within cropping systems. Meanwhile, Sharma et al. [25] documented improvements in soil physical, chemical and biological properties by enhancing soil structure [26], moisture retention, cation exchange capacity (CEC) and nutrient availability, while also stimulating beneficial microbial activity with biochar, and Ralebitso-Senior and Orr [27] analysed how biochar influences microbial communities. However, each of these focused on isolated effects rather than the integrated pathway connecting soil-health enhancements to GHG flux reductions. Similarly, Ambika et al. [28] addressed engineered biochar for Cr(VI) remediation, and Iboko et al. [29] conducted a meta-analysis of biochar + nitrogen fertilizer effects on GHG emissions—but systematic linkage of these outcomes with productivity gains across agroecosystems remains limited. Moreover, variations in biochar feedstock, pyrolysis conditions, soil types and climatic environments have led to contradictory findings, limiting the generalization of results. Long-term field data, particularly from tropical and sub-Saharan agricultural systems, are also scarce, while interactions between biochar and other management practices such as fertilizer application and tillage remain poorly understood. Consequently, there is a need for a comprehensive synthesis that bridges these disciplinary and contextual gaps. This review therefore aims to critically evaluate existing literature to elucidate how and under what conditions biochar serves as a sustainable solution for mitigating GHG emissions and enhancing soil productivity, with emphasis on mechanisms, environmental contexts and implications for climate-smart agriculture.

Therefore, the overall objective of this review is to critically evaluate the potential of biochar as a sustainable solution for mitigating GHG emissions while enhancing soil productivity across agricultural systems.

1.1 | GHGs

1.1.1 | Methane (CH_4)

Methane (CH_4) is a potent GHG with a GWP approximately 27–30 times stronger than CO_2 over a 100-year horizon [8, 10]. Its atmospheric concentration has risen steeply in recent decades, reaching approximately 1875 ppb—the highest in at least 800,000 years and about 2.5 times preindustrial levels [30, 31]. The recent acceleration in CH_4 accumulation has been attributed not only to human activities but also to increasing emissions from natural wetlands driven by climate warming [32], underscoring its sensitivity to climatic feedback.

Anthropogenic activities remain the dominant source of CH_4 , with fossil fuel extraction, livestock production and croplands jointly contributing roughly 50% of global emissions [33, 34]. Agricultural soils and management practices play a key role in shaping CH_4 dynamics. Nitrogen fertilizers, for instance, influence CH_4 emissions by altering soil redox balance and microbial processes, stimulating methanogenesis while inhibiting methanotrophic activity [35–37]. Excessive nitrogen rates can exacerbate CH_4 fluxes, particularly in flooded environments.

Rice paddies remain the largest anthropogenic source of agricultural CH_4 , emitting substantial amounts under anaerobic soil conditions [38, 39]. With climate change intensifying heat and rainfall variability, rice systems face dual challenges: declining productivity and rising CH_4 emissions. Globally, rice cultivation is estimated to account for approximately 10%–12% of anthropogenic CH_4 emissions, and up to 18% when broader farm-level sources are included [40, 41]. Warmer temperatures and prolonged flooding further enhance methanogenic pathways, raising concerns for tropical and Asian rice-growing regions.

Recent studies have introduced both refined and transformative mitigation options. Proven field-level interventions include alternate wetting and drying (AWD), which can cut CH_4 emissions by 30%–70% without yield penalties [39, 42]. Precision fertilization, nitrification inhibitors, deep placement of urea and integrated nutrient management improve nitrogen-use efficiency while reducing CH_4 generation [43]. The deployment of low-emission or methane-suppressing rice cultivar breeding lines with modified root exudation and rhizosphere microbiomes has gained traction in recent CH_4 mitigation research [44]. Advanced biological and soil-based interventions are also emerging. These include the deliberate enhancement of anaerobic methanotrophic archaea to boost in-soil CH_4 oxidation, the use of biochar and organic amendments to suppress methanogenesis and the adoption of climate-adaptive cultivation systems that reduce flooding durations and improve soil aeration [43, 45]. Collectively, these innovations highlight the increasing potential for integrating microbial ecology, soil science and precision agriculture to sustainably curb methane emissions in crop-based systems.

1.1.2 | Carbon Dioxide (CO_2)

CO_2 is the primary GHG, with levels rising from 280 ppm (preindustrial) to 415 ppm today [46]. Human activities—fossil

fuel combustion and deforestation—drive these increases. Agriculture, forestry and land use account for ~24% of CO₂ emissions [47]. Deforestation, reducing carbon sinks, exacerbates emissions [48].

The concentration of carbon dioxide (CO₂) in the air rose to 411.43 parts per million (ppm) in 2019 from 315.98 ppm in 1959, as depicted in Figure 1 [50]. Scientific estimates suggest that the combined impact of elevated CO₂ levels and positive water feedback could lead to a 3°C–5°C increase in the average global surface temperature by the Year 2100 [47]. Manure application, while enhancing soil fertility, contributes to CO₂ emissions via microbial decomposition and anaerobic processes [51]. Improper manure storage further intensifies emissions [52].

1.1.3 | Nitrous Oxide (N₂O)

Though less abundant, N₂O is 300 times more potent than CO₂ [53] and constitutes ~6% of global GHG emissions [54]. Agricultural soils contribute ~78% of anthropogenic N₂O emissions [55]. Synthetic fertilizers and manure accelerate N₂O release through microbial processes [56], with significant implications for ozone depletion [57].

1.2 | Effect of GHG Emissions on Crop Productivity

GHG emissions, including carbon dioxide (CO₂), methane (CH₄) and nitrous oxide (N₂O), significantly threaten global food security by altering climate conditions essential for crop productivity.

1.2.1 | Carbon Dioxide and Crop Growth

Elevated atmospheric CO₂ can enhance photosynthesis, potentially increasing crop yields. However, this effect varies across species and may be offset by other climate stressors such as temperature fluctuations, ozone alterations and nutrient limitations [58]. Increased CO₂ also reduces the protein and nitrogen content in crops like alfalfa and soybean, lowering forage quality for livestock [58]. Additionally, C3 plants experience decreased zinc, iron and protein concentrations under elevated CO₂, affecting human nutrition [59, 60].

1.2.2 | Temperature Rise, Heat Stress and Growth Disruptions

Global warming induced by GHGs leads to higher average temperatures, causing heat stress that disrupts photosynthesis, flowering and fruit formation. Elevated temperatures reduce the efficiency of Rubisco, a key enzyme in carbon fixation [61, 62]. Heat stress also damages thylakoid membranes, impairing photosynthesis [63, 64]. Additionally, high temperatures decrease pollen viability and cause floral abnormalities, reducing reproductive success [65, 66].

In Figure 2, the correlation between the average annual temperature and the yearly maize yield from 2004 to 2010 in the research location is illustrated. In 2004 and 2005, as the temperature rose from 24.79°C to 24.87°C, respectively, the maize yield decreased from 1.4 to 1.29 t/Ha. This trend continued into 2006, with a temperature increase to 26.48°C resulting in a further decline in maize yield to 1.22 t/Ha.

Evapotranspiration rises with increasing temperatures, exacerbated by prolonged droughts [68]. Heat stress dehydrates rice plants, leading to crop loss [69]. Higher temperatures extend the warm season, shortening crop cycles and reducing maize yields by affecting pollination and seed germination [70]. A 1°C rise in temperature can decrease maize yield by 10% [71]. Climate change is also expected to extend growing seasons, potentially disrupting crop adaptation and yield [72, 73].

1.2.3 | Changes in Precipitation Patterns

Climate change alters rainfall patterns, leading to more intense or prolonged droughts. These shifts disrupt crop cycles, affecting germination, water availability and nutrient uptake. With over 80% of global crop yields dependent on rainfall, such changes have critical implications [74]. Figure 3 illustrates the correlation between yearly precipitation and maize crop yield from 2004 to 2015 within the specified study region. In 2004, there was a reduction in rainfall from 1456.7 to 1286.2 mm, which coincided with a decrease in maize yield from 1.4 to 1.29 t/Ha. This pattern persisted into 2005. However, a notable shift occurred in 2006 when maize yield dropped to 1.22 t/Ha, despite a significant rise in rainfall to 1470.7 mm. Between 2006 and 2007, there was

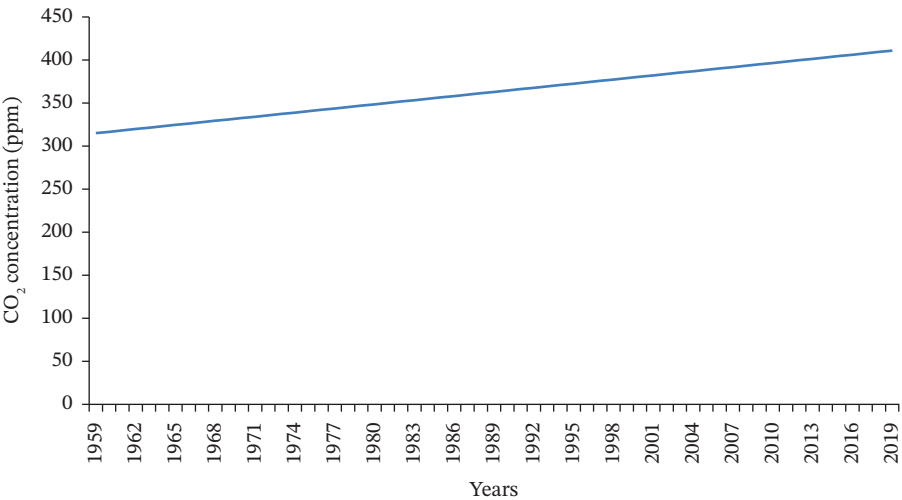


FIGURE 1 | The increase in CO₂ concentration in the atmosphere (source [49]).

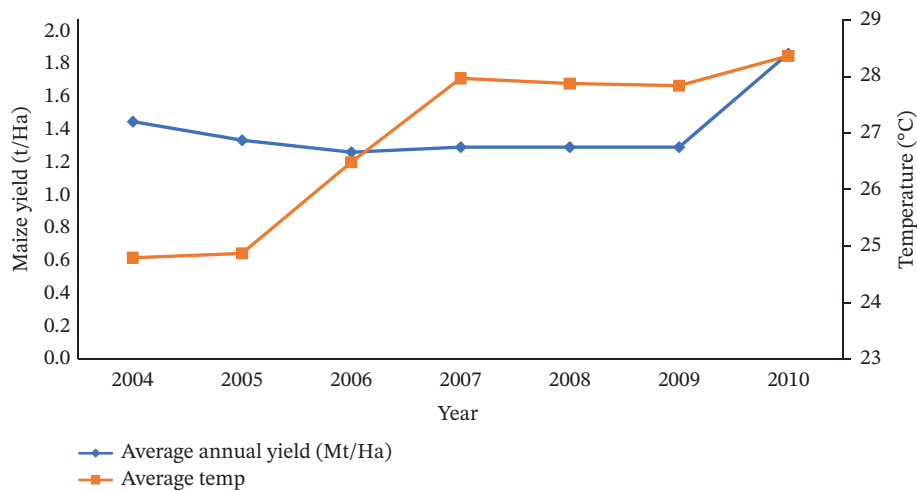


FIGURE 2 | Relationship between total average annual temperature (°C) and annual maize yield (t/Ha) (sources: [67]).

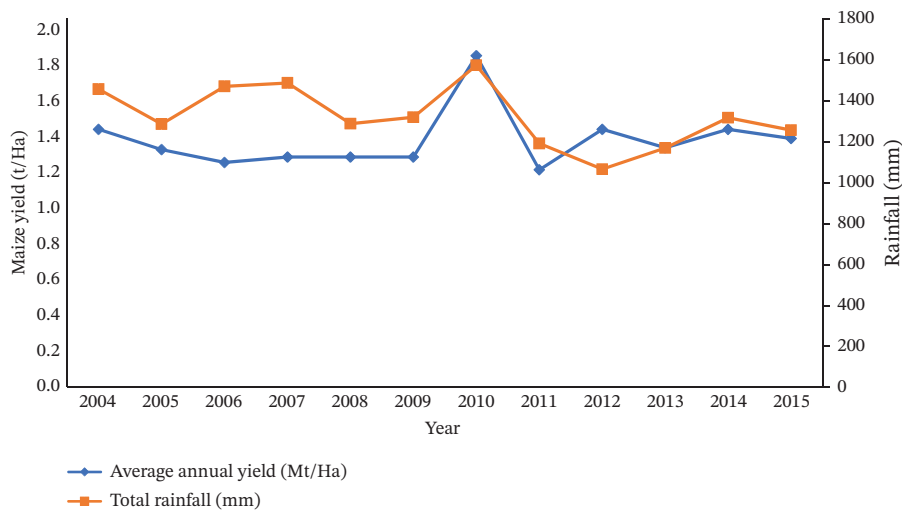


FIGURE 3 | Relationship between annual rainfall (mm) and annual maize yield (t/Ha) (sources: [67]).

a slight increase in rainfall, and correspondingly, maize yield saw a slight rise. This trend remained consistent in 2008 and 2009, maintaining a steady maize yield of 1.25 t/Ha.

Insufficient soil moisture weakens plant resilience, increasing susceptibility to pests and diseases [75]. Conversely, excessive rainfall in regions like the Atlantic coast and European mountains leads to yield losses, poor soil workability and reduced operational days for machinery [74]. In Nigeria, extreme weather events cause crop production fluctuations, affecting prices and food security [76]. Similarly, monsoon-induced floods in Thailand threaten agricultural output [77].

1.2.4 | Pests, Diseases and Climate Change

Rising temperatures favour pests and diseases, altering their distribution and increasing infestations [78]. Climate change drives insect population dynamics, worsening crop losses [79]. Warmer conditions create ecological niches for pest migration [80] and facilitate plant infections by altering pathogen evolution and host–pathogen interactions [81, 82]. Elevated CO₂ intensifies crop diseases, including powdery mildew in cucurbits and wheat

blight [83, 84]. Additionally, humidity and heat worsen potato blight and oilseed rape canker [85].

1.2.5 | Water Resources and Soil Fertility

Climate change disrupts precipitation and evaporation cycles, reducing irrigation water availability and increasing water pollution risks [86]. Saltwater intrusion from rising sea levels damages water infrastructure [86]. Soil organic matter decomposition releases dissolved organic matter, impacting water quality [87]. Climate-driven soil degradation accelerates erosion, nutrient leaching and loss of soil organic matter, increasing compaction and reducing fertility [88–90]. Higher temperatures intensify organic matter decomposition, lowering CEC and leading to soil acidification [91, 92].

1.3 | Biochar Effects on Soil Properties

The application of biochar to agricultural soils has been shown to influence a range of physical, chemical and biological soil properties, thereby potentially enhancing soil productivity and contributing to GHG mitigation.

1.3.1 | Soil Physical Properties

Studies have shown that biochar application can improve soil structure, reduce bulk density and improve aggregate stability. For example, Ebido et al. [93] found that rice husk biochar (RHB) added to a coarse-textured Ultisol increased soil organic carbon (SOC) and improved aggregate stability by up to ~17% at the highest rate ($\approx 60 \text{ t ha}^{-1}$) compared to the unfertilized control. Improvements in porosity and water-holding capacity have also been reported, particularly in sandy soils: biochar's porous structure increases spaces for water retention and microbial habitat, thus improving retention and movement of water in the soil profile. More broadly, reviews show that biochar tends to reduce soil bulk density, increase porosity and improve aggregate stability/mean weight diameter (MWD) [94]. For example, an 8-year field experiment in northeast China in a Mollisol found aggregate stability increased by ~10.9%–23.5% with biochar application; the authors attribute this to the porous structure of biochar and its capacity to serve as binding sites for micro-aggregates [95]. In one long-term U.S. no-till field, application of papermill biochar reduced bulk density from ~1.40 to ~1.26 g cm^{-3} and increased aggregate stability by up to ~67% after 10 years of continuous use [96]. The high specific surface area and porosity of biochar allow for particle–microbe–mineral interactions, enhancing binding of soil particles into aggregates; hydrophilic functional groups on the biochar surface (carboxyl, hydroxyl) improve electrostatic or hydrogen-bond-mediated particle binding.

In sandy or coarse-textured soils, biochar's highly porous matrix offers additional storage of water and enhances plant-available water content (PAWC). For instance, a meta-analysis found that coarse-textured soils amended with biochar had ~30% greater plant-available water capacity [97]. Another review summarizes that biochar improved field water-holding capacity, soil available water content and reduced saturated hydraulic conductivity (sometimes) under certain conditions [98]. The mechanism involved included the fact that biochar provides both macro- and micropores; the micropores hold water against gravity, while macropores enhance infiltration and connectivity; increased porosity and improved aggregate structure reduce compaction and improve water movement and retention. However, some caveats remain. In very weathered tropical soils, one study found biochar increased aggregate stability by up to 33% but had no detectable effect on field-saturated hydraulic conductivity or water-retention characteristics after 10 months [99].

1.3.2 | Soil Chemical Properties

On the chemical side, biochar application exerts multiple beneficial effects on soil fertility and nutrient dynamics. Generally, biochar tends to raise soil pH, particularly when produced from feedstocks at high pyrolysis temperatures ($\geq 500^\circ\text{C}$), owing to the concentration of basic cations such as Ca^{2+} , Mg^{2+} , K^+ and Na^+ in its ash fraction [100]. This liming effect is especially valuable for acidic tropical soils, such as the Alfisols of the Nigerian-derived savannah, which are often prone to nutrient fixation and aluminium toxicity. For instance, Obalum et al. [101], in their review of Nigerian agroecologies, observed that most locally produced biochars are alkaline ($\text{pH} > 8$) and their application frequently ameliorates soil acidity while enhancing nutrient availability and base saturation. Similarly, Oguntunde et al. [102]

reported that the application of maize–stalk biochar at $10\text{--}20 \text{ t ha}^{-1}$ increased soil pH from 5.3 to 6.8 in a degraded Alfisol in southwest Nigeria, creating a more favourable environment for crop nutrient uptake. Biochar also enhances CEC and nutrient retention through its high surface area, porous structure and abundance of negatively charged functional groups (carboxyl, phenolic and hydroxyl moieties). These characteristics increase the soil's ability to retain exchangeable cations (NH_4^+ , K^+ , Ca^{2+} , Mg^{2+}), thereby reducing nutrient leaching and improving fertilizer-use efficiency [103]. In tropical and subtropical soils, where leaching losses are common due to high rainfall, the addition of biochar has been shown to significantly enhance nutrient retention and plant nutrient uptake. El-Naggar et al. [104] applied biochars produced from rice straw, silvergrass residues and umbrella tree to sandy soils and reported CEC increases of 906%, 180% and 130%, respectively, compared to unamended soil.

Moreover, biochar serves as a sink and stabilizing agent for SOC. Its carbon is highly aromatic and recalcitrant, contributing to long-term carbon sequestration and improving soil organic matter quality [105]. In addition, biochar can physically protect native organic matter by promoting the formation of organo-mineral complexes and microaggregates, thereby enhancing the stability of SOC [106]. Ebido et al. [93] also reported that RHB addition to an Ultisol significantly increased SOC concentration, supporting the notion that biochar enhances both the labile and stable carbon pools [107]. Furthermore, biochar can modulate nitrogen dynamics by adsorbing ammonium (NH_4^+) and nitrate (NO_3^-), thus reducing N losses through leaching or volatilization [106]. The retention of these ions not only enhances soil fertility but also minimizes environmental pollution from reactive nitrogen. For instance, Li et al. [96] reported that biochar-amended soils retained 35% more $\text{NH}_4^+\text{-N}$ and 28% more $\text{NO}_3^-\text{-N}$ compared to unamended soils, leading to improved nitrogen-use efficiency in soybean systems (Table 1).

1.3.3 | Soil Biological Properties

On the biological and biochemical side, biochar significantly influences soil microbial communities, enzyme activities and overall biochemical functioning. Through its porous structure, surface chemistry and organic carbon content, biochar creates new microhabitats and energy sources for soil microorganisms [100]. These pores can protect microbes from environmental stress (e.g., desiccation and predation) while providing surfaces for colonization, thereby enhancing microbial abundance and diversity [108]. Biochar's porous matrix also modifies soil aeration, moisture and nutrient availability, all of which are key determinants of microbial activity. By improving soil moisture retention, biochar stabilizes the habitat for microbial communities during dry periods, particularly in coarse-textured tropical soils [109]. Furthermore, the sorption of labile organic molecules and toxins on biochar surfaces can buffer microbial processes by reducing substrate loss and toxicity [110]. Biochar amendments often result in increased microbial biomass and enzymatic activity, which in turn accelerate nutrient cycling. For instance, Ebido et al. [93] reported that RHB application in an Ultisol enhanced microbial biomass carbon and increased dehydrogenase and phosphatase activities, reflecting improved biological functioning and nutrient turnover (Table 2).

TABLE 1 | Effects of biochar on soil chemical properties.

Soil chemical property	Main findings	Benefits to soil/crops	Citations
Soil pH (liming effect)	Biochar raises soil pH, especially when produced at $\geq 500^{\circ}\text{C}$ due to high levels of basic cations (Ca^{2+} , Mg^{2+} , K^{+} , Na^{+}). Locally produced Nigerian biochars commonly have $\text{pH} > 8$. Application of maize-stalk biochar ($10\text{--}20\text{ t ha}^{-1}$) increased soil pH from 5.3 to 6.8 in an Alfisol	Reduces soil acidity, alleviates Al toxicity, improves nutrient availability, enhances root growth and fertilizer-use efficiency	[101, 102]
Cation exchange capacity (CEC)	Biochar improves CEC due to high surface area, porosity and negatively charged functional groups (carboxyl, phenolic, hydroxyl). In sandy soils, CEC increased by 906%, 180% and 130% using rice straw, silvergrass and umbrella-tree biochars	Enhances nutrient retention (NH_4^{+} , K^{+} , Ca^{2+} , Mg^{2+}), reduces leaching losses, increases fertilizer-use efficiency	[103, 104]
Soil organic carbon (SOC)	Biochar contributes highly aromatic, recalcitrant carbon, improving long-term carbon sequestration. Promotes formation of organo-mineral complexes and microaggregates. Rice husk biochar increased SOC in Ultisols	Enhances stable and labile carbon pools, improves soil structure, boosts microbial activity and increases soil resilience	[93, 105, 106]
Nitrogen dynamics	Biochar adsorbs NH_4^{+} and NO_3^{-} , increases soil N retention and reduces leaching and volatilization. Biochar-amended soils retained 35% more $\text{NH}_4^{+}\text{-N}$ and 28% more $\text{NO}_3^{-}\text{-N}$ compared to controls	Improves nitrogen-use efficiency, reduces fertilizer requirements and lowers environmental pollution from reactive nitrogen	[211, 212]

TABLE 2 | Effects of biochar on soil biological properties.

Biological property	Main findings	Benefits to soil and crop productivity	Citations
Microbial habitat formation	Biochar provides porous microhabitats that support microbial colonization and protect microbes from desiccation, predation and temperature stresses. The pores and surfaces act as ecological niches for bacteria and fungi	Enhances microbial abundance, diversity and resilience; promotes stable microbial networks crucial for nutrient cycling	[100, 108]
Microbial biomass and diversity	Biochar increases microbial biomass carbon, microbial diversity and functional group abundance due to improved aeration, moisture and bioavailable carbon	Greater microbial activity enhances soil fertility, nutrient release and organic matter decomposition	[93, 213]
Soil moisture and aeration effects on microbes	Biochar improved water retention stabilizes microbial habitats, especially in sandy tropical soils. Enhanced aeration supports aerobic microbes while reducing stress periods in dry seasons	Sustains microbial functioning under drought; improves decomposition, nutrient mobilization and soil resilience	[109, 214]
Sorption of labile molecules and toxins	Biochar adsorbs labile organic substrates, pesticides and toxic compounds, reducing their mobility and preventing microbial inhibition	Creates a safer and more buffered biochemical environment for microbes, improving enzymatic activity and soil health	[110, 215]
Enzyme activities (e.g., dehydrogenase, phosphatase, urease)	Biochar stimulates soil enzyme activities involved in carbon, nitrogen and phosphorus cycling. Increased dehydrogenase and phosphatase activities reported in Ultisols after rice husk biochar addition	Faster nutrient cycling, improved organic matter turnover and enhanced availability of N, P and C for crops	[93, 216]
Microbial community structure and functional genes	Biochar shifts microbial communities towards beneficial taxa (e.g., plant growth-promoting rhizobacteria, N-fixers and mycorrhizae). Enhances functional genes related to nitrification, denitrification and carbon metabolism	Improved nutrient-use efficiency, reduced GHG-producing microbes (e.g., CH ₄ methanogens), better crop-soil interactions	[96, 217]
Plant-microbe interactions	Biochar enhances root colonization by beneficial microbes and mycorrhizal fungi; reduces pathogen abundance through improved soil environment and sorption of toxins	Improved plant health, reduced disease incidence, enhanced nutrient uptake and root performance	[218, 219]

1.4 | Biochar as an Agent of GHG Sequestration

At present, crop yield uncertainties and rising GHG emissions have marred the overall productive capacity of agriculture systems, putting future food security targets in jeopardy. Indeed, this peculiar situation emphasizes the importance of transitioning from modern intensive farming to more sustainable agricultural management, which can boost crop productivity while reducing GHG emission.

Biochar, a carbon-rich charcoal material, is produced by a dry carbonization process, either under the complete or partial absence of O₂, at high temperatures ranging from 300°C to 1000°C [111]. Globally, biochar has attracted considerable attention as a versatile organic amendment with significant potential for mitigating the global warming effects [112], increasing crop productivity [100] and C-sequestration [113]. The availability of wide-ranging feedstock materials, as well as the pyrolysis temperature conditions, can produce biochar of varied physical and structural attributes, including but not limited to mechanical strength, porosity, surface area, particle size, and density and structural complexity [100, 114].

Biochar application can replenish key soil nutrients in low-fertility soils due to its unique surface charge density, and the predominant negatively charged surfaces of biochar also promote cation adsorption [115, 116]. Since the sources and sinks of three potent GHGs (CO₂, N₂O and CH₄) are major components of the C budget in ecosystems, the inclusion of biochar as a soil amendment is critical because it can sequester C and, more importantly, enable soil to negate anthropogenic CO₂ emissions [117]. According to estimates, biochar produced from 2.2 Gt of feedstock material can remove up to 0.49 Gt C from the atmosphere each year, implying greater merits for its use as a key climate change mitigation strategy [118].

1.4.1 | Effect of Biochar on CO₂ Sequestration

Carbon sequestration is a process by which atmospheric CO₂ is captured and stored to prevent it from being emitted into the atmosphere [119]. It is essential that the carbon is transferred to a passive carbon pool, i.e., stable or inert, in order to decrease C emission to the atmosphere. Transferring even a small amount of the carbon that cycles between the atmosphere and plants to a much slower biochar cycle would impact greatly on atmospheric CO₂ concentration. Biochar is biologically and chemically more stable than the original carbon form, due to its molecular structure and its origins. Furthermore, it is difficult for the sequestered carbon to be released as CO₂, making this a good method for carbon sequestration [120].

Biochar plays a crucial role in carbon sequestration by capturing and storing carbon in a stable form within the soil. The process involves multiple mechanisms, including:

1. **Pyrolysis and Carbon Stabilization:** Biochar is produced through pyrolysis, a process that involves the thermal decomposition of biomass in an oxygen-limited environment [121]. During pyrolysis, labile carbon (e.g., carbohydrates, proteins) is converted into recalcitrant carbon, forming aromatic structures that resist microbial degradation [122]. A comprehensive 8-year study examined the decomposition of biochar derived from ryegrass using compound-specific ¹⁴C analysis. The results demonstrated an exceptionally

slow decomposition rate, with the biochar losing only $7 \times 10^{-4}\%$ of its carbon content per day under optimal conditions [123]. This suggests that nearly 400 years would be required for just a 1% reduction in its carbon content. This stabilized carbon remains in the soil for hundreds to thousands of years, preventing its rapid decomposition and release as CO₂ back into the atmosphere [124]. This longevity ensures that the sequestered carbon remains stored in the soil, providing a reliable and long-term solution for carbon sequestration;

2. **Reduced Microbial Respiration and Mineralization:** Biochar has a high carbon-to-nitrogen (C:N) ratio, which limits its decomposition by microbes [125]. Its complex chemical structure inhibits microbial respiration and slows down carbon mineralization, reducing CO₂ emissions [126]. Biochar amendments possess the capacity to modify enzymatic activity through their influence on microbial community composition and metabolic processes, thereby instigating alterations in the rates of organic matter decomposition and subsequent CO₂ sequestration [127]. It also promotes microbial interactions that enhance carbon stabilization by forming microbial biofilms on its surface [128].
3. **Soil Carbon Protection via Adsorption and Aggregate Formation:** Biochar adsorbs dissolved organic carbon (DOC), preventing its leaching and subsequent oxidation into CO₂ [129]. The potential for biochar (charcoal produced from pyrolysed filtercake) to mitigate carbon and nutrient leaching in a cultivated Brazilian Ferralsol after vinasse application was evaluated [129]. Results of their experiment revealed that biochar-amended soil preferentially retained high-molecular-weight, humic-like DOC species, as revealed by fluorescence spectroscopy and optical indices. Thus, biochar amendments in vinasse application areas may decrease carbon leaching.
4. **Enhanced Plant Growth and Photosynthetic Carbon Fixation:** By improving soil fertility, water retention and CEC, biochar enhances plant growth [130]. This leads to greater CO₂ uptake by plants through photosynthesis, thereby increasing biomass production. Some of this biomass is eventually returned to the soil as organic matter, further contributing to long-term carbon sequestration.

Several studies have been carried out on the effect of biochar on the sequestration of atmospheric CO₂, with contrasting results. Some studies report a reduction in CO₂ emissions; Sun et al. [130] reported that the application of biochar at the rate of 30 t/ha reduced CO₂ emissions by 31.5% from a pine forest soil. Application of biochar significantly reduced CO₂ emissions as reported by Qiao and Wu [131]. Gui et al. [132] in their work aimed to explore if the extra sorption of carbon dioxide (CO₂) exists in the biochar-amended soil. They put biochar and mineral-rich biochar into soils to perform laboratory CO₂ sorption experiments. Their results demonstrate that all biochars increased soil carbon storage and meanwhile further sorb CO₂ for more carbon sequestration. It has been reported that the biochar technology could deliver emission reductions of 3.4~6.3 Pg CO₂ annually and persist in soil for hundreds to thousands of years [133, 134]. Sheng et al. [135] also found suppression of total CO₂ release in both 500°C- and 700°C-derived biochar-amended soils. By using

a generalized framework for quantifying, the potential contribution biochar can make towards achieving national carbon emissions reduction goals, assuming use of only sustainably supplied biomass, that is, residues from existing agricultural, livestock, forestry and wastewater treatment operations [136]. Results showed that biochar can play a role in worldwide CDR strategies, with carbon dioxide removal potential of $6.23 \pm 0.24\%$ of total GHG emissions in the 155 countries covered based on 2020 data over a 100-year timeframe, and more than 10% of national emissions in 28 countries.

Others report an opposite effect; a field experiment by Zhang et al. [137] reported that wheat straw biochar had significantly increased CO₂ emissions by 12%. Hawthorne et al. [138] found that CO₂ emission from Douglas-fir forest soil was higher under biochar application at the rate of 10% compared to the rate of 1%. Some studies report no effect at all. Fidel et al. [139] observed no reduction in CO₂ emissions after application of biochar in a field study with four cropping systems (continuous corn, switchgrass, low diversity grass mix and high diversity grass-forb mix). Cropping system however had a significant effect in the field study, with soils in grass and grass-forb cropping systems emitting more CO₂ than the continuous corn cropping system. Another field experiment in a pasture ecosystem showed no significant effects of biochar amendment on soil CO₂ emissions [140].

These contrasting effects of biochar on sequestration of atmospheric CO₂ could be because biochar influences soil total CO₂ emissions differently depending on biochar type, soil type and experimental design [135, 141–144]. Cely et al. [145] found that biochar derived from wood chips exhibited a negative priming effect in soils, whereas biochar produced from a mixture of paper sludge and wheat husk induced a positive priming effect. These differences may be attributed to variations in biochar properties, including carbon content, carbon aromaticity, volatile matter, fixed carbon, easily oxidized organic carbon, metal content and phenolic compounds, as well as surface characteristics. Their study indicated that biochar application increased soil CO₂ emissions by approximately 25%.

1.4.2 | Effect of Biochar on N₂O Sequestration

The major source of N₂O emissions from agricultural soils is the application of synthetic nitrogen (N) fertilizer, as highlighted by Wang et al. [144]. Studies demonstrate that biochar can positively influence soil physical properties, such as reducing bulk density [146] and enhancing soil moisture [147], potentially leading to a decrease in soil N₂O emissions. The global trend towards adopting no-tillage and reduced tillage practices is driven by their demonstrated benefits in enhancing soil organic matter status, structural condition and water regime [148, 149]. Extensive studies indicate that reduced tillage induces a significant alteration in soil structure, increasing porosity and decreasing bulk density over the long term [150]. The transition from traditional mouldboard ploughing to no-tillage has been associated with a decrease in N₂O emissions [151].

Literature acknowledges the potential of soil compaction to elevate N₂O emissions from agricultural soils. For instance, Hu et al. [152] conducted a review on the effects of soil compaction on productivity and the environment, using New Zealand as a case study. Their findings suggest that the role of soil

compaction in explaining variability in N₂O emissions remains unclear. Ruser et al. [153] reported a 20% increase in the bulk density of fine silty soil, leading to a rise in N₂O emission. Studies examining N₂O emissions in different contexts further support these observations. In a Czech Republic cattle overwintering area, Simek et al. [154] observed higher N₂O emissions in trampled areas, though statistical significance was not achieved, attributed to high spatial variability. Similarly, a study in Scotland simulated trampling in a wet dairy pasture soil, revealing a threefold increase in N₂O emissions [155]. In New Zealand, van der Weerden and Styles [156] and van der Weerden et al. [157] noted elevated N₂O fluxes from compacted treatments after urine application in pasture on silt loam soil. In oak forests, Goutal et al. [158] found higher N₂O production in trafficked plots compared to the control treatment, particularly below 0.3 m depth, where soil air-filled porosity was significantly reduced. These reactions were because compaction-induced changes in the pore system, as highlighted by Dörner and Horn [159], negatively impact pore size, tortuosity and connectivity. This alteration in the pore structure directly influences fluid transport in soil [160, 161], potentially leading to anaerobic conditions and altered soil processes [162, 163]. In summary, limited pore continuity and reduced gas transport capacity within and between aggregates influence N₂O production, consumption and transport to the soil surface, as outlined by Ball [164].

Rondon et al. [165] were among the first to report a reduction in soil N₂O emissions following biochar application. Conducted in a low-fertility oxisol Colombian savannah, their study revealed a reduction of up to 50% for soybeans and up to 80% for grass in N₂O emissions. Additionally, Liu et al. [166] conducted a laboratory study on two types of biochar derived from rice straw and dairy manure, finding a correlation between reduced copy numbers of the monooxygenase gene *amoA* and the nitrite reductase gene *nirS* (genes responsible for nitrification and denitrification) and a subsequent reduction in N₂O emissions when biochar was applied.

In a study assessing the impact of biochar on soil GHG emissions at both laboratory and field scales, Fidel et al. [139] observed a 27% suppression of N₂O emissions in a corn cropping system. However, they noted no significant effect at the laboratory scale across varying soil temperature and moisture levels. This disparity in N₂O emission results between laboratory and field-scale studies emphasizes that laboratory experiments may not reliably predict the impact of biochar at the field scale. Yanai et al. [147] conducted a brief laboratory incubation study, applying municipal biowaste-derived biochar at a rate of 180 tonnes per hectare. They observed a noteworthy reduction in N₂O emissions within a wetted volcanic ash soil. Correspondingly, Zhang et al. [167] demonstrated a significant decline in total N₂O emissions in a hydroagric stagnic anthrosol by 40%–51% and 21%–28% after the addition of biochar (created via slow pyrolysis of wheat straw at 350°C–550°C at a rate of 40 tonnes per hectare compared to control treatments, with or without N-fertilizer, respectively). This aligns with the results of Sarkhot et al. [168], who also found a 26% reduction in cumulative N₂O flux when using dairy manure-derived biochar (DBC).

Furthermore, Cayuela et al. [169] conducted a comprehensive meta-analysis encompassing both short- and long-term studies evaluating the impact of biochar on N₂O emissions. They observed a 54% reduction in soil N₂O emission under controlled

laboratory conditions and a 28% reduction in field conditions. Their meta-analysis highlighted significant factors influencing N₂O emissions, including the type of feedstock used for biochar production, pyrolysis conditions and the properties of the resulting biochar. Additionally, they found a direct relationship between the reduction of N₂O emissions and the application rates of biochar [169]. The study also proposed that interactions among biochar, soil texture and nitrogen fertilizer form play a pivotal role in affecting soil N₂O emissions [169].

The reduction of soil nitrous oxide (N₂O) emissions by biochar is attributed to several mechanisms, such as: (1) Improved Soil Aeration and Reduced Denitrification: Biochar enhances soil structure, increasing porosity and aeration [170], which promotes oxygen diffusion [171]. This reduces anaerobic conditions, limiting denitrification, a major source of N₂O emissions. The availability of oxygen to soil microorganisms via water-filled pore space (WFPS) depends on soil moisture and aeration, ultimately influencing the activities of nitrifiers and denitrifiers. A study by Bateman and Baggs [172] investigated the contributions of nitrification and denitrification to N₂O emissions at different WFPS levels. The findings revealed that nitrification was the primary cause of N₂O generation at 35%–60% WFPS, while denitrification dominated at 70% WFPS and higher [172]. (2) Adsorption and Retention of Nitrogen Compounds: Biochar has a high surface area and CEC, allowing it to adsorb ammonium (NH₄⁺) and nitrate (NO₃⁻) [173, 174]. By retaining nitrogen, biochar reduces substrate availability for nitrification and denitrification, thereby lowering N₂O production [175, 176]. Zhong et al. [177] investigated the potential for N₂O production through bacterial and fungal nitrification and denitrification in both rhizosphere and nonrhizosphere soils, along with the abundance of microbial genes associated with these processes. Their findings revealed that inorganic fertilizers and biochar significantly influenced N₂O production potential and gene abundance. They concluded that partially replacing inorganic fertilizers with biochar could mitigate N₂O emissions by reducing bacterial nitrification and denitrification. (3) Altering Microbial Communities: Biochar influences soil microbial dynamics, particularly by suppressing denitrifiers responsible for N₂O production. Case et al. [178] investigated the effect of biochar on soil N₂O emissions and N cycling processes by quantifying soil N immobilization, denitrification, nitrification and mineralization rates using ¹⁵N pool dilution techniques and the FLUAZ numerical calculation model. They examined whether biochar amendment affected N₂O emissions and the availability and transformations of N in soils. Results showed that biochar suppressed cumulative soil N₂O production by 91% in near-saturated, fertilized soils. Cumulative denitrification was reduced by 37%, which accounted for 85%–95% of soil N₂O emissions. Biochar also enhances N₂O-reducing bacteria, which convert N₂O to harmless N₂ gas [179]. (4) pH Modification and Inhibition of N₂O Formation: Biochar tends to increase soil pH [114], which can reduce nitrification rates (lowering NO₃⁻ availability). In the study of Ippolito et al. [180] in a hardwood-based fast pyrolysis in which biochar was applied (0 wt.%, 1 wt.%, 2 wt.% and 10 wt.%) to calcareous soil, it was reported that biochar at higher applications dramatically lowers soil NO₃⁻-N concentrations and prevents NO₃⁻-N from accumulating over time. An incubation experiment was performed on the salt-affected soil collected from a three-year consecutive experiment at biochar application gradients of 7.5, 15 and 30 t·ha⁻¹ and under nitrogen (N) fertilization [181]. Biochar addition inhibited nitrification in salt-affected irrigation-

silting soil by shifting the community structures of ammonia-oxidizing bacteria and ammonia-oxidizing archaea and reducing the relative abundance of dominant functional ammonia-oxidizers, such as *Nitrosospira*, *Nitrosomonas* and *Nitrosopumilus*. Biochar also tends to increase soil pH, which can shift denitrification pathways towards complete reduction of N₂O to N₂ gas, minimizing emissions. Biochar was found to suppress N₂O and NO emissions by altering soil pH during denitrification [182]. Using acid acrisols and two biochar types, the study found that biochar's alkalizing effect, not labile carbon, influenced product stoichiometry. Acid-leached biochar lost their suppression ability, confirming pH's critical role in denitrification product dynamics. In order to study the influence of biochar addition on N₂O emissions from soils with different pH levels [183], a 40-day incubation experiment was carried out, and four treatments (control, nitrogen fertilizer application, biochar amendment and N plus biochar amendment) were set up separately in soils with three different natural pH levels (acidic vegetable soil, neutral rice soil and alkaline soil). Results showed that adding biochar significantly decreased N₂O emissions by 20.8% and 47.6% in acidic vegetable soil for both N and no-N addition treatments, respectively. Thus, biochar amendment could be used as an effective management practice for mitigating N₂O emissions from acidic and alkaline soils (Table 3).

1.4.3 | Effect of Biochar on CH₄ Sequestration

Biochar has been utilized to mitigate methane (CH₄) emissions. In a study by Yu et al. [184], a notable reduction in CH₄ emissions from forest soils was observed with the incorporation of 10% w/w chicken manure biochar. Similarly, Xiao et al. [185] demonstrated that biochar significantly enhanced CH₄ uptake in a chestnut plantation in China, irrespective of the application rate. The increase in CH₄ uptake within the soil can be attributed to biochar-induced elevation of soil pH, promoting the growth of methanotrophs [186, 187]. Additionally, biochar application leads to a reduction in soil bulk density and an increase in soil porosity, facilitating CH₄ oxidation and uptake by soil microbes [188, 189]. The reduction in CH₄ due to biochar can be linked to the inhibitory effect of biochar chemicals on soil methanotrophs [190]. The porous structure of biochar provides new habitats for soil microbes, enhancing CH₄ uptake by methanotrophs more than CH₄ production, owing to improved soil aeration [189]. A study by Karhu et al. [189] highlighted that CH₄ uptake increased in biochar-amended soil due to enhanced soil aeration and improved CH₄ diffusion through the soil profile. From a laboratory experiment, Liu et al. [21] found that CH₄ emissions from paddy soil amended with bamboo char and straw char at 2.5% application rate were reduced by 51.1% and 91.2% in 49 days.

Lee et al. [191] reported a global meta-analysis conducted regarding the effectiveness of biochar. The study by Jeffery et al. [187] demonstrated that biochar application effectively mitigates methane (CH₄) emissions while enhancing SOC and crop yield. Their quantitative meta-analysis revealed that biochar substantially reduces CH₄ emissions, especially in flooded (paddy) fields and acidic soils where flooding occurs as part of field management. Rajalekshmi et al. [192] found that the application of RHB in wetland Ultisol significantly enhanced soil carbon content and increased carbon accumulation in rice crops. The study revealed that RHB contributed substantial refractory carbon to the soil, improving its nutrient status and productivity.

TABLE 3 | Effect of biochar on N₂O sequestration.

Theme/factor	Main findings		Benefits/implications	Citations
Synthetic N-fertilizer	Major driver of N ₂ O emissions in agricultural soils		Highlights need for improved N management strategies	[220]
Biochar—physical properties	Biochar reduces bulk density and improves moisture retention		Enhances aeration and reduces N ₂ O generation potential	[146, 147]
Reduced/no-tillage	Improves SOM, soil structure and porosity; lowers bulk density; and decreases N ₂ O emissions compared to conventional ploughing		Supports climate-smart, conservation agriculture practices	[148–151]
Soil compaction	Higher bulk density alters pore connectivity; increases N ₂ O emissions		Avoiding compaction reduces denitrification-induced N ₂ O flux	[152, 153]
Livestock trampling studies	Trampled and compacted areas show higher N ₂ O emissions (sometimes with high spatial variability)		Highlights the risk of emissions hotspots in grazed systems	[154, 155]
Compaction + urine deposition	Compacted pasture soils show intensified N ₂ O flux after urine application		Importance of manure/urine management in grazing systems	[156, 157]
Forest and woodland compaction	Trafficked plots in oak forests show increased N ₂ O production, especially at depth		Forest soil trafficking can influence greenhouse gas dynamics	[158]
Mechanistic understanding of compaction	Reduces pore size, tortuosity, connectivity; restricts gas transport; promotes anaerobic zones		Identifies structural drivers of N ₂ O variability; guides soil management reforms	[159–164]
Early evidence of biochar mitigation	Biochar reduced N ₂ O by up to 50%–80% in oxisols		Strong evidence for mitigation in low-fertility tropical soils	[155]
Biochar impacts on microbial genes	Reduces amoA and nirS gene copies linked to nitrification/denitrification		Demonstrates microbial mechanism behind N ₂ O reduction	[166]
Biochar + no-till systems	No-tillage + biochar significantly lower N ₂ O emissions vs. conventional tillage		Integrated strategies amplify mitigation effects	[221]
High-rate biochar applications	Substantial N ₂ O reductions in volcanic ash soils at 180 t ha ⁻¹		Shows the sensitivity of N ₂ O response in certain soil types	[147]
Wheat-straw biochar	Reduced N ₂ O by 40%–51% (with N) and 21%–28% (without N)		Effective across fertilized and unfertilized systems	[167]
Manure-derived biochar	26% reduction in cumulative N ₂ O emissions		Demonstrates the value of recycled organic waste biochars	[168]
Meta-analysis (biochar)	N ₂ O emissions reduced by 54% (lab) and 28% (field). Effects vary by feedstock, pyrolysis and rate		Strong global evidence base for biochar as a mitigation tool	[169]
Improved aeration mechanism	Biochar improves porosity, oxygen diffusion; reduces anaerobic hotspots		Limits denitrification and associated N ₂ O production	[170–172]
N retention mechanism	Adsorbs NH ₄ ⁺ and NO ₃ ⁻ , decreasing substrates for N ₂ O-producing pathways		Enhances nitrogen-use efficiency	[173–177]
Microbial community shifts	Biochar suppresses denitrifiers; increases N ₂ O-reducing bacteria; reduces cumulative denitrification		Direct biological control of N ₂ O emissions	[178, 179]
pH modification mechanism	Biochar increases soil pH, reducing nitrification rates and shifting denitrification towards complete reduction to N ₂		Particularly effective in acidic or salt-affected soils	[148, 180–183]

TABLE 4 | Effects of biochar on methane (CH₄) sequestration.

Theme/mechanism	Main findings	Benefits/implications	Citations
CH ₄ reduction in forest and plantation soils	CH ₄ emissions reduced with 10% w/w chicken manure biochar; increased CH ₄ uptake in chestnut plantations regardless of rate	Demonstrates biochar effectiveness across forest and perennial systems	[185, 210]
pH enhancement promoting methanotrophs	Biochar increases soil pH, stimulating methanotrophic growth and CH ₄ oxidation	Improves microbial CH ₄ consumption and reduces emissions	[186, 187]
Soil structural improvement (bulk density↓, porosity↑)	Increased soil porosity and reduced bulk density enhance CH ₄ oxidation and diffusion	Supports microbial CH ₄ uptake under better-aerated conditions	[188, 189]
Chemical inhibition of methanotrophs	Certain biochar-derived compounds affect methanotroph communities	Identifies chemical pathways influencing CH ₄ dynamics	[190]
Paddy soil CH ₄ reduction	Bamboo and straw biochar (2.5%) reduced CH ₄ emissions by 51%–91% in 49 days	Strong mitigation potential in flooded rice systems	[191]
Global evidence from meta-analysis	Biochar reduces CH ₄ emissions, improves SOC and increases crop yield, especially in paddy soils and acidic environments	Confirms global-scale CH ₄ mitigation effectiveness	[187, 193]
Rice husk biochar (RHB) effects	RHB improved SOC and rice carbon accumulation; reduced CH ₄ emissions by 50%–60% compared to FYM	Enhances soil quality while lowering greenhouse gas emissions	[193]
Mangrove wood biochar in rice systems	Biochar reduced cumulative CH ₄ emissions by 21%–25% across two seasons; biochar + fertilizer > fertilizer alone	Effective for tropical lowland rice systems	[194]
Corn residue biochar under drip-irrigated maize	At 15–30 t ha ⁻¹ , biochar reduced CH ₄ and N ₂ O emissions, increased SOC and lowered GWP	Valuable mitigation tool in semiarid maize production	[195]
Enhanced aeration suppressing methanogenesis	Biochar increases O ₂ diffusion, reducing methanogenic archaea and increasing methanotroph abundance	Limits CH ₄ production and enhances oxidation in flooded soils	[170, 196–200]
Electron-accepting properties (quinone groups)	Biochar acts as an electron acceptor, promoting anaerobic CH ₄ oxidation; quinone groups reduce CH ₄ by facilitating microbial pathways	Provides alternative pathways for microbial metabolism competing with methanogenesis	[208]
pH modification and nutrient effects	Alkaline biochar (high-temperature pyrolysis) boosts methanotrophic activity in acidic soils	Particularly effective in acidic paddy soils	[148, 184, 209]
Reduced substrate availability (DOC adsorption)	Biochar adsorbs dissolved organic carbon (DOC), reducing precursors for methanogenesis	Limits CH ₄ production at microbial substrate level	[210], 225]

Importantly, RHB application led to a notable 50%–60% reduction in methane (CH_4) emissions compared to farmyard manure (FYM), highlighting its effectiveness in mitigating GHG emissions while enhancing soil and crop quality.

An experiment was conducted to evaluate the potential of mangrove tree wood (*Rhizophora apiculata*) biochar on CH_4 mitigation, soil properties and the productivity of rice cultivated in a clay loam soil in Thailand [193]. The treatment biochar alone significantly reduced cumulative methane (CH_4) emissions compared to the nonamended control: 21.1% in the first season, 24.9% in the second season. The treatment biochar + fertilizer also produced lower CH_4 emissions than the fertilizer-alone treatment. A 2-year field study conducted in the sandy loam soils of Inner Mongolia, China, examined the effects of corn residue-derived biochar on GHG emissions, SOC, and GWP under film-mulched, drip-irrigated maize production [194]. Corn residue-derived biochar, particularly at 15–30 t ha⁻¹, effectively reduced CH_4 and N_2O emissions, improved SOC storage and lowered net GWP, demonstrating its potential as a climate mitigation and soil-improvement strategy in semiarid sandy loam soils under drip-irrigated maize production in northern China.

Biochar reduces methane (CH_4) emissions from the soil through several mechanisms: (1) Enhanced Aeration and Redox Potential: Biochar improves soil structure, increasing aeration and oxygen diffusion [170]. This promotes aerobic microbial activity, suppressing methanogenic (CH_4 -producing) archaea that thrive in anaerobic conditions. The effect of biochar addition on CH_4 emissions, and the abundance and community composition of methanogens and methanotrophs over two rice cultivation seasons were studied [195]. Biochar application decreased CH_4 emissions by reducing methanogenic archaea abundance in the studied flooded paddy soil. Methanogens and methanotrophs regulate CH_4 emissions in paddy soils [196]. Feng et al. [197] and Qin et al. [198] found significant decreases in CH_4 emissions by biochar addition and explained the result by increases in methanotrophic bacteria biodiversity and abundance. The addition of biochar to paddy soil reduces soil bulk density and enhances aeration, thereby suppressing methanogenic activity. This reduction in bulk density is attributed to biochar's high porosity [199]. As a nutrient-rich amendment [200], biochar stimulates rice root growth [201], leading to increased oxygen secretion [202, 203]. Enhanced oxygen availability further inhibits methanogens and their activity. Kim et al. [204] found that biochar application not only boosted rice yield but also reduced CH_4 emissions by improving soil aeration and oxygen supply, thereby suppressing methanogenesis. (2) Electron Transfer and Methane Suppression: Biochar acts as an electron acceptor, facilitating alternative microbial pathways that compete with methanogenesis. Biochar contains quinone, phenolic and other redox-active groups that can accept electrons from microbial metabolism, especially from anaerobic respiration [205]. These functional groups undergo reversible oxidation and reduction, influencing microbial electron transfer. Theoretically, carbonyl and quinone moieties in biochar can function as electron acceptors, facilitating CH_4 consumption in paddy soils. Zhang et al. [206] reported that the quinone ($\text{C}=\text{O}$) structure in biochar contributes to anaerobic CH_4 oxidation. This process can account for up to 50% of total CH_4 consumption in wetlands [207]. Therefore, biochar may help mitigate CH_4 emissions partly by

enhancing anaerobic CH_4 oxidation through its electron-accepting capacity in paddy soils. (3) Nutrient and pH Modification: Biochar can increase soil pH, which inhibits methanogenic archaea that prefer acidic conditions. The alkaline nature of biochar is mainly due to the ash content [148]. Weber and Quicker [208] observed a large decrease in methanotrophic activity when soil pH decreases from 6.3 to 5.6. Generally, at higher pyrolysis temperatures, the biochar produced is higher alkaline, and hence, its application significantly promotes the activity of methanotrophic in acid paddy soil [209]. (4) Reduced Substrate Availability: Biochar adsorbs DOC, decreasing the availability of methanogenic precursors like acetate and hydrogen. Soil DOC serves as a key substrate for methanogens and plays a crucial role in CH_4 production [196]. Yu et al. [184] found that applying hen biochar to paddy fields reduced soil DOC, likely due to adsorption within the biochar pores. Zheng et al. [210] reported that biochar amendment decreased DOC content by 52% and 71% at application rates of 20 and 40 t ha⁻¹, respectively (Table 4).

2 | Conclusion

This review revealed that GHG emissions, including CO_2 , CH_4 and N_2O , detrimentally affect crop productivity, posing a serious threat to global food security. These gases trap heat in the Earth's atmosphere, leading to global warming and climate change, ultimately impacting agriculture and crop yields. Decreased crop productivity due to climate change can have far-reaching economic and social impacts. Reduced yields may lead to food shortages, increased food prices and economic instability, affecting both farmers and consumers, particularly in vulnerable regions. Biochar aids in mitigating atmospheric CO_2 by sequestration of C (for it is difficult for the sequestered carbon to be released as CO_2 , making this a good method for carbon sequestration). Biochar can positively influence soil physical properties, such as reducing bulk density and enhancing soil moisture, potentially leading to a decrease in soil N_2O emissions. The decrease in soil N_2O emissions was due to enhanced oxygen levels in the soil by biochar through improved aeration. Biochar has been utilized to mitigate methane (CH_4) emissions. The reduction in CH_4 due to biochar can be linked to the inhibitory effect of biochar chemicals on soil methanotrophs.

Biochar's potential as an agent of GHG sequestration lies in its ability to effectively capture and store carbon while promoting soil health and reducing emissions of other potent GHGs. When integrated into sustainable land management practices, biochar can play a vital role in mitigating climate change and fostering a more sustainable and resilient agricultural system. However, further research and widespread adoption are essential to fully realize the potential of biochar on a global scale.

2.1 | Potential Areas of Future Research

Future studies should explore the economic feasibility of integrating biochar into agricultural systems by assessing the cost-effectiveness of its production and application relative to potential benefits, including increased crop yields, carbon sequestration and reduced emissions. In addition, research should examine the development and implementation of policies and regulatory frameworks that drive the adoption of biochar in agriculture, as well as assess the potential barriers and incentives

at local, national and international levels for the integration of biochar into sustainable land management practices.

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Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

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References

1. Y. Malhi, J. Franklin, N. Seddon, et al., "Climate Change and Ecosystems: Threats, Opportunities and Solutions," *Philosophical Transactions of the Royal Society B* 375, no. 1794 (2020): 20190104, <https://doi.org/10.1098/rstb.2019.0104>.
2. P. H. Raven and D. L. Wagner, "Agricultural Intensification and Climate Change are Rapidly Decreasing Insect Biodiversity," *Proceedings of the National Academy of Sciences* 118, no. 2 (2021): e2002548117, <https://doi.org/10.1073/pnas.2002548117>.
3. J. Lynch, M. Cain, D. Frame, and R. Pierrehumbert, "Agriculture's Contribution to Climate Change and Role in Mitigation is Distinct From Predominantly Fossil CO₂-Emitting Sectors," *Frontiers in Sustainable Food Systems* 4 (2021): 518039, <https://doi.org/10.3389/fsufs.2020.518039>.
4. S. Lenka, N. K. Lenka, V. Sejian, and M. Mohanty, "Contribution of Agriculture Sector to Climate Change," *Climate Change Impact on Livestock: Adaptation and Mitigation*, ed. V. Sejian, et al. (2015), 548, https://doi.org/10.1007/978-81-322-2265-1_3.
5. A. Shakoor, M. S. Arif, S. M. Shahzad, et al., "Does Biochar Accelerate the Mitigation of Greenhouse Gaseous Emissions From Agricultural Soil?—A Global Meta-Analysis," *Environmental Research* 202 (2021): 111789, <https://doi.org/10.1016/j.envres.2021.111789>.
6. S. A. Montzka, E. J. Dlugokencky, and J. H. Butler, "Non-CO₂ Greenhouse Gases and Climate Change," *Nature* 476, no. 7358 (2011): 43–50, <https://doi.org/10.1038/nature10322>.
7. P. Forster, V. Ramaswamy, P. Artaxo, et al., "Changes in Atmospheric Constituents and in Radiative Forcing," in *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*, ed. S. Solomon (Cambridge University Press, 2007), <http://www.ipcc.ch/pdf/assessment-report/ar4/wg1/ar4-wg1-chapter2.pdf>.
8. EPA, in *Greenhouse Gas Mitigation Potential in U.S. Forestry and Agriculture* (U.S. Environmental Protection Agency, 2005).
9. S. Sakata, A. Z. Akililu, and R. Pizarro, *Greenhouse Gas Emissions Data: Concepts and Data Availability* (2024), OECD Statistics Working Papers 2024/03.
10. IPCC, in *Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the IPCC Sixth Assessment Report* (Cambridge University Press, 2021).
11. D. I. Stern and R. K. Kaufmann, "Anthropogenic and Natural Causes of Climate Change," *Climatic Change* 122, no. 1 (2014): 257–269, <https://doi.org/10.1007/s10584-013-1007-x>.
12. K. R. Shivanna, "Climate Change and Its Impact on Biodiversity and Human Welfare," *Proceedings of the Indian National Science Academy* 88, no. 2 (2022): 160–171, <https://doi.org/10.1007/s43538-022-00073-6>.
13. C. J. Oraegbunam, A. Kimura, T. Yamamoto, et al., "Bacterial Communities and Soil Properties Influencing Dung Decomposition and Gas Emissions Among Japanese Dairy Farms," *Journal of Soil Science and Plant Nutrition* 23, no. 3 (2023): 3343–3348, <https://doi.org/10.1007/s42729-023-01250-2>.
14. M. M. M. Islam, Y. K. Gaihre, M. N. Islam, et al., "Reducing Greenhouse Gas Emissions and Improving Rice Yield: The Influence of Cultivars, Soil Salinity, and Nitrogen Management," *Science of the Total Environment* 997 (2025): 180192, <https://doi.org/10.1016/j.scitotenv.2025.180192>.
15. Y. K. Gaihre, U. Singh, S. M. M. Islam, et al., "Nitrous Oxide and Nitric Oxide Emissions and Nitrogen Use Efficiency as Affected by Nitrogen Placement in Lowland Rice Fields," *Nutrient Cycling in Agroecosystems* 110, no. 2 (2018): 277–291, <https://doi.org/10.1007/s10705-017-9897-z>.
16. J. E. Hickman, M. Havlikova, C. Kroeze, and C. A. Palm, "Current and Future Nitrous Oxide Emissions From African Agriculture," *Current Opinion in Environmental Sustainability* 3, no. 5 (2011): 370–378, <https://doi.org/10.1016/j.cosust.2011.08.001>.
17. OECD Press Release, *Shaping Globalisation* (2000).
18. IPCC, *Changes in Atmospheric Constituents and in Radiative Forcing* (Cambridge University Press, 2007).
19. B. Linquist, K. J. Van Groenigen, M. A. Adviento-Borbe, C. Pittelkow, and C. Van Kessel, "An Agronomic Assessment of Greenhouse Gas Emissions From Major Cereal Crops," *Global Change Biology* no. 1 (2012): 194–209, <https://doi.org/10.1111/j.1365-2486.2011.02502.x>.
20. M. Shibata, S. Koeda, T. Noji, et al., "Design of New Extraction Surfactants for Membrane Proteins From Peptide Gemini Surfactants," *Bioconjugate Chemistry* 27, no. 10 (2016): 2469–2479, <https://doi.org/10.1021/acs.bioconjchem.6b00417>.
21. Y. Liu, M. Yang, Y. Wu, H. Wang, Y. Chen, and W. Wu, "Reducing CH₄ and CO₂ Emissions From Waterlogged Paddy Soil With Biochar," *Journal of Soils and Sediments* 11, no. 6 (2011): 930–939, <https://doi.org/10.1007/s11368-011-0376-x>.
22. Y. Xiao, S. Yang, J. Xu, J. Ding, S. Sun, and Z. Jiang, "Effect of Biochar Amendment On Methane Emissions From Paddy Field Under Water-Saving Irrigation," *Sustainability* 10, no. 5 (2018): 1371, <https://doi.org/10.3390/su10051371>.
23. Z. L. Van, B. Singh, S. Joseph, S. Kunber, A. L. Cowie, and Chan, "Biochar and Emissions of Non-CO₂ Greenhouse Gases From Soil," in *Biochar for Environmental Management: Science and Technology*, ed. J. Lehmann and S. Joseph (Earthscan, 2009), 227–250.
24. D. A. Laird, R. C. Brown, J. E. Amonette, and J. Lehmann, "Review of the Pyrolysis Platform for Coproducing Bio-Oil and Biochar," *Biofuels, Bioproducts and Biorefining* 3, no. 5 (2009): 547–562, <https://doi.org/10.1002/bbb.169>.
25. M. Kumar, X. Xiong, Y. Sun, et al., "Critical Review on Biochar-Supported Catalysts for Pollutant Degradation and Sustainable Bio-refinery," *Advanced Sustainable Systems* 4, no. 10 (2020): 1900149, <https://doi.org/10.1002/adsu.201900149>.
26. M. Sharma, R. Kaushal, P. Kaushik, and S. Ramakrishna, "Carbon Farming: Prospects and Challenges," *Sustainability* 13, no. 19 (2021): 11122, <https://doi.org/10.3390/SU131911122>.
27. T. K. Ralebitso-Senior and C. H. Orr, "Microbial Ecology Analysis of Biochar-Augmented Soils," in *Book: Biochar Application*, ed. K. Ralebitso-Senior and C. H. Orr (Elsevier Inc, 2016), <https://doi.org/10.1016/B978-0-12-803433-0.00001-1>.
28. S. Ambika, M. Kumar, L. Pisharody, et al., "Modified Biochar as a Green Adsorbent for Removal of Hexavalent Chromium From Various Environmental Matrices: Mechanisms, Methods, and Prospects," *Chemical Engineering Journal* 439 (2022): 135716, <https://doi.org/10.1016/j.cej.2022.135716>.
29. M. P. Iboko, E. R. Dossou-Yovo, S. E. Obalum, et al., "Paddy Rice Yield and Greenhouse Gas Emissions: Any Trade-Off Due to Co Application of Biochar and Nitrogen Fertilizer? A Systematic Review," *Heliyon* 9, no. 11 (2023): e2213, <https://doi.org/10.1016/j.heliyon.2023.e22132>.

30. E. J. Dlugokencky and S. Houweling, "The Global Methane Budget 2000–2017," *Earth System Science Data* 12 (2020): 1561–1623, <https://doi.org/10.5194/essd-12-1561-2020>.
31. E. G. Nisbet, M. R. Manning, E. J. Dlugokencky, et al., "Atmospheric Methane: Comparison Between Methane's Record in 2006–2022 and During Glacial Terminations," *Global Biogeochemical Cycles* 37, no. 8 (2023): <https://doi.org/10.1029/2023GB007875>.
32. R. B. Jackson, M. Saunio, A. Martinez, et al., "Human Activities Now Fuel Two-Thirds of Global Methane Emissions," *Environmental Research Letters* 19, no. 10 (2024): 101002DOI, <https://doi.org/10.1088/1748-9326/ad6463>.
33. A. Shakoor, M. S. Arif, S. M. Shahzad, et al., "Does Biochar Accelerate the Mitigation of Greenhouse Gaseous Emissions From Agricultural Soil? A Global Meta-Analysis," *Environmental Research* 202 (2021): 111789, <https://doi.org/10.1016/j.envres.2021.111789>.
34. J. Broucek, "Production of Methane Emissions From Ruminant Husbandry: A Review," *Journal of Environmental Protection* 5, no. 15 (2024): 1482–1493DOI, <https://doi.org/10.4236/jep.2014.515141>.
35. Z. Wang, X. Zhang, L. Liu, et al., "Inhibition of Methane Emissions From Chinese Rice Fields by Nitrogen Deposition Based on the DNDC Model," *Agricultural Systems* 184 (2020): 102919, <https://doi.org/10.1016/j.agsy.2020.102919>.
36. F. Chai, L. Li, S. Xue, and J. Liu, "Auxiliary Voltage Enhanced Microbial Methane Oxidation Co-Driven by Nitrite and Sulfate Reduction," *Chemosphere* 250 (2020): 126259, <https://doi.org/10.1016/j.chemosphere.2020.126259>.
37. S. M. M. Islam, Y. K. Gaihare, M. R. Islam, et al., "Mitigating Greenhouse Gas Emissions From Irrigated Rice Cultivation Through Improved Fertilizer and Water Management," *Journal of Environmental Management* 307 (2022): 114520, <https://doi.org/10.1016/j.jenvman.2022.114520>.
38. X. Yan, H. Akiyama, K. Yagi, and H. Akimoto, "Global Estimations of the Inventory and Mitigation Potential of Methane Emissions From Rice Cultivation Conducted Using the 2006 Intergovernmental Panel on Climate Change Guidelines," *Global Biogeochemical Cycles* 23, no. 2 (2009): GB2002, <https://doi.org/10.1029/2008GB003299>.
39. Y. K. Gaihare, S. M. M. Islam, and U. Singh, "Chapter 15-Managing Rice Soils for Mitigating Greenhouse Gases Emissions," *Agriculture Toward Net Zero Emissions* (2025): 305–326.
40. S. Hussain, J. Huang, J. Huang, et al., "Rice Production Under Climate Change: Adaptations and Mitigating Strategies," in *Environment, Climate, Plant and Vegetation Growth* (Springer, 2020), 659–686, https://doi.org/10.1007/978-3-030-49732-3_26.
41. FAO, "Methane Emissions in Livestock and Rice Systems," *Sources, Quantification, Mitigation and Metrics* (2023), <https://doi.org/10.4060/cc7607en>.
42. S. M. M. Islam, Y. K. Gaihare, M. R. Islam, et al., "Effects of Water Management on Greenhouse Gas Emissions From Farmers' Rice Fields in Bangladesh," *Science of the Total Environment* 734 (2020): 139382, <https://doi.org/10.1016/j.scitotenv.2020.139382>.
43. M. A. Habib, S. M. M. Islam, M. A. Haque, et al., "Effects of Irrigation Regimes and Rice Varieties on Methane Emissions and Yield of Dry Season Rice in Bangladesh," *Soil Systems* 7, no. 2 (2023): 41, <https://doi.org/10.3390/soilsystems7020041>.
44. F. Chen, L. Wang, Y. Wang, et al., "Retrieval of Dominant Methane (CH₄) Emission Sources, the First High-Resolution (1–2 m) Dataset of Storage Tanks of China in 2000–2021," *Earth System Science Data* 16, no. 7 (2024): 3369–3382, <https://doi.org/10.5194/essd-16-3369-2024>.
45. Y. Zhao, M. Saunio, P. Bousquet, et al., "Reconciling the Bottom-Up and Top-Down Estimates of the Methane Chemical Sink Using Multiple Observations," *Atmospheric Chemistry and Physics* 23, no. 1 (2023): 789–807, <https://doi.org/10.5194/acp-23-789-2023>.
46. G. Wu, X. M. Chen, J. Ling, et al., "Effects of Soil Warming and Increased Precipitation On greenhouse Gas Fluxes in Spring Maize Seasons in the North China Plain," *Science of the Total Environment* 734 (2020): 139269, <https://doi.org/10.1016/j.scitotenv.2020.139269>.
47. IPCC, in *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A: Global and Sectoral Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, ed. C. B. Field, V. R. Barros, D. J. Dokken, et al. (Cambridge University Press, 2014), 1132.
48. FAOSTAT, *FAOSTAT Statistical Database* (FAO (Food and Agriculture Organization of the United Nations), 2016).
49. NOAA, in *Earth System Research Laboratory* (NOAA) (2010), <https://www.esrl.noaa.gov>.
50. NOAA, in *Earth System Research Laboratory* (NOAA) (2020), <https://www.esrl.noaa.gov>.
51. M. M. Hanafiah, A. J. Ibraheem, and K. K. Razman, "Emissions of Carbon Dioxide and Methane From Dairy Cattle Manure," *IOP Conference Series: Earth and Environmental Science* 880, no. 1 (2021): 012037, <https://doi.org/10.1088/1755-1315/880/1/012037>.
52. M. H. M. El, T. J. Barzee, R. B. Franco, R. Zhang, S. Kaffka, and F. Mitloehner, "Anaerobic Digestion and Alternative Manure Management Technologies for Methane Emissions Mitigation on Californian Dairies," *Atmosphere* 14, no. 1 (2023): 120, <https://doi.org/10.3390/atmos14010120>.
53. K. Anderson, *What is Nitrous Oxide (N₂O)?* (2025), <https://greenly.earth/en-gb/blog/ecology-news/what-is-nitrous-oxide-n2o>.
54. P. Smith, D. Martino, Z. Cai, et al., eds., *Climate Change 2007: Mitigation. Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* (Cambridge University Press, 2007).
55. H. O. P. Mbow, A. Reisinger, J. Canadell, and P. O'Brien, in *Special Report on Climate Change, Desertification, Land Degradation, Sustainable Land Management, Food Security, and Greenhouse Gas Fluxes in Terrestrial Ecosystems (SR2)* (IPCC, 2017).
56. A. Charles, P. Rochette, J. K. Whalen, D. A. Angers, M. H. Chantigny, and N. Bertrand, "Global Nitrous Oxide Emission Factors From Agricultural Soils After Addition of Organic Amendments: A Meta-Analysis," *Agriculture, Ecosystems & Environment* 236 (2017): 88–98, <https://doi.org/10.1016/j.agee.2016.11.021>.
57. R. W. Portmann, J. S. Daniel, and A. R. Ravishankara, "Stratospheric Ozone Depletion Due to Nitrous Oxide: Influences of Other Gases," *Philosophical Transactions of the Royal Society B: Biological Sciences* 367, no. 1593 (2012): 1256–1264, <https://doi.org/10.1098/rstb.2011.0377>.
58. J. Hatfield, G. Takle, R. Grotjahn, et al., "Ch. 6: Agriculture," *Climate Change Impacts in the United States: The Third National Climate Assessment*, ed. J. M. Melillo, T. C. Richmond, and G. W. Yohe (U.S. Global Change Research Program, 2014), 150–174, <https://doi.org/10.7930/J02213FR>.
59. G. S. Malhi, M. Kaur, and P. Kaushik, "Impact of Climate Change on Agriculture and Its Mitigation Strategies: A Review," *Sustainability* 13, no. 3 (2021): 1318, <https://doi.org/10.3390/su13031318>.
60. S. S. Myers, A. Zanobetti, I. Kloog, et al., "Increasing CO₂ Threatens Human Nutrition," *Nature* 510, no. 7503 (2014): 139–142, <https://doi.org/10.1038/nature13179>.
61. S. Mathur, D. Agrawal, and A. Jajoo, "Photosynthesis: Response to High Temperature Stress," *Journal of Photochemistry and Photobiology B: Biology* 137 (2014): 116–212, <https://doi.org/10.1016/j.jphotobiol.2014.01.010>.
62. R. G. Jensen, "Activation of Rubisco Regulates Photosynthesis at High Temperature and CO₂," *Proceedings of the National Academy of Sciences of the United States of America* 97, no. 24 (2000): 12937–12938, <https://doi.org/10.1073/pnas.97.24.12937>.

63. H. Zhu, Y. Wu, and Y. Zheng, "Effects of Heat Shock on Photosynthesis-Related Characteristics and Lipid Profile of *Cycas multipinnata* and *C. panzhihuaensis*," *BMC Plant Biology* 22, no. 1 (2022): 442, <https://doi.org/10.1186/s12870-022-03825-0>.
64. M. Hasanuzzaman, K. Nahar, M. M. Alam, R. Roychowdhury, and M. Fujita, "Physiological, Biochemical, and Molecular Mechanisms of Heat Stress Tolerance in Plants," *International Journal of Molecular Sciences* 14, no. 5 (2013): 9643–9684, <https://doi.org/10.3390/ijms14059643>.
65. M. Iovane, A. Cirillo, L. G. Izzo, C. Di Vaio, and G. Aronne, "High Temperature and Humidity Affect Pollen Viability and Longevity in *Olea europaea* L.," *Agronomy* 12, no. 1 (2022): 1, <https://doi.org/10.3390/agronomy12010001>.
66. N. V. Khodorova and M. Boitel-Conti, "The Role of Temperature in the Growth and Flowering of Geophytes," *Plants* 2, no. 4 (2013): 699–771, <https://doi.org/10.3390/plants2040699>.
67. G. P. Cudjoe, P. Antwi-Agyei, and B. A. Gyampoh, "The Effect of Climate Variability on Maize Production in the Ejura-Sekyedumase Municipality, Ghana," *Climate* 9, no. 10 (2021): 145, <https://doi.org/10.3390/cli9100145>.
68. K. Yamauchi, "Climate Change Impacts on Agriculture and Irrigation in the Lower Mekong Basin," *Paddy and Water Environment* 12, no. S2 (2014): S227–S240, <https://doi.org/10.1007/s10333-013-0388-9>.
69. R. Wassmann, S. V. K. Jagadish, K. Sumfleth, et al., "Regional Vulnerability of Climate Change Impacts on Asian Rice Production and Scope for Adaptation," *Advances in Agronomy* 102 (2009): 91–133, [https://doi.org/10.1016/S0065-2113\(09\)01003-7](https://doi.org/10.1016/S0065-2113(09)01003-7).
70. J. I. Lizaso, M. Ruiz-Ramos, L. Rodríguez, et al., "Impact of High Temperatures in Maize: Phenology and Yield Components," *Field Crops Research* 216 (2018): 129–140, <https://doi.org/10.1016/j.fcr.2017.11.013>.
71. W. Shi and F. Tao, "Vulnerability of African Maize Yield to Climate Change and Variability During 1961–2010," *Food Security* 6, no. 4 (2014): 471–481, <https://doi.org/10.1007/s12571-014-0370-4>.
72. C. Rosenzweig and F. N. Tubiello, "Adaptation and Mitigation Strategies in Agriculture: An Analysis of Potential Synergies," *Mitigation and Adaptation Strategies for Global Change* 12, no. 5 (2007): 855–873, <https://doi.org/10.1007/s11027-007-9103-8>.
73. V. Potopová, P. Zahradníček, L. Türkott, P. Štěpánek, and J. Soukup, "The Effects of Climate Change on Variability of the Growing Seasons in the Elbe River Lowland, Czech Republic," *Land-Atmosphere Interactions* 2015 (2015): 546920, <https://doi.org/10.1155/2015/546920>.
74. J. E. Olesen and M. Bindi, "Consequences of Climate Change for European Agricultural Productivity, Land Use and Policy," *European Journal of Agronomy* 16, no. 4 (2002): 239–262, [https://doi.org/10.1016/S1161-0301\(02\)00004-7](https://doi.org/10.1016/S1161-0301(02)00004-7).
75. S. A. Zayan, "Impact of Climate Change on Plant Diseases and IPM Strategies," in *Plant Pathology and Management of Plant Diseases*, ed. S. Topolovec-Pintarić (IntechOpen, 2019), <https://doi.org/10.5772/intechopen.87055>.
76. J. O. Ajetomobi, "Sensitivity of Crop Yield to Extreme Weather in Nigeria," in *Paper Presented at the 5th Conference of African Association Agricultural Economists (AAAE), Held in Addis Ababa, Ethiopia (September 23-26, 2016)* (2016), <http://ageconsearch.umn.edu/bitstream/246919/2/259.%20Sensitivity%20of%20crop%20yield%20to%20extreme%20weather%20in%20Nigeria.pdf>.
77. C. Pendino, "Biome Breakdown: The Effects of Climate Change on Agriculture in Nigeria and Thailand," *Global Majority E-Journal* 8, no. 1 (2017): 4–17.
78. A. Prakash, J. Rao, A. K. Mukherjee, et al., in *Climate Change: Impact on Crop Pests; Applied Zoologists Research Association (AZRA)* (Central Rice Research Institute, 2014).
79. B. B. Fand, A. L. Kamble, and M. Kumar, "Will Climate Change Pose Serious Threat to Crop Pest Management: A Critical Review," *International Journal of Science and Research* 2 (2012): 1–14.
80. FAO, in *Climate Related Transboundary Pests and Diseases* (2020), <http://www.fao.org/3/a-ai785e.pdf>.
81. S. P. Cohen and J. E. Leach, "High Temperature-Induced Plant Disease Susceptibility: More Than the Sum of Its Parts," *Current Opinion in Plant Biology* 56 (2020): 235–241, <https://doi.org/10.1016/j.pbi.2020.02.008>.
82. A. C. Velasquez, C. D. M. Castroverde, and S. Y. He, "Plant-Pathogen Warfare Under Changing Climate Conditions," *Current Biology* 28, no. 10 (2018): R619–R634, <https://doi.org/10.1016/j.cub.2018.03.054>.
83. M. R. Khan and T. F. Rizvi, "Effect of Elevated Levels of CO₂ on Powdery Mildew Development in Five Cucurbit Species," *Scientific Reports* 10, no. 1 (2020): 4986, <https://doi.org/10.1038/s41598-020-61790-w>.
84. Z. Vary, E. Mullins, J. C. McElwain, and F. M. Doohan, "The Severity of Wheat Diseases Increases When Plants and Pathogens are Acclimated to Elevated Carbon Dioxide," *Global Change Biology* 21, no. 7 (2015): 2661–2669, <https://doi.org/10.1111/gcb.12899>.
85. B. K. Singh, M. Delgado-Baquerizo, E. Egidi, J. E. Leach, H. Liu, and P. Trivedi, "Climate Change Impacts on Plant Pathogens, Food Security and Paths Forward," *Nature Reviews Microbiology* 21, no. 10 (2023): 640–656, <https://doi.org/10.1038/s41579-023-00900-7>.
86. G. Howard, R. Calow, A. Macdonald, and J. Bartram, "Climate Change and Water and Sanitation: Likely Impacts and Emerging Trends for Action," *Annual Review of Environment and Resources* 41, no. 1 (2016): 253–276, <https://doi.org/10.1146/annurev-enviro-110615-085856>.
87. S. Sukanya and S. Joseph, "Climate Change Impacts on Water Resources: An Overview," *Visualization Techniques for Climate Change With Machine Learning and Artificial Intelligence* 2023 (2023): 55–76, <https://doi.org/10.1016/B978-0-323-99714-0.00008-X>.
88. E. A. Davidson and I. A. Janssens, "Temperature Sensitivity of Soil Carbon Decomposition and Feedbacks to Climate Change," *Nature* 440, no. 7081 (2006): 65–173, <https://doi.org/10.1038/nature04514>.
89. D. K. Benbi and R. Kaur, "Modeling Soil Processes in Relation to Climate Change," *Journal of the Indian Society of Soil Science* 57 (2009): 433–444.
90. S. Mondal, "Impact of Climate Change on Soil Fertility," *Climate Change and the Microbiome*, ed. D. K. Choudhary, 63 (2021), 551–569, https://doi.org/10.1007/978-3-030-76863-8_28.
91. A. O. Adekiya, B. B. Ayorinde, E. T. Alori, C. Aremu, and W. S. Ejue, "Effects of Lime on Soil Chemical Characteristics and Performance of Cowpea (*Vigna unguiculata* (L.) Walp.) on Oxidic Haplustalf of a Derived Savanna Ecology of Nigeria," *Research on Crops* 24, no. 2 (2023): 298–306, <https://doi.org/10.31830/2348-7542.2023.ROC-885>.
92. B. P. Singh, A. L. Cowie, and K. Y. Chan, eds., *Soil Health and Climate Change, Soil Biology* (Springer, 2011), 414, https://doi.org/10.1007/978-3-642-20256-8_4.
93. N. E. Ebido, I. G. Edeh, B. O. Unagwu, et al., "Rice-Husk Biochar Effects on Organic Carbon, Aggregate Stability and Nitrogen-Fertility of Coarse-textured Ultisols Evaluated Using *Celosia argentea* Growth," *SAINS TANAH-Journal of Soil Science and Agroclimatology* 18, no. 2 (2021): 177–187, <https://doi.org/10.20961/stjssa.v18i2.56330>.
94. Q. Sun, J. Meng, Y. Lan, et al., "Long-Term Effects of Biochar Amendment on Soil Aggregate Stability and Biological Binding Agents in Brown Earth," *Catena* 205 (2021): 105460, <https://doi.org/10.1016/j.catena.2021.105460>.
95. Z. Sun, Z. Zhang, K. Zhu, et al., "Biochar Altered Native Soil Organic Carbon by Changing Soil Aggregate Size Distribution and Native SOC in Aggregates Based on an 8-Year Field Experiment," *Science of the Total Environment* 708 (2020): 134829, <https://doi.org/10.1016/j.scitotenv.2019.134829>.

96. Y. Li, G. Feng, and H. Tewolde, "Biochar Derived From Papermill Factories Improves Soil Physical and Hydraulic Properties in No-Till Cotton Fields," *Biochar* 5, no. 1 (2023): 35, <https://doi.org/10.1007/s42773-023-00235-9>.
97. I. Edeh, O. Masek, and W. Buss, "A Meta-Analysis on Biochar's Effects on Soil Water Properties—New Insights and Future Research Challenges," *The Science of the Total Environment* 714, no. 202–203 (2020): 136857, <https://doi.org/10.1016/j.scitotenv.2020.136857>.
98. S. Ahmad Bhat, A. Kuriqi, M. U. D. Dar, et al., "Application of Biochar for Improving Physical, Chemical, and Hydrological Soil Properties: A Systematic Review," *Sustainability* 14, no. 17 (2022): 11104, <https://doi.org/10.3390/su141711104>.
99. E. Arthur and F. Ahmed, "Rice Straw Biochar Affects Water Retention and Air Movement in a Sand-Textured Tropical Soil," *Archives of Agronomy and Soil Science* 63, no. 14 (2017): 2035–2047, <https://doi.org/10.1080/03650340.2017.1322196>.
100. J. Lehmann and S. Joseph, *Biochar for Environmental Management: Science, Technology and Implementation* (Routledge, 2015).
101. S. E. Obalum, N. E. Ebido, J. C. Akubue, et al., "Contributions of Soil Hydrothermal Regime to Biochar Effectiveness: Chronicling Empirical Evidence Among Nigerian Agroecologies," in *Glimpse into the Frontiers of Research in Soil Science in Nigeria*, ed. N. Abdu (2024), 174–205.
102. P. G. Oguntunde, M. Fosu, A. E. Ajayi, and N. Van De Giesen, "Effects of Charcoal Production on Maize Yield, Chemical Properties and Texture of Soil," *Biology and Fertility of Soils* 39, no. 4 (2004): 295–299, <https://doi.org/10.1007/s00374-003-0707-1>.
103. B. Glaser, J. Lehmann, and W. Zech, "Ameliorating Physical and Chemical Properties of Highly Weathered Soils in the Tropics With Charcoal—A Review," *Biology and Fertility of Soils* 35, no. 4 (2002): 219–230, <https://doi.org/10.1007/s00374-002-0466-4>.
104. A. El-Naggar, S. S. Lee, Y. M. Awad, et al., "Influence of Soil Properties and Feedstocks on Biochar Potential for Carbon Mineralization and Improvement of Infertile Soils," *Geoderma* 2018 332 (2002): 100–108, <https://doi.org/10.1016/j.geoderma.2018.06.017>.
105. J. Lehmann and M. Rondon, "Biochar Soil Management on Highly Weathered Soils in the Humid Tropics," in *Biological Approaches to Sustainable Soil Systems*, ed. N. Uphoff, A. S. Ball, E. Fernandes, et al. (CRC Press, 2006), 517–530.
106. D. Woolf, J. Lehmann, S. Ogle, A. W. Kishimoto-Mo, B. McConkey, and J. Baldock, "Greenhouse Gas Inventory Model for Biochar Additions to Soil," *Environmental Science and Technology* 55, no. 21 (2021): 14795–14805, <https://doi.org/10.1021/acs.est.1c02425>.
107. D. Laird, P. Fleming, B. Wang, R. Horton, and D. Karlen, "Biochar Impact on Nutrient Leaching From a Midwestern Agricultural Soil," *Geoderma* 158, no. 3–4 (2010): 436–442, <https://doi.org/10.1016/j.geoderma.2010.05.012>.
108. J. E. Thies and M. C. Rillig, "Characteristics of Biochar: Biological Properties," in *Biochar for Environmental Management*, ed. J. Lehmann and S. Joseph (Earthscan, 2009), 85–106.
109. M. Zhang, Y. Liu, Q. Wei, et al., "Effects of Biochar and Vermicompost on Growth and Economic Benefits of Continuous Cropping Pepper at Karst Yellow Soil Region in Southwest China," *Frontiers of Plant Science* 14 (2023): 1238663, <https://doi.org/10.3389/fpls.2023.1238663>.
110. M. Mierzwa-Hersztek, K. Wolny-Koładka, K. Gondek, A. Gałazka, and K. Gawryjolek, "Effect of Coapplication of Biochar and Nutrients on Microbiocenotic Composition, Dehydrogenase Activity Index and Chemical Properties of Sandy Soil," *Waste and Biomass Valorization* 11, no. 8 (2020): 3911–3923, <https://doi.org/10.1007/s12649-019-00757-z>.
111. T. M. Agbede, A. Oyewumi, A. O. Adekiya, et al., "Assessing the Synergistic Impacts of Poultry Manure and Biochar on Nutrient-Depleted Sand and Sandy Loam Soil Properties and Sweet Potato Growth and Yield," *Experimental Agriculture* 58, no. e54 (2022): 1–21, <https://doi.org/10.1017/S0014479722000497>.
112. W. Ashiq, M. Nadeem, W. Ali, et al., "Biochar Amendment Mitigates Greenhouse Gases Emission and Global Warming Potential in Dairy Manure-Based Silage Corn in Boreal Climate," *Environmental Pollution* 265 (2020): 114869, <https://doi.org/10.1016/j.envpol.2020.114869>.
113. W. Wang, X. Wu, A. Chen, X. Xie, Y. Wang, and C. Yin, "Mitigating Effects of Ex Situ Application of Rice Straw on CH₄ and N₂O Emissions From Paddy-Upland Coexisting System," *Scientific Reports* 6, no. 1 (2016): 37402, <https://doi.org/10.1038/srep37402>.
114. A. O. Adekiya, O. V. Adebiyi, A. L. Ibaba, C. Aremu, and R. O. Ajibade, "Effects of Wood Biochar and Potassium Fertilizer on Soil Properties, Growth and Yield of Sweet Potato (*Ipomea batata*)," *Heliyon* 8, no. 11 (2022): e11728, <https://doi.org/10.1016/j.heliyon.2022.e11728>.
115. T. Kongthod, S. Thanachit, S. Anusontpornperm, and W. Wiriyakitnatekul, "Effects of Biochars and Other Organic Soil Amendments on Plant Nutrient Availability in an Ustoxic Quartzipsamment," *Pedosphere* 25, no. 5 (2015): 790–798, [https://doi.org/10.1016/s1002-0160\(15\)30060-6](https://doi.org/10.1016/s1002-0160(15)30060-6).
116. Y. Lou, S. Joseph, L. Li, E. R. Graber, X. Liu, and G. Pan, "Water Extract From Straw Biochar Used for Plant Growth Promotion: An Initial Test," *Bioresources* 11, no. 1 (2016): 249–266, <https://doi.org/10.15376/biores.11.1.249-266>.
117. L. Montanarella and E. Lugato, "The Application of Biochar in the EU: Challenges and Opportunities," *Agronomy* 3, no. 2 (2013): 462–473, <https://doi.org/10.3390/agronomy3020462>.
118. D. Woolf, J. E. Amonette, F. A. Street-Perrott, J. Lehmann, and S. Joseph, "Sustainable Biochar to Mitigate Global Climate Change," *Nature Communications* 1, no. 1 (2010): 1–9, <https://doi.org/10.1038/ncomms1053>.
119. M. H. Duku, S. Gu, and E. B. Hagan, "Biochar Production Potential in Ghana—A Review," *Renewable and Sustainable Energy Reviews* 15, no. 8 (2011): 3539–3551, <https://doi.org/10.1016/j.rser.2011.05.010>.
120. S. T. Shafie, M. M. Salleh, L. L. Hang, M. Rahman, and W. A. W. A. K. Ghani, "Effect of Pyrolysis Temperature on the Biochar Nutrient and Water Retention Capacity," *Journal of Purity, Utility Reaction and Environment* 1, no. 6 (2012): 293–307.
121. S. Rangabhashiyam and P. Balasubramanian, "The Potential of Lignocellulosic Biomass Precursors for Biochar Production: Performance, Mechanism and Wastewater Application—A Review," *Industrial Crops and Products* 128 (2019): 405–423, <https://doi.org/10.1016/j.indcrop.2018.11.041>.
122. J. Xing, G. Xu, and G. Li, "Comparison of Pyrolysis Process, Various Fractions and Potential Soil Applications Between Sewage Sludge-Based Biochars and Lignocellulose-Based Biochars," *Ecotoxicology and Environmental Safety* 208 (2021): 111756, <https://doi.org/10.1016/j.ecoenv.2020.111756>.
123. Y. Kuzyakov, I. Bogomolova, and B. Glaser, "Biochar Stability in Soil: Decomposition During Eight Years and Transformation as Assessed by Compound-Specific ¹⁴C Analysis," *Soil Biology and Biochemistry* 70 (2014): 229–236, <https://doi.org/10.1016/j.soilbio.2013.12.021>.
124. R. Lal, "Soil Carbon Management and Climate Change," *Carbon Management* 4, no. 4 (2013): 439–462, <https://doi.org/10.4155/cmt.13.31>.
125. C. L. Phillips, K. M. Meyer, M. Garcia-Jaramillo, et al., "Towards Predicting Biochar Impacts on Plant-Available Soil Nitrogen Content," *Biochar* 4, no. 1 (2022): 9, <https://doi.org/10.1007/s42773-022-00137-2>.
126. E. S. Button, J. Pett-Ridge, D. V. Murphy, Y. Kuzyakov, D. R. Chadwick, and D. L. Jones, "Deep-C Storage: Biological, Chemical and Physical Strategies to Enhance Carbon Stocks in Agricultural Subsoils," *Soil Biology and Biochemistry* 170 (2022): 108697, <https://doi.org/10.1016/j.soilbio.2022.108697>.

127. S. Li and D. Tasnady, "Biochar for Soil Carbon Sequestration: Current Knowledge, Mechanisms, and Future Perspectives," *Chimia* 9, no. 3 (2023): 67, <https://doi.org/10.3390/c9030067>.
128. S. Mukherjee, B. Sarkar, V. K. Aralappanavar, et al., "Biochar-Microorganism Interactions for Organic Pollutant Remediation: Challenges and Perspectives," *Environmental Pollution* 308 (2022): 119609, <https://doi.org/10.1016/j.envpol.2022.119609>.
129. A. J. Eykelbosh, M. S. Johnson, and E. G. Couto, "Biochar Decreases Dissolved Organic Carbon But Not Nitrate Leaching in Relation to Vinasse Application in a Brazilian Sugarcane Soil," *Journal of Environmental Management* 149 (2015): 9–17, <https://doi.org/10.1016/j.jenvman.2014.09.033>.
130. Z. Sun, E. W. Bruun, E. Arthur, et al., "Effect of Biochar on Aerobic Processes, Enzyme Activity, and Crop Yields in Two Sandy Loam Soils," *Biology and Fertility of Soils* 50, no. 7 (2014): 1087–1097, <https://doi.org/10.1007/s00374-014-0928-5>.
131. Y. Qiao and C. Wu, "Nitrogen Enriched Biochar Used as CO₂ Adsorbents: A Brief Review," *Carbon Capture Science & Technology* 2 (2022): 100018, <https://doi.org/10.1016/j.ccst.2021.100018>.
132. X. Gui, X. Xu, Z. Zhang, et al., "Biochar-Amended Soil Can Further Sorb Atmospheric CO₂ for More Carbon Sequestration," *Communications Earth & Environment* 6, no. 1 (2025): 5, <https://doi.org/10.1038/s43247-024-01985-5>.
133. J. Lehmann, A. Cowie, C. A. Masiello, et al., "Biochar in Climate Change Mitigation," *Nature Geoscience* 14, no. 12 (2021): 883–892, <https://doi.org/10.1038/s41561-021-00852-8>.
134. S. Joseph, A. L. Cowie, B. N. Zwieterman, et al., "How Biochar Works, and When It Doesn't: A Review of Mechanisms Controlling Soil and Plant Responses to Biochar," *GCB Bioenergy* 13, no. 11 (2021): 1731–1764, <https://doi.org/10.1111/gcbb.12885>.
135. Y. Q. Sheng, Y. Zhan, and L. Zhu, "Reduced Carbon Sequestration Potential of Biochar in Acidic Soil," *Science of the Total Environment* 572 (2016): 129–137, <https://doi.org/10.1016/j.scitotenv.2016.07.140>.
136. D. Lefebvre, S. Fawzy, C. A. Aquije, et al., "Biomass Residue to Carbon Dioxide Removal: Quantifying the Global Impact of Biochar," *Biochar* 5, no. 1 (2023): 65, <https://doi.org/10.1007/s42773-023-00258-2>.
137. A. Zhang, Y. Liu, G. Pan, et al., "Effect of Biochar Amendment on Maize Yield and Greenhouse Gas Emissions From a Soil Organic Carbon Poor Calcareous Loamy Soil From Central China Plain," *Plant and Soil* 351, no. 1 (2012): 263–275, <https://doi.org/10.1007/s11104-011-0957-x>.
138. I. Hawthorne, M. S. Johnson, R. S. Jassal, T. A. Black, N. J. Grant, and S. M. Smukler, "Application of Biochar and Nitrogen Influences Fluxes of CO₂, CH₄ and N₂O in a Forest Soil," *Journal of Environmental Management* 1, no. 192 (2017): 203–214, <https://doi.org/10.1016/j.jenvman.2016.12.066>.
139. R. B. Fidel, D. A. Laird, and T. B. Parkin, "Effect of Biochar on Soil Greenhouse Gas Emissions at the Laboratory and Field Scales," *Soil Systems* 3, no. 1 (2019): 8, <https://doi.org/10.3390/soilsystems3010008>.
140. C. Scheer, P. R. Grace, D. W. Rowlings, S. Kimber, and L. Van Zwieterman, "Effect of Biochar Amendment on the Soil-Atmosphere Exchange of Greenhouse Gases From an Intensive Subtropical Pasture in Northern New South Wales, Australia," *Plant and Soil* 345, no. 1–2 (2011): 47–58, <https://doi.org/10.1007/s11104-011-0759-1>.
141. X. Y. Liu, L. M. Wang, and S. W. Zhu, "Urban Sprawl and Residential Carbon Emission: Panel Data on Southern Cities in China," *Journal of Southeast University (Philosophy and Social Science)* 18, no. 05 (2016): 101–108+148, <https://doi.org/10.13916/j.cnki.issn1671-511x.2016.05.014>.
142. D. Wu, M. Senbayram, H. Zang, et al., "Effect of Biochar Origin and Soil pH on Greenhouse Gas Emissions From Sandy and Clay Soils," *Applied Soil Ecology* 129 (2018): 121–127, <https://doi.org/10.1016/j.apsoil.2018.05.009>.
143. A. E. E. A. Z. Amin, "Carbon Sequestration, Kinetics of Ammonia Volatilization and Nutrient Availability in Alkaline Sandy Soil as a Function on Applying Calotropis Biochar Produced at Different Pyrolysis Temperatures," *Science of the Total Environment* 726 (2020): 138489, <https://doi.org/10.1016/j.scitotenv.2020.138489>.
144. L. Wang, C. Gao, K. Yang, et al., "Effects of Biochar Aging in the Soil on Its Mechanical Property and Performance for Soil CO₂ and N₂O Emissions," *Science of the Total Environment* 782 (2021): 146824, <https://doi.org/10.1016/j.scitotenv.2021.146824>.
145. P. Cely, G. Gascó, J. Paz-Ferreiro, and A. Méndez, "Agronomic Properties of Biochars From Different Manure Wastes," *Journal of Analytical and Applied Pyrolysis* 111 (2015): 173–182.
146. H. Singh, B. K. Northup, C. W. Rice, and P. V. Varasdas, "Biochar Applications Influence Soil Physical and Chemical Properties, Microbial Diversity, and Crop Productivity: A Meta-Analysis," *Biochar* 4, no. 1 (2022): 8, <https://doi.org/10.1007/s42773-022-00138-1>.
147. Y. Yanai, K. Toyota, and M. Okazaki, "Effects of Charcoal Addition on N₂O Emissions From Soil Resulting From Rewetting Air Dried Soil in Short Term Laboratory Experiments," *Soil Science & Plant Nutrition* 53, no. 2 (2007): 181–188, <https://doi.org/10.1111/j.1747-0765.2007.00123.x>.
148. A. O. Adekiya, S. O. Ojieniyi, and T. M. Agbede, "Soil Physical and Chemical Properties and Cocoyam Yield Under Different Tillage Systems in Tropical Alfisol," *Experimental Agriculture* 47, no. 3 (2011): 477–488, <https://doi.org/10.1017/s001447971100041x>.
149. V. Šimanský, E. Tobiašová, and J. Chlupík, "Soil Till-Age and Fertilization of Orthic Luvisol and Their Influence on Chemical Properties, Soil Structure Stability and Carbon Distribution in Water-Stable Macro-Aggregates," *Soil and Tillage Research* 100 (2008): 125–132.
150. Y. M. Zhang, J. Chen, L. Wang, X. Q. Wang, and Z. H. Gu, "The Spatial Distribution Patterns of Biological Soil Crusts in the Gurbantunggut Desert, Northern Xinjiang, China," *Journal of Arid Environments* 68, no. 4 (2007): 599–610, <https://doi.org/10.1016/j.jaridenv.2006.06.012>.
151. J. J. Almaraz, F. Mabood, X. Zhou, I. Strachan, B. Ma, and D. L. Smith, "Performance of Agricultural Systems Under Contrasting Growing Season Conditions in South-Western Quebec," *Journal of Agronomy and Crop Science* 195, no. 5 (2009): 319–327, <https://doi.org/10.1111/j.1439-037x.2009.00369.x>.
152. W. Hu, J. Drewry, M. Beare, A. Eger, and K. Muller, "Compaction Induced Soil Structural Degradation Affects Productivity and Environmental Outcomes: A Review and New Zealand Case Study," *Geoderma* 395 (2021): 115035, <https://doi.org/10.1016/j.geoderma.2021.115035>.
153. R. Ruser, H. Flessa, R. Schilling, H. Steindl, and F. Beese, "Soil Compaction and Fertilization Effects on Nitrous Oxide and Methane Fluxes in Potato Fields," *Soil Science Society of America Journal* 62, no. 6 (1998): 1587–1595, <https://doi.org/10.2136/sssaj1998.03615995006200060016x>.
154. M. Simek, P. Brucek, J. Hynst, E. Uhlirova, and S. O. Petersen, "Effects of Excretal Returns and Soil Compaction on Nitrous Oxide Emissions From a Cattle Overwintering Area," *Agriculture, Ecosystems & Environment* 112 (2006): 186–191, <https://doi.org/10.1016/j.agee.2005.08.018>.
155. B. Ball, K. Cameron, H. Di, and S. Moore, "Effects of Trampling of a Wet Dairy Pasture Soil on Soil Porosity and on Mitigation of Nitrous Oxide Emissions by a Nitrification Inhibitor, Dicyandiamide," *Soil Use & Management* 28, no. 2 (2012): 194–201, <https://doi.org/10.1111/j.1475-9772.2012.00389.x>.
156. T. J. van der Weerden and T. M. Styles, "Reducing Nitrous Oxide Emissions From Grazed Winter Forage Crops," *Proceedings of the New Zealand Grassland Association* 74 (2012): 57–62, <https://doi.org/10.33584/jnzg.2012.74.2884>.
157. T. J. van der Weerden, T. M. Styles, A. J. Rutherford, C. A. M. de Klein, and R. Dynes, "Nitrous Oxide Emissions From Cattle Urine Deposited onto Soil Supporting a Winter Forage Kale Crop,"

- New Zealand Journal of Agricultural Research* 60, no. 2 (2017): 119–130, <https://doi.org/10.1080/00288233.2016.1273838>.
158. N. Goutal, P. Renault, and J. Ranger, “Forwarder Traffic Impacted Over at Least Four Years Soil Air Composition of Two Forest Soils in Northeast France,” *Geoderma* 193 (2013): 29–40, <https://doi.org/10.1016/j.geoderma.2012.10.012>.
 159. J. Dörner and R. Horn, “Anisotropy of Pore Functions in Structured Stagnic Luvisols in the Weichselian Moraine Region in N Germany,” *Journal of Plant Nutrition and Soil Science* 169 (2006): 213–220, <https://doi.org/10.1002/jpln.200521844>.
 160. H. Kim, S. Anderson, P. Motavalli, and C. Gantzer, “Compaction Effects on Soil Macropore Geometry and Related Parameters for an Arable Field,” *Geoderma* 160, no. 2 (2010): 244–251, <https://doi.org/10.1016/j.geoderma.2010.09.030>.
 161. F. E. Berisso, P. Schjønning, T. Keller, et al., “Persistent Effects of Subsoil Compaction on Pore Size Distribution and Gas Transport in Loamy Soil,” *Soil and Tillage Research* 122 (2012): 45–51.
 162. R. Ruser, H. Flessa, R. Russow, G. Schmidt, F. Buegger, and J. C. Munch, “Emission of N₂O, N₂ and CO₂ From Soil Fertilized With Nitrate: Effect of Compaction, Soil Moisture and Rewetting,” *Soil Biology and Biochemistry* 38, no. 2 (2006): 263–274, <https://doi.org/10.1016/j.soilbio.2005.05.005>.
 163. W. C. T. Chamen, A. P. Moxey, W. Towers, B. Balana, and P. D. Hallett, “Mitigating Arable Soil Compaction: A Review and Analysis of Available Cost and Benefit Data,” *Soil and Tillage Research* 146 (2015): 10–25, <https://doi.org/10.1016/j.still.2014.09.011>.
 164. B. Ball, “Soil Structure and Greenhouse Gas Emissions: A Synthesis of 20 Years of Experimentation,” *European Journal of Soil Science* 64, no. 3 (2013): 357–373, <https://doi.org/10.1111/ejss.12013>.
 165. M. Rondon, J. A. Ramirez, and J. Lehmann, “Greenhouse Gas Emissions Decrease With Charcoal Additions to Tropical Soils,” in *3rd USDA Symposium on Greenhouse Gases and Carbon Sequestration in Agriculture and Forestry* (2005).
 166. L. Liu, G. Shen, M. Sun, X. Cao, G. Shang, and P. Chen, “Effect of Biochar on Nitrous Oxide Emission and Its Potential Mechanisms,” *Journal of the Air and Waste Management Association* 64, no. 8 (2014): 894–902, <https://doi.org/10.1080/10962247.2014.899937>.
 167. F. Zhang, J. Qi, F. M. Li, C. S. Li, and C. B. Li, “Quantifying Nitrous Oxide Emissions From Chinese Grasslands With a Process-Based Model,” *Biogeosciences* 7, no. 6 (2010): 2039–2050, <https://doi.org/10.5194/bg-7-2039-2010>.
 168. D. V. Sarkhot, A. A. Berhe, and T. A. Ghezzehei, “Impact of Biochar Enriched With Dairy Manure Effluent on Carbon and Nitrogen Dynamics,” *Journal of Environmental Quality* 41, no. 4 (2012): 1107–1114, <https://doi.org/10.2134/jeq2011.0123>.
 169. M. L. Cayuela, Z. L. Van, B. P. Singh, S. Jeffery, A. Roig, and M. A. Sánchez-Monedero, “Biochar’s Role in Mitigating Soil Nitrous Oxide Emissions: A Review and Meta-Analysis,” *Agriculture, Ecosystems & Environment* 191 (2014): 5–16.
 170. Y. Chang, L. Rossi, L. Zotarelli, B. Gao, M. A. Shahid, and A. Sarkhosh, “Biochar Improves Soil Physical Characteristics and Strengthens Root Architecture in Muscadine Grape (*Vitis rotundifolia* L.),” *Chemical and Biological Technologies in Agriculture* 8, no. 1 (2021): 7, <https://doi.org/10.1186/s40538-020-00204-5>.
 171. F. Sønderholm and C. J. Bjerrum, “Minimum Levels of Atmospheric Oxygen From Fossil Tree Roots Imply New Plant–Oxygen Feedback,” *Geobiology* 19 (2021): 250–260, <https://doi.org/10.1111/gbi.12435>.
 172. E. J. Bateman and E. M. Baggs, “Contributions of Nitrification and Denitrification to N₂O Emissions From Soils at Different Water-Filled Pore Space,” *Biology and Fertility of Soils* 41, no. 6 (2005): 379–388, <https://doi.org/10.1007/s00374-005-0858-3>.
 173. A. Mukherjee, A. R. Zimmerman, and W. Harris, “Surface Chemistry Variations Among a Series of Laboratory-Produced Biochars,” *Geoderma* 163, no. 3–4 (2011): 247–255, <https://doi.org/10.1016/j.geoderma.2011.04.021>.
 174. K. Jindo, H. Mizumoto, Y. Sawada, M. A. Sanchez-Monedero, and T. Sonoki, “Physical and Chemical Characterization of Biochars Derived From Different Agricultural Residues,” *Biogeosciences* 11, no. 23 (2014): 6613–6621, <https://doi.org/10.5194/bg-11-6613-2014>.
 175. T. J. Clough, L. M. Condron, C. Kammann, and C. Müller, “A Review of Biochar and so Nitrogen Dynamics,” *Agronomy* 3, no. 2 (2013): 275–293, <https://doi.org/10.3390/agronomy3020275>.
 176. M. U. Hassan, M. Aamer, A. Mahmood, et al., “Management Strategies to Mitigate N₂O Emissions in Agriculture,” *Life* 12, no. 3 (2022): 439, <https://doi.org/10.3390/life12030439>.
 177. L. Zhong, G. Li, J. Qing, et al., “Biochar Can Reduce N₂O Production Potential From Rhizosphere of Fertilized Agricultural Soils by Suppressing Bacterial Denitrification,” *European Journal of Soil Biology* 109 (2022): 103391, <https://doi.org/10.1016/j.ejsobi.2022.103391>.
 178. S. D. C. Case, N. P. McNamara, D. S. Reay, A. W. Stott, H. K. Grant, and J. Whitaker, “Biochar Suppresses N₂O Emissions While Maintaining N Availability in a Sandy Loam Soil,” *Soil Biology and Biochemistry* 81 (2015): 178–185, <https://doi.org/10.1016/j.soilbio.2014.11.012>.
 179. X. Wang, B. Xiang, J. Li, et al., “Using Adaptive and Aggressive N₂O-Reducing Bacteria to Augment Digestate Fertilizer for Mitigating N₂O Emissions From Agricultural Soils,” *Science of the Total Environment* 903 (2023): 166284, <https://doi.org/10.1016/j.scitotenv.2023.166284>.
 180. J. A. Ippolito, M. E. Stromberger, R. D. Lentz, and R. S. Dungan, “Hardwood Biochar Influences Calcareous Soil Physicochemical and Microbiological Status,” *Journal of Environmental Quality* 43, no. 2 (2014): 681–689, <https://doi.org/10.2134/jeq2013.08.0324>.
 181. R.-J. Yao, H.-Q. Li, J.-S. Yang, X.-P. Wang, W.-P. Xie, and X. Zhang, “Biochar Addition Inhibits Nitrification by Shifting Community Structure of Ammonia-Oxidizing Microorganisms in Salt-Affected Irrigation-Silting Soil,” *Microorganisms* 10, no. 2 (2022): 436, <https://doi.org/10.3390/microorganisms10020436>.
 182. A. Obia, G. Cornelissen, J. Mulder, and P. Dörsch, “Effect of Soil Ph Increase by Biochar on NO, N₂O and N₂ Production During Denitrification in Acid Soils,” *PLoS One* 10, no. 9 (2015): e0138781, <https://doi.org/10.1371/journal.pone.0138781>.
 183. F. Lin, H. Wang, H. Shaghaleh, et al., “Effects of Biochar Amendment on N₂O Emissions From Soils With Different pH Levels,” *Atmosphere* 15, no. 1 (2024): 68, <https://doi.org/10.3390/atmos15010068>.
 184. L. Yu, J. Tang, R. Zhang, Q. Wu, and M. Gong, “Effects of Biochar Application on Soil Methane Emission at Different Soil Moisture Levels,” *Biology and Fertility of Soils* 49, no. 2 (2012): 119–128, <https://doi.org/10.1007/s00374-012-0703-4>.
 185. Y. H. Xiao, *Effects of Different Application Rates of Biochar on the Soil Greenhouse Gas Emission in Chinese Chestnut Stands* (Zhejiang A and F University, 2016), Master Thesis.
 186. E. Anders, A. Watzinger, F. Rempt, et al., “Biochar Affects the Structure Rather Than the Total Biomass of Microbial Communities in Temperate Soils,” *Agricultural and Food Science* 22, no. 4 (2013): 404–423, <https://doi.org/10.23986/afsci.8095>.
 187. S. Jeffery, D. Abalos, M. Prodana, et al., “Biochar Boosts Tropical But Not Temperate Crop Yields,” *Environmental Research Letters* 12, no. 5 (2017): 053001, <https://doi.org/10.1088/1748-9326/aa67bd>.
 188. P. Brassard, G. Stephane, and R. Vijaya, “Soil Biochar Amendment as a Climate Change Mitigation Tool: Key Parameters and Mechanisms Involved,” *Journal of Environmental Management* 181 (2016): 484–497, <https://doi.org/10.1016/j.jenvman.2016.06.063>.
 189. K. Karhu, K. Mattila, I. Bergström, and K. Lång, “Biochar Addition to Agricultural Soil Increased CH₄ Uptake and Water Holding Capacity–

- Results From a Short-Term Pilot Field Study," *Agriculture, Ecosystems & Environment* 140, no. 1 (2011): 309–313, <https://doi.org/10.1016/j.agee.2010.12.005>.
190. K. A. Spokas, "Impact of Biochar Field Aging on Laboratory Greenhouse Gas Production Potentials," *GCB Bioenergy* 5, no. 2 (2013): 165–176, <https://doi.org/10.1111/gcbb.12005>.
191. J.-M. Lee, H.-C. Jeong, H.-S. Gwon, et al., "Effects of Biochar on Methane Emissions and Crop Yields in East Asian Paddy Fields: A Regional Scale Meta-Analysis," *Sustainability* 15, no. 12 (2023): 9200, <https://doi.org/10.3390/su15129200>.
192. K. Rajalekshmi, B. Bastin, and S. Sasidharan, "Impact of Rice Husk Biochar on Soil Carbon Sequestration, Methane Emission, and Rice Yield in Wetland Soil (Ultisol)," in *Reducing Carbon Footprint in Different Sectors for Sustainability* (Intechopen, 2024), <https://doi.org/10.5772/intechopen.1005103>.
193. P. Sriphiroom, A. Chidthaisong, K. Yagi, S. Tripetchkul, N. Boonapatcharoen, and S. Towprayoon, "Effects of Biochar on Methane Emission, Grain Yield, and Soil in Rice Cultivation in Thailand," *Carbon Management* 12, no. 2 (2021): 109–121, <https://doi.org/10.1080/17583004.2021.1885257>.
194. W. Yang, G. Feng, D. Miles, et al., "Impact of Biochar on Greenhouse Gas Emissions and Soil Carbon Sequestration in Corn Grown Under Drip Irrigation With Mulching," *Science of the Total Environment* 729 (2020): 138752, <https://doi.org/10.1016/j.scitotenv.2020.138752>.
195. L. Qi, Z. Ma, S. X. Chang, et al., "Biochar Decreases Methanogenic Archaea Abundance and Methane Emissions in a Flooded Paddy Soil," *Science of the Total Environment* 752 (2021): 141958, <https://doi.org/10.1016/j.scitotenv.2020.141958>.
196. R. Conrad, "Microbial Ecology of Methanogens and Methanotrophs," *Advances in Agronomy*, 96 (Academic Press, 2007), 1–63.
197. Y. Feng, Y. Xu, Y. Yu, Z. Xie, and X. Lin, "Mechanisms of Biochar Decreasing Methane Emission From Chinese Paddy Soils," *Soil Biology and Biochemistry* 46 (2012): 80–88, <https://doi.org/10.1016/j.soilbio.2011.11.016>.
198. X. Qin, Y. E. Li, H. Wang, et al., "Long-Term Effect of Biochar Application on Yield-Scaled Greenhouse Gas Emissions in a Rice Paddy Cropping System: A Four-Year Case Study in South China," *Science of the Total Environment* 569 (2016): 1390–1401, <https://doi.org/10.1016/j.scitotenv.2016.06.222>.
199. M. Waqas, A. S. Nizami, A. S. Aburiazza, M. A. Barakat, I. M. I. Ismail, and M. I. Rashid, "Optimization of Food Waste Compost With the Use of Biochar," *Journal of Environmental Management* 216 (2018): 70–81, <https://doi.org/10.1016/j.jenvman.2017.06.015>.
200. L. Chen, M. Liu, A. Ali, et al., "Effects of Biochar on Paddy Soil Fertility Under Different Water Management Modes," *Journal of Soil Science and Plant Nutrition* 20, no. 4 (2020): 1810–1818, <https://doi.org/10.1007/s42729-020-00252-8>.
201. Y. Xiang, Q. Deng, H. Duan, and Y. Guo, "Effects of Biochar Application on Root Traits: A Meta-Analysis," *GCB Bioenergy* 9, no. 10 (2017): 1563–1572, <https://doi.org/10.1111/gcbb.12449>.
202. D. Dong, M. Yang, C. Wang, et al., "Responses of Methane Emissions and Rice Yield to Applications of Biochar and Straw in a Paddy Field," *Journal of Soils and Sediments* 13, no. 8 (2013): 1450–1460, <https://doi.org/10.1007/s11368-013-0732-0>.
203. K. E. Ma, Q. Qiu, and Y. Lu, "Microbial Mechanism for Rice Variety Control on Methane Emission From Rice Field Soil," *Global Change Biology* 16, no. 11 (2010): 3085–3095, <https://doi.org/10.1111/j.1365-2486.2009.02145.x>.
204. J. Kim, G. Yoo, D. Kim, W. Ding, and H. Kang, "Combined Application of Biochar and Slow-Release Fertilizer Reduces Methane Emission But Enhances Rice Yield by Different Mechanisms," *Applied Soil Ecology* 117 (2017): 57–62, <https://doi.org/10.1016/j.apsoil.2017.05.006>.
205. M. Wu, X. Han, T. Zhong, M. Yuan, and W. Wu, "Soil Organic Carbon Content Affects the Stability of Biochar in Paddy Soil," *Agriculture, Ecosystems & Environment* 223 (2016): 59–66, <https://doi.org/10.1016/j.agee.2016.02.033>.
206. X. Zhang, J. Xia, J. Pu, et al., "Biochar-Mediated Anaerobic Oxidation of Methane," *Environmental Science and Technology* 53, no. 12 (2019): 6660–6668, <https://doi.org/10.1021/acs.est.9b01345>.
207. F. S. Segarra, V. Samarkin, M. Y. Yoshinaga, and K. U. Hinrichs, "Joye SB High Rates of Anaerobic Methane Oxidation in Freshwater Wetlands Reduce Potential Atmospheric Methane Emissions," *Nature Communications* 6 (2015): 7477.
208. K. Weber and P. Quicker, "Properties of Biochar," *Fuel* 217 (2018): 240–261, <https://doi.org/10.1016/j.fuel.2017.12.054>.
209. Y. Wang, R. Yin, and R. Liu, "Characterization of Biochar From Fast Pyrolysis and Its Effect on Chemical Properties of the Tea Garden Soil," *Journal of Analytical and Applied Pyrolysis* 110 (2014): 375–381, <https://doi.org/10.1016/j.jaap.2014.10.006>.
210. J. F. Zheng, J. H. Chen, G. X. Pan, et al., "Biochar Decreased Microbial Metabolic Quotient and Shifted Community Composition 4 Years After a Single Incorporation in a Slightly Acid Rice Paddy From Southwest China," *Science of the Total Environment* 571 (2016): 206–217, <https://doi.org/10.1016/j.scitotenv.2016.07.135>.
211. R. I. Solomon, "Biochar Amendments for Reducing Nitrate Leaching From Soils of Different Textural Classes in the Nigerian Savanna," *Turkish Journal of Agriculture-Food Science and Technology* 10, no. 8 (2022): 1363–1368, <https://doi.org/10.24925/turjaf.v10i8.1363-1368.4855>.
212. R. Lv, Y. Wang, X. Yang, et al., "Adsorption and Leaching Characteristics of Ammonium and Nitrate From Paddy Soil as Affected by Biochar Amendment," *Plant Soil and Environment* 67, no. 1 (2021): 8–17, <https://doi.org/10.17221/276/2020-pse>.
213. E. Kabir, K. H. Kim, and E. E. Kwon, "Biochar as a Tool for the Improvement of Soil and Environment," *Frontiers of Environmental Science* 11 (2023): 1324533, <https://doi.org/10.3389/fenvs.2023.1324533>.
214. J. A. Antonangelo, X. Sun, and H. J. Eufraide-Junior, "Biochar Impact on Soil Health and Tree-Based Crops: A Review," *Biochar* 7, no. 1 (2025): 51, <https://doi.org/10.1007/s42773-025-00450-6>.
215. G. B. Melas, O. Ortiz, A. M. Roshdy, M. Y. Hendawi, D. Triantakoustantis, and S. Shaddad, "Assessing the Modulatory Effects of Biochar on Soil Health Status in Response to Pesticide Application," *Earth* 6, no. 2 (2025): 27, <https://doi.org/10.3390/earth6020027>.
216. G. Wang, G. Geng, L. Xu, et al., "Rice Husk Biochar Resuscitates the Microecological Functions of Heavy-Metal Contaminated Soil After Washing by Enriching Functional Bacteria," *Journal of Hazardous Materials* 480 (2024): 136430, <https://doi.org/10.1016/j.jhazmat.2024.136430>.
217. M. Afzal, X. Tan, Y. Ouyang, et al., "Bacterial-Charged Biochar Enhances Plant Growth and Mitigates Microplastic Toxicity by Altering Microbial Communities and Soil Metabolism," *Plant Stress* 17 (2025): 100916, <https://doi.org/10.1016/j.stress.2025.100916>.
218. S. Bolan, D. Hou, L. Wang, et al., "The Potential of Biochar as a Microbial Carrier for Agricultural and Environmental Applications," *Science of the Total Environment* 886 (2023): 163968, <https://doi.org/10.1016/j.scitotenv.2023.163968>.

219. B. Bhatt, S. K. Gupta, S. Mukherjee, and R. Kumar, "A Comprehensive Review on Biochar Against Plant Pathogens: Current State-of-the-Art and Future Research Perspectives," *Heliyon* 10, no. 17 (2024): e37204, <https://doi.org/10.1016/j.heliyon.2024.e37204>.
220. F. Conen, K. E. Dobbie, and K. A. Smith, "Predicting N₂O Emissions From Agricultural Land Through Related Soil Parameters," *Global Change Biology* 6, no. 4 (2000): 417–426, <https://doi.org/10.1046/j.1365-2486.2000.00319.x>.
221. Q. Deng, D. Hui, J. Wang, et al., "Corn Yield and Soil Nitrous Oxide Emission Under Different Fertilizer and Soil Management: A Three-Year Field Experiment in Middle Tennessee," *PLoS One* 10, no. 4 (2015): e0125406, <https://doi.org/10.1371/journal.pone.0125406>.